Original article

Epileptic Disord 2018; 20 (5): 401-12

West syndrome, developmental and epileptic encephalopathy, and severe CNS disorder associated with WWOX mutations

Qudsia Shaukat¹, Jozef Hertecant², Ayman W. El-Hattab², Bassam R. Ali³, Jehan Suleiman^{1,4}

¹ Division of Neurology, Department of Pediatrics, Tawam Hospital, Al Ain

² Division of Genetic and Metabolic Disorders, Departments of Pediatrics, Tawam Hospital, Al Ain

³ Departments of Pathology, College of Medicine and Health Sciences, UAE University, Al Ain

⁴ Department of Pediatrics, College of Medicine and Health Sciences UAE University, Al Ain, United Arab Emirates

Received October 02, 2017; Accepted July 22, 2018

ABSTRACT - Aims. Mutations in the WWOX gene have been reported in a number of patients with various neurological disorders including spino-cerebellar ataxia, intellectual disability, epilepsy, and epileptic encephalopathy. We aimed to study the clinical, electrographic, and imaging features of two new cases with WWOX mutations and compare them to previously reported cases with WWOX mutations. Methods. We assessed two unrelated children from two consanguineous families who had severe neurological disorder including early-onset spastic quadriplegia, profound developmental delay, epilepsy, and West syndrome. Results. Based on whole-exome sequencing, we identified homozygous null mutations in WWOX in both children, and further addressed the genotype-phenotype correlation. In addition, we provide a detailed review of the previously reported cases of WWOX-related neurological disorders and compare them to the children in this report. Conclusions. The findings in this report expand the clinical phenotype associated with WWOX mutations and confirm a well characterised severe central nervous system disorder in association with biallelic null mutations in WWOX. This syndrome consists of profound psychomotor delay, early-onset spastic quadriplegia, and refractory epilepsy including epileptic encephalopathy, acquired microcephaly, and growth restriction. This can be associated with progressive brain atrophy, suggestive of neurodegeneration. Identification of this phenotype by clinicians may help with early diagnosis and appropriate genetic counselling.

Correspondence:

Jehan Suleiman Department of Pediatric, Division of Neurology, Tawam Hospital, P.O. Box 18258, Al-Ain, United Arab Emirates <jsuleiman10@gmail.com> <jsuleiman@seha.ae>

Key words: *WWOX*, West syndrome, epileptic encephalopathy, intellectual disability, microcephaly, spasticity

Central nervous system (CNS) diseases in children including those with severe epilepsy and motor disorders impose big challenges for diagnosis and potential treatments. Many children with undiagnosed CNS disorders are labelled with "cerebral palsy", particularly those with spasticity and seizures as the main features of their disorder. With this label, the search for an underlying diagnosis is often not pursued further (Gupta and Appleton, 2001; Leach et al., 2014). The advent of next-generation sequencing has led to a molecular revolution with rapid growth in the number of identified monogenic determinants underlying many neurological disorders, including severe epilepsy and epileptic encephalopathy (Charng et al., 2016; Fogel et al., 2016; McTague et al., 2016). Epileptic encephalopathies form a large heterogeneous group of severe childhood-onset epilepsies, associated with frequent epileptiform activity on EEG and developmental delay (Berg et al., 2010). West Syndrome (WS) is one of the common epileptic encephalopathies, and is characterised by epileptic spasms along with "hypsarrhythmia", as a specific EEG pattern. Until recently, up to 30% of cases of WS had no identified cause (Osborne et al., 2010). However, many genes have been identified recently in association with WS, therefore an aetiology is now known in a larger proportion of cases. One of the genes described recently in association with CNS disorders and epileptic encephalopathy is WWOX which encodes for a WW domain-containing oxidoreductase; a cytoplasmic protein involved in many cellular processes including growth, differentiation, and tumour suppression (Bednarek et al., 2000; Chang et al., 2014). WWOX is located on a common fragile site, FRA16D, on the long (g) arm of chromosome 16 at position 23 (16q23) and contains nine exons. Somatic WWOX mutations have been reported in different human cancers and its role as a tumour suppressor gene is well recognized (Bednarek et al., 2001). More recently, germline mutations were described in children with neurological disorders, which indicates that WWOX plays an important role in CNS function and development (Abdel-Salam et al., 2014; Mallaret et al., 2014; Ben-Salem et al., 2015; Mignot et al., 2015; Tabarki et al., 2015a; Elsaadany et al., 2016). It is thought that WWOX is highly expressed in various parts of the CNS including the cerebrum, cerebellum, brain stem, and spinal cord. A portion of WWOX is thought to be located in the mitochondria, to carry out its oxidoreductase function (Chang et al., 2014).

Here, we report two children with a severe CNS disorder and epileptic encephalopathy in the form of West syndrome, in association with homozygous null mutations in *WWOX*.

Subjects and methods

Two unrelated children with early-onset spastic quadriplegia, psychomotor retardation, and WS were evaluated at the paediatric neurology service at Tawam Hospital, Al-Ain, United Arab Emirates. Written informed consent was obtained from both families. This study was approved by Al-Ain Medical Human Research Ethics Committee according to the national regulations (protocol number 13/95-CRD 297/13), and funded by the United Arab Emirates University (grant number 31M135).

