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Heinrich Unverricht
(1853-1912)



Gonzalo Rodriguez Lafora
(1886-1971)

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Progressive Myoclonus Epilepsies: State-of-the-Art

Invited Editors:
**Berge A. Minassian, Pasquale Striano,
Giuliano Avanzini**

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Progressive Myoclonus Epilepsy: The Gene-Empowered Era

The progressive myoclonus epilepsies (PMEs) are “progressive”, because they worsen with time and are generally fatal. They are “myoclonus”, because these patients generally have frequent, often constant, myoclonus, which is commonly, though not invariably, cortical. They are “epilepsies”, because in addition to the myoclonus the patients suffer convulsive and other types of seizures, which are soon intractable, and which in many cases precipitate death in status epilepticus. But the PMEs are in most cases more. They are often associated with blindness, because the retina is nervous tissue, and what is wrong with that nervous tissue is what is wrong with the brain itself, namely neurodegeneration. The PMEs are therefore also ataxia, and dementia, and general demise of the brain. The PMEs are yet more. They are children who are born and grow and go to school and love and are loved, before they are struck. They are therefore not static early tragedies that families adjust to from the get-go, but are children with whom the family has grown and who are slipping daily a little bit more into greater pain, towards greater intellectual and emotional separation, and towards death. And yet that death does not come fast in most cases, because these are children generally with otherwise healthy bodies. They and their families therefore suffer for years, sometimes decades before the end.

Yet again the PMEs are something more. Almost all are monogenic diseases. As such, however horrible, they are genetically simple. As such, and because each has an open window of health prior to significant neurodegeneration, they will be among the first brain diseases to be treated by interventions such as gene replacement therapy. Also, because they are genetically simple, they will be understood ahead of genetically complex neurological disorders.

We are therefore dealing with the worst and best of all neurological worlds. This series of articles reviews the PMEs and provides the most up-to-date knowledge of their basic mechanisms. It concludes with an outlook

at upcoming gene knowledge empowered therapies. It is hoped that the next book written on this subject will include real examples of available cures, and a dream shared by all neurologists, that when they see a family with a PME they would be able to say: “This is what you have. Take this.”

The editors would like to thank Dr. Maria Majno from the Mariani Foundation for her tireless work in helping organize this series, her insights, and her steady gentle prodding to realize this project. At the Mariani Foundation, we would also like to thank Ms. Valeria Basilico for her outstanding productive coordination.

We are very grateful to Pr. Alexis Arzimanoglou at *Epileptic Disorders* for his expert guidance and chief-editing towards ensuring excellent clinical and scientific accessibility of all the manuscripts. The educational missions of both the Mariani Foundation and *Epileptic Disorders* are certainly being fulfilled, and so well.

A word of appreciation is due to all the leaders and teachers in the field of PMEs over the decades. We are certainly forgetting many, but these names include the team from the Montreal Neurological Institute, Drs. Eva and Fred Andermann, Samuel Berkovic now in Australia, previously trained with the Andermanns, and Guy Rouleau; the group in Helsinki led by Anna-Elina Lehesjoki; the Marseille group (the late Dr Joseph Roger and his team: Charlotte Dravet, Pierre Genton and Michelle Bureau); and the Los Angeles group (Antonio Delgado-Escueta and his trainee now a leader in Spain, Jose-Maria Serratosa).

Finally, the greatest thanks go to each and every child we have all seen, who taught us so much about the brain generally, for the sake of countless future patients, and all the families who teach us every day what humanity is at its best.

Berge A. Minassian, Pasquale Striano,
Giuliano Avanzini

TEST YOURSELF



- 1) What is the main difference between a Lafora body of Lafora disease and the amyloid plaque of Alzheimer's disease?
- 2) Is Unverricht-Lundborg disease caused by complete loss of function of the cystatin B protein?
- 3) What is common in the numerous neuronal ceroid lipofuscinoses?
- 4) Do you know a PME which is at the same time a motor neuron disease?
- 5) Do you know a potassium channel PME?
- 6) Which PME is associated with renal failure?
- 7) Which genetic PME is not due to nuclear genome mutations?
- 8) What viral infections could mimic a PME?
- 9) Which PME has a cherry-red spot?
- 10) Which PME is caused by a dodecamer repeat expansion and is not Unverricht-Lundborg disease and is not even a human disease?

Note: Reading the series of articles provides an answer to all questions. Correct answers may be accessed on the website, www.epilepticdisorders.com, under the section "The EpiCentre".

The history of progressive myoclonus epilepsies

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ABSTRACT – The history of the progressive myoclonus epilepsies (PMEs) spans more than a century. However, the recent history of PME begins with a consensus statement published in the wake of the Marseille PME workshop in 1989 (Marseille Consensus Group, 1990). This consensus helped define the various types of PME known at the time and set the agenda for a new era of genetic research which soon led to the discovery of many PME genes. Prior to the Marseille meeting, and before the molecular era, there had been much confusion and controversy. Because investigators had but limited and biased experience with these rare disorders due to the uneven, skewed distribution of PMEs around the world, opinions and nosologies were based on local expertise which did not match well with the experiences of other researchers and clinicians. The three major areas of focus included: (1) the nature and limits of the concept of PME in varying scopes, which was greatly debated; (2) the description of discrete clinical entities by clinicians; and (3) the description of markers (pathological, biological, neurophysiological, etc.) which could lead to a precise diagnosis of a given PME type, with, in the best cases, a reliable correlation with clinical findings. In this article, we shall also examine the breakthroughs achieved in the wake of the 1989 Marseille meeting and recent history in the field, following the identification of several PME genes. As in other domains, the molecular and genetic approach has challenged some established concepts and has led to the description of new PME types. However, as may already be noted, this approach has also confirmed the existence of the major, established types of PME, which can now be considered as true diseases.

Key words: progressive myoclonus epilepsy, Lafora, Unverricht-Lundborg, Kufs

The concept of progressive myoclonus epilepsy

The relationship between "myoclonia" and epilepsy was recognized by Prichard in 1822 (quoted by Rabot [1899]). Delasiauve had also noticed

the existence of myoclonic jerks in patients with epilepsy and in his 1854 treatise on epilepsy, labelled them "*petit mal moteur*". The myoclonic jerks, well described by many authors, were found in patients with various conditions, ranging from a comparatively benign, non-progressive type that would later

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Figure 1. Herman Lundborg (1868-1943).

Herman Lundborg wrote his dissertation in 1903 at the Karolinska Institutet, in Stockholm, about a family with the condition previously described by Unverricht, which he studied from a clinical point of view but also from a genetic perspective. His interest in genetics led him to found the notorious State Institute of Racial Biology, in Uppsala, in 1922. He came under strong criticism and disrepute due to his adherence to Nazi ideology and his advocacy of eugenics and the sterilisation of “genetically unworthy” persons.

be described as “*impulsive petit mal*” (Janz & Christian, 1957), to many more severe examples. Following Friedreich’s “*paramyoclonus multiplex*” (1881), it was admitted that the jerks probably originated in the spinal cord. No clear disease entity was associated with these jerks (Friedreich, 1881).

The concept of “progressive myoclonus epilepsy” (Minassian *et al.*, 2016) was introduced by Herman Lundborg (figure 1) in 1903 (Lundborg, 1903), on the basis of several Swedish families with a common ancestor and (among other markers of “degeneration”) a particular form of epilepsy associated with progressive myoclonus and varying degrees of severity (figure 2). He acknowledged the previous reports from Estonia by Heinrich Unverricht (Unverricht, 1891) (figure 3) who had described two families with “*Myoklonie*” (1891) or “*familiäre*

Myoklonie” (1895). Both authors had aptly described a fairly “pure” type of PME which did not include major symptoms other than the myoclonias and epileptic seizures. It took, however, nearly a century for this condition to be rightly recognised as “Unverricht-Lundborg” disease (ULD). Their contributions were widely read and commented upon, but failed to convince later authors that they had described a recognisable, specific condition. In order to reach a consensus, there were obviously too few cases in the patienthood of major neurologists at the time. When Lafora (figure 4) described the pathological inclusion found in the brain of a patient with a “myoclonic epilepsy”, which he also aptly described, he did not believe that his patient was different from those of Unverricht and Lundborg (Lafora, 1911).

Hunt (figure 5) contributed to the complexity of the matter by describing patients with signs of Friedreich’s ataxia associated with action myoclonus and (in some cases) epilepsy (Hunt, 1921). The “Ramsay Hunt syndrome” (RHS; not to be confused with the description by the same author of the herpes infection of the geniculate ganglion, with resulting facial paresis and skin eruption) covered many clinical conditions, including ULD (Roger *et al.*, 1968). RHS was finally discarded as a useful entity (Andermann *et al.*, 1989a), however, at that time not for the right reasons, but because it was felt that the recent recognition of mitochondrial diseases with progressive myoclonus and seizures had cleared the way.