DNA samples from peripheral blood were extracted for the affected children and their parents at the hospital laboratory, as recommended by the manufacturer. Chromosomal microarray analysis (CMA) and wholeexome sequencing (WES) was performed as a service at Baylor Genetics Laboratory (www.bmgl.com), Texas, USA, according to the generally accepted international standards.

Results

Clinical data

The first child was a two-year-old boy who was born vaginally at term. Birth weight was 3.5 kg (SD: -0.23), length 48 cm (SD: -1.07), and head circumference (HC) 35 cm (SD: -0.63). There were concerns about clenched fists and limb stiffness soon after birth. Head ultrasound showed periventricular cysts, which led to suspicion of periventricular leukomalacia. At five weeks of age, parents reported paucity of spontaneous movements and the presence of sudden brief body jerks. EEG performed at six weeks of age was reported as normal. The child had poor feeding and nasogastric feeds were commenced. At the age of six months, his growth parameters were as follows: weight 5.1 kg (SD: -3.95), length 61.6 cm (SD: -2.86), and HC 39.7 cm (SD: -3.38), consistent with failure to thrive and acquired microcephaly (figure 1A). He had severe global developmental delay, no visual fixation or vocalisation, axial hypotonia, limb spasticity and rigidity (with variable degree), and minimal spontaneous movements (hypokinesia). Optic atrophy (bilateral) was confirmed by an ophthalmologist. At the age of seven months, he continued to have severe psychomotor delay and his parents reported arm stiffening episodes with facial redness, which they noted a few months earlier. EEG showed hypsarrhythmia and epileptic spasms and tonic seizures were captured during the EEG (figure 2A). Oral prednisolone was given as per UKISS protocol (Lux et al., 2005) with a partial response.

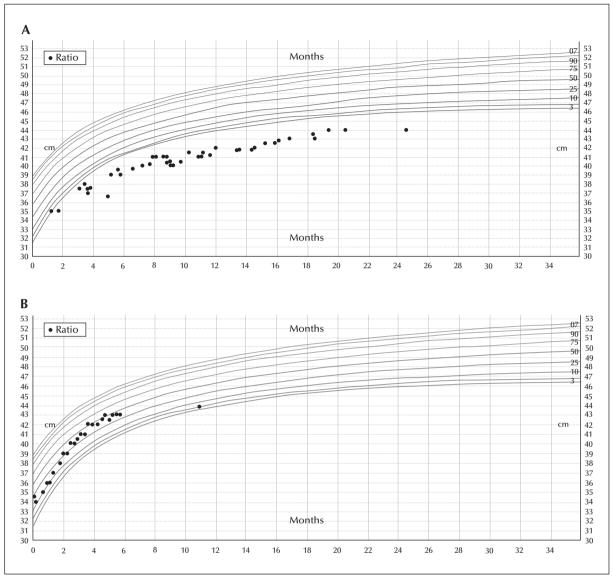


Figure 1. Head circumference growth charts for the first child (A) and second child (B), showing acquired microcephaly (2000 CDC [Centers for Disease Control and Prevention] growth charts).

Vigabatrin was added and the spasms resolved, however, he soon developed other seizure types, including focal myoclonic jerks and clonic seizures, which improved after adding levetiracetam. At two years of age, he remained profoundly delayed with infrequent mixed-type seizures and spastic quadriplegia. EEG showed slow background and infrequent multifocal epileptic activity.

Family history was notable for parents being first cousins with four healthy children. Two cousins had neurological diseases with spasticity and profound developmental delay. They eventually required ventilation and one of them died at the age of four years. The following investigations were normal: lactate, ammonia, acylcarnitine, uric acid, homocysteine, plasma amino acids, urine organic acids, and transferrin isoforms. Cranial magnetic resonance imaging (MRI) at two months of age showed thin corpus callosum, wide Sylvian fissure, and periventricular connatal cysts in the left frontal region (Tan *et al.*, 2010) (*figure 3A*).

The second child was a 12-month-old boy who was born via emergency Caesarean section due to foetal distress and meconium stained fluid at 41 weeks of gestation. He required initial resuscitation and subsequent intubation and ventilation. His birth weight

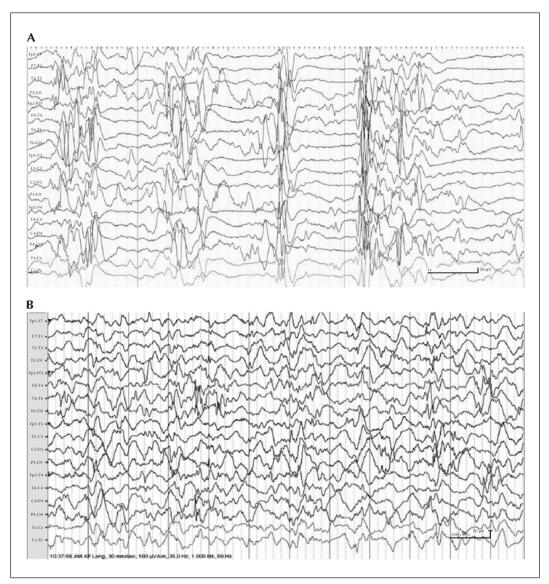


Figure 2. (A) EEG of the first child at age eight months shows high-voltage disorganized background, intermixed with multifocal highamplitude spike and polyspikes, followed by voltage background consistent with hypsarrhythmia and burst suppression. (B) EEG of the second child at age four months showing high-voltage disorganized slow background, intermixed with multifocal high-amplitude spike and polyspikes, consistent with modified hypsarrhythmia.

was 3.745 kg (SD: 0.21), length 50 cm (SD: -0.31), and HC 34.5 cm (SD: -0.87). He was found to have limb hypertonia early in life. Seizures were noticed at five weeks of age and were focal involving lip smacking and arm jerking. EEG showed low-amplitude background of mixed delta and theta activity and frequent sharp and slow waves in the right fronto-temporal region. Phenobarbital was given and seizures improved. At four months, the child failed to attain any developmental milestones. He developed flexor spasms and EEG showed modified hypsarrhythmia in sleep (*figure 2B*) and epileptic spasms were captured. Vigabatrin was added with a partial response. He had mild spasticity in all limbs and was commenced on oral baclofen. He required tracheostomy at four months due to repeated failure of extubation and the need for assisted ventilation.

The child was evaluated again at the age of 11 months. He had profound psychomotor delay and remained ventilator-dependent. He had subtle right hemi spasms occurring in clusters despite being on four antiepileptic medications including vigabatrin, sodium valproate, clobazam, and phenobarbital. Clinical assessment revealed no visual fixation or

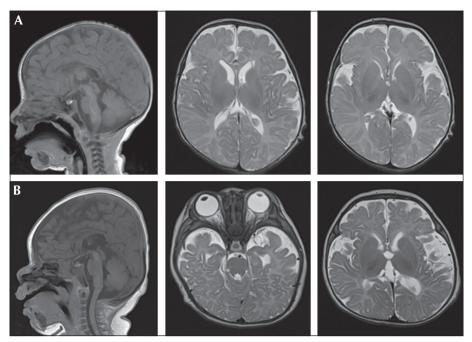


Figure 3. Cranial MRI of the two patients (left: sagittal T1; middle and right: axial T2). (A) The first child at age two months showing hypoplastic corpus callosum, particularly in the anterior portion, wide Sylvian fissure, and small connatal cysts adjacent to the frontal horn of left lateral ventricle. (B) The second child at age two months showing thin corpus callosum, mild ventricular dilatation, and wide CSF spaces, particularly in frontal and temporal regions (indicating reduced cerebral volume) and bilateral posterior polymicrogyria.

communication, axial hypotonia, and limb hypertonia and hyperreflexia. Growth parameters were as follows: weight 7.4 kg (SD: -2.79), length 68 cm (SD: -2.44) and HC 44 cm (SD: -1.62), consistent with failure to thrive and acquired microcephaly (*figure 1A*).

The child was the first born to first-cousin parents, with unremarkable family history.

The following investigations were normal: echocardiogram, lactate, ammonia, acylcarnitine profile, uric acid, homocysteine, plasma amino acids, urine organic acid, and urine sulfocystine. Cranial MRI at two months of age revealed bilateral parietal polymicrogyria, dysplastic thin corpus callosum, wide CSF spaces and reduced white matter volume, particularly in the temporal lobe (*figure 3B*).

Genetic testing

WES in the first child revealed a novel homozygous deletion affecting exons 3 to 4 of the *WWOX* gene (NM_016373.3), at position chr16;78143675-78149052. This finding was confirmed by CMA, which revealed homozygous copy number loss within chromosome band 16q23.1 of approximately 11 Kb in size, including exon 3 and 4 of *WWOX*, consistent with the WES findings. Parental CMA revealed a heterozygous copy number loss involving *WWOX* in both the mother and

the father. No other relevant variants or copy number variations were detected by WES or CMA.

For the second child, CMA did not identify any copy number changes associated with known microdeletion or microduplication syndromes. However, WES revealed a previously reported homozygous pathogenic splice site mutation, c.606-1G>A, in *WWOX* (NM_016373.3) at position chr16;78453766, intron 6 (Tabarki *et al.*, 2015a). This was confirmed by Sanger sequencing. Both parents were heterozygous for this mutation. No other relevant variants were detected by WES.

Discussion

The genetics of epileptic and neurodevelopmental encephalopathies is a rapidly growing field and the recent advances in next-generation sequencing have helped in identifying many monogenic variants in association with epileptic encephalopathy, with autosomal recessive, autosomal dominant or X-linked inheritance (McTague *et al.*, 2016).

Recent reports have revealed the importance of *WWOX* in children with neurological disorders including epilepsy. Here, we report two new cases with *WWOX*-related CNS disorder and present a

summary of all reported patients from previous studies (*table 1*).