There were, however, efforts to try and introduce order to the PMEs. Van Bogaert approached the issue from a mixed neuropathological and clinical point of view, and supported the concept of PME, but failed to establish clear boundaries between the various types (Van Bogaert, 1968). In 1973, Diebold defined a nucleus of “hereditary myoclonus-epilepsy-dementia nuclear syndromes” (*erbliche myoklonisch-epileptisch-dementielle Kernsyndrome*), which he differentiated from the “borderline syndromes” occurring in diseases which only fit the PME definition in some cases (Diebold, 1973). Heralding the modern approach, the Montreal group also acknowledged the concept of PME and proposed a classification that was, subjectively, based on the relative frequency of these rare conditions (Berkovic *et al.*, 1986). Before the genetic advances of the past twenty years had really had an impact, the Marseille group (Genton *et al.*, 1990) proposed to divide the PMEs into those with known biochemical mechanisms (e.g. MERRF and sialidosis), those with a definite and reliable pathological marker (e.g. Lafora’s disease, the neuronal ceroid lipofuscinoses [NCLF]), and those without any marker (the “degenerative” types, with purely clinical diagnosis and exclusion of other aetiologies: e.g. ULD and DRPLA).

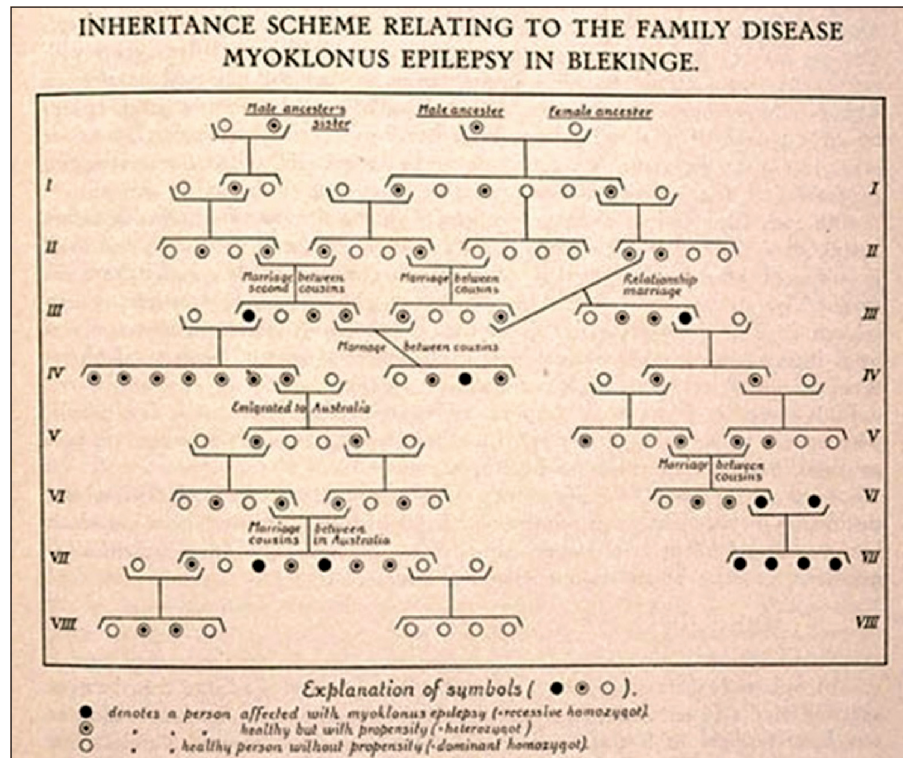


Figure 2. A pedigree showing recessive transmission in a family with Unverricht-Lundborg disease (From Lundborg & Runnstrom [1921]).

Clinical descriptions and pathological markers

Table 1 summarises, for the major PME, the progression from clinical descriptions to molecular elucidation, which is currently nearly complete. However, it appears that the process was fairly uneven. Some descriptions preceded the molecular characterisation of the condition by more than a hundred years, while in other cases, a “new” disease was described on the basis of a singular clinical, pathological or genetic feature.