The two children presented in this report had earlyonset epilepsy (myoclonic seizures in the first child and focal motor seizures in the second child) and epileptic encephalopathy in the form of WS along with severe neurological disorder in the form of progressive microcephaly, early-onset spasticity, and severe psychomotor retardation. The developmental outcome was unfavourable with profound impairment despite improvement of epileptic activity (e.g. in the first child). This supports the new concept of "developmental and epileptic encephalopathy" in genetic encephalopathies (Scheffer et al., 2017), as compared to the traditional concept regarding epileptic encephalopathy in which it was thought that "the epileptic activity itself contributes to cognitive and behavioural impairments beyond that expected from the underlying pathology alone (e.g. cortical malformation)" (Berg et al., 2010). We believe that, in these children, the underlying cause itself (mutations in WWOX) caused both the severe cognitive impairment and the epileptic encephalopathy. In support of this hypothesis is the fact that the cognitive and psychomotor impairment preceded the onset of epileptic encephalopathy and did not improve with achievement of better control of epileptic activity in some of these cases.

Biallelic missense mutations in *WWOX* were reported in association with a form of childhood-onset cerebellar ataxia with epilepsy and intellectual disability and, in addition, lower limb spasticity was present in some of these patients (Gribaa *et al.*, 2007; Mallaret *et al.*, 2014). This phenotype is milder than that of the children described in this report. This is likely due to the nature of the mutations that were missense rather than null, which result in partial rather than complete loss of function of the protein.

Reviewing the previous reports of children with various biallelic null mutations (nonsense, splice site and deletions) in WWOX shows that many of these children had a clinically distinct form of severe CNS disorder consisting of profound global developmental delay, spastic quadriplegia of early onset, refractory epilepsy, progressive microcephaly, and growth restriction (table 1) (Abdel-Salam et al., 2014; Ben-Salem et al., 2015; Mignot et al., 2015; Tabarki et al., 2015a; Valduga et al., 2015; Elsaadany et al., 2016). Other features in some cases include ophthalmic involvement such as optic atrophy (present in Case 1 in this report), retinal disease (Abdel-Salam et al., 2014; Ben-Salem et al., 2015; Mignot et al., 2015; Tabarki et al., 2015a; Elsaadany et al., 2016), and early death (Mignot et al., 2015; Tabarki et al., 2015a; Valduga et al., 2015). Many children in these reports had early-onset epileptic encephalopathy, most commonly West syndrome (*table 1*).

Of note is that some children had compound heterozygous mutations in the form of deletion and missense, for example, the two siblings in the study of Mignot et al. (Case 3 and 4) (table 1). These children had profound developmental delay as well as severe epilepsy and epileptic encephalopathy, however, they did not have spastic quadriplegia microcephaly or abnormality on cranial MRI. This could be considered an intermediate phenotype which supports genotype-phenotype correlation in cases with WWOX mutations. Interestingly, all reported patients with WWOX mutations so far, whether biallelic null, biallelic missense or compound null and missense mutations, had severe developmental delay/intellectual disability (ID) and epilepsy (23/23 for both features) (table 1), thus potentially adding WWOX to the growing and heterogeneous list of genes associated with ID, as well as epilepsy.

Most children with WWOX biallelic null mutations in previous reports, as well the cases presented here, had loss of volume in supratentorial structures including thinning of the corpus callosum, progressive brain atrophy involving grey and white matter, particularly in frontal and temporal regions, as well as widening of sylvian fissures evident on brain imaging (Abdel-Salam et al., 2014; Mignot et al., 2015; Tabarki et al., 2015a; Elsaadany et al., 2016). WWOX is known to be involved in control of cell survival or death through different mechanisms and interactions with various molecules involved in the regulation of cell signalling, gene transcription, and lipid metabolism (Chang et al., 2014). It is postulated that WWOX plays a critical role in neuronal development, differentiation, and protection, and that loss of WWOX function leads to neuronal injury and neurodegeneration by mechanisms that are still unclear, but possibly involve mitochondrial dysfunction and apoptosis (Chang et al., 2014; Tabarki et al., 2015b), hence leading to brain atrophy and loss of volume.

The rare finding of polymicrogyria in Case 2 and a previously reported case (Ben-Salem *et al.*, 2015) indicates that *WWOX* may play a role in neuronal migration, and more cases and research are needed to confirm the role of *WWOX* as a gene implicated in malformation of cortical development (Parrini *et al.*, 2016).

The findings in this report support a clinically recognisable form of CNS disease in association with *WWOX* biallelic null mutations with the following features: severe early-onset disease with profound psychomotor delay, spastic quadriplegia, refractory seizures including epileptic encephalopathy, particularly WS, progressive microcephaly, and growth restriction. Other common features include poor vision and optic atrophy, variable degrees of rigidity and hypokinesia, Table 1. Clinical, electrographic, imaging and genetic features in patients with mutations in WWOX.

| Study | | Griba | Gribaa e <i>t al.</i> , 2007, Mallaret e <i>t al.</i> , 2014 | , Mallaret | et al., 2014 | _ | Abdel- Salam et al., 2014 | Ben Salem et al. 2014 | | M | Mignot e <i>t al.</i> , 2015 | 2015 | |
|----------------------------------|---------|---|--|-------------|--------------|---------------------|--------------------------------------|-----------------------------|--|---|------------------------------|------------------------------|-------------------------|
| Families/number of | | | | 2/6 | | | 1/1 | 1/1 | | | 4/5 | | |
| Case | - | 2 | 3 | 4 | 5 | 9 | - | - | - | 2 | e S | 4 | 5 |
| Age ^{&} /sex | 19y/F | 18y/F | 16y/M | 10y/F | NA/M | NA/F | 12m/F | 5m/M | 4y/F | 6m/F | 24m/M | 24/F | 10m/F |
| Severe developmental delay/ID | + | + | + | + | + | + | + | + | + | + | + | + | + |
| Spastic quadriplegia (SQ) | | | | | | | + | + | | + | | | + |
| Early onset SQ <3 m | na | na | па | na | na | na | NA | NA | na | AN | na | na | NA |
| Microcephaly | ٩N | AN | ΝA | AN | ν | ΝA | + | + | Possible | + | | | |
| Growth restriction | ٩N | NA | AN | AN | NA | ΝA | + | + | NA | AA | NA | AA | NA |
| Early death <3y | | | 1 | | | | + | na [@] | | + | | | + |
| Ataxia | + | + | + | + | + | + | | | | | | | |
| Ophthalmic involvement | +Nystag | mus +Nystagı | +Nystagmus +Nystagmus +Nystagmus +Nystagmus | ıs +Nystagn | - sn | | +Optic atrophy Abnormal ERG | +Optic atrophy | +Poor eye contact | +Abnormal ERG | , | | +Abnormal ERG |
| Other clinical features | s | Delayed walking a Dysarthria Hyporeflexia | und talkin | ۵۵ | Spastic lc | Spastic lower limbs | | SNHL- unilateral | Hypokin- esia | Rigidity Dystonia Hypokinesia Apnea | ia. | | Rigidity Hypokinesia |
| Epilepsy | + | + | + | + | + | + | + | + | + | + | + | + | + |
| Early onset <3m | | | | ı | | ı | + | + | + | + | | | + |
| Seizure type | GTC | GTC | GTC | GTC | GTC | GTC | Hemicon- vulsions Myoclonic | Focal tonic IS | Focal tonic | Focal and gener- alised | Focal SG Myoclonic | Focal SG | IS Tonic |
| EEC | z | Generali SSW | Generalised Occipital SSW paroxysms | z | NA | A | Generalised bursts of SSW | Hyps | Slow disor- ganised BG Occipital SSW | Slow disor- ganised BG Multifocal SSW | - Slow BG Frontal SSW | Slow BG Multifocal SSW | Modified hyps |
| EE | | | | | NA | NA | + | + | + | + | + | + | + |
| EE type | na 1 | na | na | na | na | na | WS c | SN c | AN AN | V N | AN AN | AN AN | AN N |
| Number of AEUS | _ | 7 | _ | 7 | K N | AN | 7 | 7 | N | K N | NA | NA | N A |
| AED response | Partial | Poor | Good | Good | na | na | Partial | Good | Partial | Partial | Partial | Partial | Partial |

| Study | | Gribaa | Gribaa e <i>t al.</i> , 2007, Mallaret e <i>t al.</i> , 2014 | Aallaret e <i>t</i> | <i>al.</i> , 2014 | | Abdel- Salam et al., 2014 | Abdel- Ben Salam Salem et al., 2014 et al. 2014 | | | Mignot | Mignot e <i>t al.</i> , 2015 | |
|-------------------------------------|----------|----------|--|---------------------|-------------------|------------------------------|---------------------------------|---|-------------|---------------|-----------|---------------------------------------|---------------|
| Cranial MRI# | | | + | + | | | + | + | + | + | + | + | + |
| MRI abnormality | na | na | + | + | na | na | + | + | + | + | | | + |
| Age at MRI | na | na | 12 y | 4 y | na | na | NA | NA | 4 y | 6 m | 24 m | 6 and 24 m | 10 m |
| Cerebral atrophy/ reduced volume | na | na | | | na | na | + | + | + | | , | 1 | + |
| hy | na | na | na | na | na | na | Temporal | NA | ΝA | na | na | na | Generalised |
| Thin corpus callosum na | | na | | | na | na | + | | + | + | | | + |
| Sylvian fissure | | na | | | na | na | | AN | | | | | + |
| widening | | | | | | | | | | | | | |
| Other imaging | | | Mild | Mild | | | Simple | Polymicro- | Mild | | | | |
| findings | | | cerebellar | cerebellar | | | gyral | gyria | ventricular | | | | |
| | | | vermis atrophy vermis | / vermis | | | pattern | (right | enlarge- | | | | |
| | | | Posterior | atrophy | | | Hippocamp. | Front- | ment | | | | |
| | | | white | | | | dysplasia | opariatal) | | | | | |
| | | | matter hyper- | | | | | | | | | | |
| | | | intensities | | | | | | | | | | |
| Summary of MRI | Not done | Not done | Mild | Mild | Not done | Not done | Supraten- | Cerebral | Reduced | Mild | Normal | Normal | Progressive |
| abnormalities | | | cerebellar | cerebellar | | | torial | atrophy | MM | myelina- | | | cerebral |
| | | | vermis atrophy vermis | / vermis | | | atrophy | Polvmicro- | volume | tion delav | | | atrophy |
| | | | Posterior | atrophy | | | Simplified | evria | Thin CC | Thin CC | | | Thin CC |
| | | | white | 1 | | | must botton | aural nattorn (right from |) |) | | | Wido |
| | | | matter | | | | gyrar parterr Thin CC | tonariatal) | | | | | svlvian |
| | | | | | | | | (approximation) | | | | | |
| | | | nyperın- tensities | | | | | | | | | | tissures |
| WWOX variant allele 1 | | c.139C | c.139C>A (p.P47T) | | c.1114G> | c.1114G>C (p.G372R) c.160G>T | c.160G>T | Deletion | Deletion | Deletion | c.45_48de | c.45_48delGGAC (p.D16Sfs*63) Deletion |) Deletion |
| | | | | | | | (p.R54*) | exon 5 | exon 1-5 | exon 6 | | | exon 1-9 |
| WWOX variant allele 2 | | c.139C | c.139C>A (p.P47T) | | c.1114G> | c.1114G>C (p.G372R) c.160G>T | c.160G>T | Deletion | Deletion | c.1005G>A | | c.140C>G (p.P47R) | c.889A>T |
| | | | | | | | (p.R54*) | exon 5 | exon 6-8 | (p.W335*) | | | (p.K297*) |
| WWOX mutation | | Hom | Hom missense | | Homr | Hom missense | Hom | Hom | CH: | CH: dele- | CH: fr | CH: frameshift/missense | CH: dele- |
| zveosity | | | | | | | nonsense | deletion | deletions | tion/nonsense | anse | | tion/nonsense |

| Families/number of cases | | | Tabarki et al., 2015a | 15a | | Valduga e <i>t al.</i> , 2015* | | Elsaadany e <i>t al.</i> , 2016 | This | This study | Total ^{\$} |
|----------------------------------|------------------|------------------|-----------------------|------------------|------------------|--------------------------------|-----------------------------|---------------------------------|---------------------------------------|---------------------------------------|---------------------|
| | | | 2/5 | | | 1/1 | | 1/2 | | 2/2 | 13/23 |
| Case | + | 2 | 3 | 4 | 5 | + | 1 | 2 | 1 | 2 | 23 |
| Age ^{&} /sex | 11m/NA | AN | NA | NA | NA | 22m/F | 7y/F | 20m/F | 24m/M | 13m/M | |
| Severe developmental delay/ID | + | + | + | + | + | + | + | + | + | + | 23/23 |
| Spastic quadriplegia (SQ) | + | + | + | + | + | + | + | + | + | + | 14/23 |
| Early onset SQ <3 m | + | + | + | + | + | + | ΝA | + | + | + | 6/6 |
| Microcephaly | + | + | + | + | + | + | | | + | + | 12/17 |
| Growth restriction | + | + | + | + | + | + | + | + | + | + | 12/12 |
| Early death <3y | + | + | + | + | + | + | | na [@] | na [@] | na [@] | 9/19 |
| Ataxia | | | | | | | | | | | 6/23 |
| Ophthalmic involvement | +Abnormal ERG | +Abnormal ERG | +Abnormal ERG | +Abnormal ERG | +Abnormal ERG | +Poor eye contact | +Optic atrophv | +Poor eye contact | +Optic atrophv | +Poor eye contact | 19/23 |
| Other clinical features | ي. | | | | | Cardiomyopathy | Respiratory failure | | Rigidity Hypokinesia | Respiratory failure Hypokinesia | |
| Epilepsy | + | + | + | + | + | + | + | + | + | + | 23/23 |
| Early onset <3m | + | + | + | + | + | + | + | + | + | + | 15/23 |
| Seizure type | Multifocal IS | Multifocal IS | Multifocal IS | Focal SG | Focal SG | Myoclonic IS | Myoclonic | Focal tonic IS | Tonic IS Focal myoclonic | Focal motor IS Hemispasms | |
| B | NA | NA | AA | ¥ Z | NA | Normal /Hyps | Abnormal BG Frequent SSW | Consistent / with EE | Hyps /Slow BG multifocal SSW | Modified hyps | |
| EE | + | + | + | + | + | + | | + | + | + | 16/21 |
| EE type | WS LGS | WS LGS | WS | NA | ٧V | WS | na | ٨٨ | WS | WS | |
| Number of AEDs | AA | AA | AA | NA | AA | 4 | 4 | 3 | 3 | 4 | |
| AED response | Poor | Poor | Poor | Poor | Poor | Partial | Poor | Partial | Partial | Poor | |
| Cranial MRI# | + | + | | | | + | + | + | + | + | 16/23 |

| Study | | Tabarl | Tabarki e <i>t al.,</i> 2015a | | | Valduga e <i>t al</i> ., 2015* | | Elsaadany et al., 2016 | Thi | This study | Total ^{\$} |
|-------------------------------------|--|--|-------------------------------|----------|----------|----------------------------------|---|---|-------------------------------------|--|---------------------|
| MRI abnormality | + | + | na | na | na | + | + | + | + | + | 14/16 |
| Age at MRI | 0 m and 11 m | NA | na | na | na | NA | 2 m and 5 m | NA | 2 m | 2 m | |
| Cerebral atrophy/ reduced volume | + | + | na | na | na | + | + | + | 1 | + | 10/16 |
| Location of atrophy | Posterior | posterior | na | na | na | Sylvian fissure | Fronto temporal | Fronto temporal | na | Fronto temporal | |
| Thin corpus callosum | + | + | na | na | na | + | + | + | + | + | 11/16 |
| Sylvian fissure widening | 1 | + | na | na | па | + | + | NA | + | + | 6/13 |
| Other imaging findings | Connatal cysts Symmetric thalamic lesions | s Symmetric thalamic lesions | | | | | | | Connatal cyst | Connatal cysts Polymicrogyria (bilateral posterior) | ia |
| Summary of MRI abnormalities | Progressive Periventricul atrophy of WM loss periventricular posteriorly WM, CC and Thin CC Flat upper upper brain brainstem stem | Periventricular Not WM loss r posteriorly Thin CC Flat upper brain stem | Not done | Not done | Not done | Thin CC Wide sylvian fissures | Hypomyel- ination Progressive frontotempo- ral atrophy Thin CC | Hypomyel- ination Progressive frontotempo- ral atrophy Thin CC | Thin CC Wide sylvian fissures | Frontotemporal atrophy Thin CC Wide sylvian fissures Polymicrogyria | ia ral |
| WWOX variant allele 1 | _ | | c.606-1G>A | | | Deletion exon 1-6 | c.13 ⁷ (p.V | c.131G >A (p.W44*) | Deletion exons 3- 4 | c.606-1G>A | |
| WWOX variant allele 2 | 2 | | c.606-1G>A | | | Deletion exon 1-6 | c.13 ⁻ (p.V | c.131G >A (p.W44*) | Deletion exons 3- 4 | c.606-1G>A | |
| WWOX mutation zygosity | | | Hom splice-site | e | | Hom deletion | Нот | Hom nonsense | Hom deletion | Hom deletion Hom splice site | ite |

Table 1. Clinical, electrographic, imaging and genetic features in patients with mutations in WWOX (Continued).

troretinogram; SMTE: sensory neural nearing 1985; OTC: generalised contry, 15: manue spasins, 50: secondary generalised, EC: electroencephanogram, 15: normal, 50: spike and slow wave; hyps: hypsarythmia; BG: background; EE: epileptic encephalopathy; WS: West syndrome; LGS: Lennox- Gastaut syndrome; AEDs: antiepileptic drugs; MRI: magnetic resonant imaging: CC: corpus callosum; WM: white matter; Hom: homozygous; Het: heterozygous; CH: compound heterozygous. "The prenatally terminated case in this study was not included in this table as clinical features are mostly not applicable. ⁵The total is calculated based on available data; cases with NA or na for any feature were subtracted from the total. [&]When the age was not specified in these reports, clues were used to determine age, *e.g.* age at MRI study. [@] na- not applicable as children were not three years old at the time this report was written. [#]When the MR images were available in the manuscript of these studies, extra interpretation was performed by JS to include features related to this table that were not described in the manuscript or figure legends in these reports. feeding difficulties, and early death. Reduced volume of cerebral white and grey matter can be seen on cranial imaging with predilection to frontal and temporal areas including widening of sylvian fissures. Although there is overlap of this phenotype with many other neurodegenerative disorders, including those associated with other genes, identifying the features of *WWOX*-related CNS disease in children with earlyonset epilepsy may help early diagnosis of affected children using targeted genetic testing, which may save time and cost.

Our findings support the role of advanced genetic testing as an important tool in the diagnosis of neurological disorders in children, particularly those with severe epilepsy and epileptic encephalopathy. Some children with spastic quadriplegia and epilepsy are labelled with cerebral palsy, particularly those with periventricular cysts or white matter loss, similar to the first child in this report. It is important to identify a potential genetic autosomal recessive disorder in these patients. Accurate and early genetic diagnosis is important and has many implications for the affected families, including specific recurrence risk, counselling for future pregnancies, and the potential for pre-implantation genetic diagnosis.

Supplementary data.

Summary didactic slides are available on the www.epilepticdisorders.com website.

Acknowledgements and disclosures.

This study was approved by Al-Ain Medical Human Research Ethics Committee according to the national regulations (protocol number 13/95-CRD 297/13). Both families consented to this publication. The study was funded by the United Arab Emirates University (grant number 31M135). JS received research awards from the National Health and Medical Research Council (NHMRC), Australia.

None of the authors have any conflict of interests to declare.

References

Abdel-Salam G, Thoenes M, Afifi HH, Korber F, Swan D, Bolz HJ. The supposed tumor suppressor gene *WWOX* is mutated in an early lethal microcephaly syndrome with epilepsy, growth retardation and retinal degeneration. *Orphanet J Rare Dis* 2014; 9: 12.

Bednarek AK, Laflin KJ, Daniel RL, Liao Q, Hawkins KA, Aldaz CM. WWOX, a novel WW domain-containing protein mapping to human chromosome 16q23.3-24.1, a region frequently affected in breast cancer. *Cancer Res* 2000; 60: 2140-5.

Bednarek AK, Keck-Waggoner CL, Daniel RL, *et al. WWOX*, the *FRA16D* gene, behaves as a suppressor of tumor growth. *Cancer Res* 2001; 61: 8068-73.

Ben-Salem S, Al-Shamsi AM, John A, Ali BR, Al-Gazali L. A novel whole exon deletion in *WWOX* gene causes early epilepsy, intellectual disability and optic atrophy. *J Mol Neurosci* 2015; 56: 17-23.

Berg AT, Berkovic SF, Brodie MJ, *et al.* Revised terminology and concepts for organization of seizures and epilepsies: Report of the ILAE Commission on Classification and Terminology, 2005-2009. *Epilepsia* 2010; 51: 676-85.

Chang HT, Liu CC, Chen ST, Yap YV, Chang NS, Sze CI. WW domain-containing oxidoreductase in neuronal injury and neurological diseases. *Oncotarget* 2014; 5: 11792-9.

Charng WL, Karaca E, Coban Akdemir Z, *et al.* Exome sequencing in mostly consanguineous Arab families with neurologic disease provides a high potential molecular diagnosis rate. *BMC Med Genomics* 2016; 9: 42.

Elsaadany L, El-Said M, Ali R, Kamel H, Ben-Omran T. W44X mutation in the *WWOX* gene causes intractable seizures and developmental delay: a case report. *BMC Med Genet* 2016; 17:53.

Fogel BL, Satya-Murti S, Cohen BH. Clinical exome sequencing in neurologic disease. *Neurol Clin Pract* 2016; 6: 164-76.

Gribaa M, Salih M, Anheim M, et al. A new form of childhood onset, autosomal recessive spinocerebellar ataxia and epilepsy is localized at 16q21-q23. *Brain* 2007; 130: 1921-8.

Gupta R, Appleton RE. Cerebral palsy: not always what it seems. *Arch Dis Child* 2001; 85: 356-60.

Leach EL, Shevell M, Bowden K, Stockler-Ipsiroglu S, Van Karnebeek CDM. Treatable inborn errors of metabolism presenting as cerebral palsy mimics: systematic literature review. *Orphanet J Rare Dis* 2014; 9: 197.

Lux AL, Edwards SW, Hancock E, *et al.* The United Kingdom Infantile Spasms Study (UKISS) comparing hormone treatment with vigabatrin on developmental and epilepsy outcomes to age 14 months: a multicentre randomised trial. *Lancet Neurol* 2005; 4:712-7.

Mallaret M, Synofzik M, Lee J, *et al.* The tumour suppressor gene *WWOX* is mutated in autosomal recessive cerebellar ataxia with epilepsy and mental retardation. *Brain* 2014; 137: 411-9.

McTague A, Howell KB, Cross JH, Kurian MA, Scheffer IE. The genetic landscape of the epileptic encephalopathies of infancy and childhood. *Lancet Neurol* 2016; 15: 304-16.

Mignot C, Lambert L, Pasquier L, *et al.* WWOX-related encephalopathies: delineation of the phenotypical spectrum and emerging genotype-phenotype correlation. *J Med Genet* 2015; 52: 61-70.

Osborne JP, Lux AL, Edwards SW, et al. The underlying etiology of infantile spasms (West syndrome): information from the United Kingdom Infantile Spasms Study (UKISS) on contemporary causes and their classification. *Epilepsia* 2010; 51: 2168-74.

Parrini E, Conti V, Dobyns WB, Guerrini R. Genetic basis of brain malformations. *Mol Syndromol* 2016; 7: 220-33.

Scheffer IE, Berkovic S, Capovilla G, *et al.* ILAE classification of the epilepsies: Position paper of the ILAE Commission for Classification and Terminology. *Epilepsia* 2017; 58: 512-21.

Tabarki B, Alhashem A, Alshahwan S, Alkuraya FS, Gedela S, Zuccoli G. Severe CNS involvement in *WWOX* mutations: Description of five new cases. *Am J Med Genet A* 2015a; 167A: 3209-13.

Tabarki B, Al Mutairi F, Al Hashem A. The fragile site WWOX gene and the developing brain. *Exp Biol Med (Maywood)* 2015b; 240: 400-2.

Tan ZY, Naidoo P, Kenning N. Case of the month, Ultrasound and MRI features of connatal cysts: clinicoradiological differentiation from other supratentorial periventricular cystic lesions. *Br J Radiol* 2010; 83: 180-3.

Valduga M, Philippe C, Lambert L, *et al*. WWOX and severe autosomal recessive epileptic encephalopathy: first case in the prenatal period. *J Hum Genet* 2015; 60: 267-71.



(1) What are the most consistent features in all reported cases with different types of WWOX mutations?

(2) List the clinical features associated with biallelic null type (nonsense, splice site, deletion) mutations of *WWOX*.

(3) West syndrome is the most common type of epilepsy associated with biallelic null mutations of *WWOX*-true or false?

Note: Reading the manuscript provides an answer to all questions. Correct answers may be accessed on the website, www.epilepticdisorders.com, under the section "The EpiCentre".