In the classic sequence of events, a clinical description occurred first, followed by a more or less specific biological or neurophysiological marker which helped ascertain the diagnosis. This was the case for the various forms of NCLF. The juvenile type of NCLF was described by Stengel, a general practitioner in 1826, in a geographic isolate of inland Norway (Stengel, 1826), but it took nearly a century to distinguish this and other forms of NCLF from other forms of “amaurotic idiocy”, which included non-PME disorders such as Tay-Sachs disease. While Batten had not initially distinguished these conditions from one another (Batten, 1902), in 1903 an ophthalmologist, Alfred Bielschowsky, characterised the ocular findings in the late infantile form of NCLF. The more specific pathological, ultra-structural changes associated with the infantile and

juvenile types of NCLF were only described in the 1970s (Zeman *et al.*, 1970). Although it took some time to differentiate NCLF from other types of degenerative childhood diseases, which included mental decline and retinal impairment, they were fairly well distinguished, on clinical grounds, from other types of PMEs. However, another condition with optional ophthalmological symptoms, sialidosis, was only clearly identified in the 1970s (Rapin *et al.*, 1978).

In the case of Lafora’s disease, the pathological marker, the presence of amyloid deposits in the brain, was described by Gonzalo Lafora in 1911, together with a fairly precise clinical depiction of the condition named after him. But it took half a century of controversies before a sound and precise clinical description of Lafora’s disease (LD) was reached in the Netherlands (Van Heycoptenham & De Jager, 1963). From this point onwards, LD was for most, but not all, a clearly identifiable entity. In subsequent years, several refinements were made to the clinical description, focusing on the characteristic EEG presentation and on the occurrence of occipital lobe seizures (Roger *et al.*, 1983; Tinuper *et al.*, 1983).

Diagnosis was much more difficult in the absence of precise markers, when the clinician was left to speculate on patient cases purely on the basis of clinical traits. Some neurophysiological features were shared



Figure 3. Heinrich Unverricht (1853-1912).

Bust erected in 1914 at Magdeburg University. During his short tenure at Dorpat (now Tartu, Estonia), which he left because of the Russification policies of the occupying forces, Heinrich Unverricht described a family with “*Myoclonie*”, i.e. with the condition now named after him, “Unverricht-Lundborg disease”. He was a prolific internist who also described other conditions (polymyositis and pneumonia). His contribution is regarded as the founding description of progressive myoclonus epilepsy.

by several, clearly different conditions. As an example, the spectacular occurrence of runs of polyspikes during REM sleep which was described in several “myoclonic” and progressive conditions, such as Ramsay-Hunt syndrome (soon to become Unverricht-Lundborg disease [ULD]), was also seen in post-anoxic myoclonus, or MERRF. Indeed, based on their own experience, various authors promoted a regional type of PME, which dominated local experiences. In Finland, close to the original sites of Unverricht’s and Lundborg’s descriptions, the “Baltic myoclonus” was the prototype of PMEs (Koskiniemi *et al.*, 1974; Koskiniemi, 1986). Likewise, RHS was repeatedly described in Marseille following Roger *et al.* (1968) and was, in the light of the Finnish publications, labelled “Mediterranean myoclonus” and considered to constitute a milder entity than the “Baltic” type and MERRF (Genton *et al.*, 1990). An explanation had already been given for the difference in severity; in Northern Europe, phenytoin, the most prescribed anticonvulsant for epilepsies with convulsive seizures (including myoclonic seizures), had clearly contributed to an artificial aggravation of the condition (Elridge *et al.*, 1983), in contrast to Mediterranean patients, who were more likely to be

treated (or over-treated) with phenobarbital, which lacked this aggravating effect.

In the 1980s, convincing descriptions of new entities emerged, such as mitochondrial encephalopathy with ragged-red fibres (MERRF) (Fukuhara *et al.*, 1980), and dentato-rubro-pallido-luysian atrophy (DRPLA) (Naito & Oyanagi, 1982), and it was tempting to ascribe previously unresolved cases to these new findings, thus rendering the RHS concept obsolete (Berkovic *et al.*, 1986; Andermann *et al.*, 1989a). The time had come to compare the experience of researchers from Europe, America and Japan; an international workshop was organised in Marseille in June 1989, which heralded the modern, genetic and molecular era in PME research.

The genetic era

Prior to 1989, the year of the Marseille conference, it had only been possible to identify the gene for only one autosomal recessive PME (NEU1; sialidosis), using classic biochemical methods (Rapin *et al.*, 1978). The Marseille conference coincided with momentous developments in the history of genetics. 1989 was the year when the promise of reverse genetics,

Table 1. Discovery and description of the main classic PME types, showing the timescale, from clinical description to diagnostic marker and genetic localisation and elucidation, in chronological order of initial clinical descriptions. In some cases, such as Lafora's disease, the discovery of a pathological marker preceded the comprehensive clinical description by many years.

| PME type | First description (year, author) | Pre-genetic diagnostic marker (year) | Locus/gene (year) |
|--|--|---|----------------------|
| Juvenile NCLF | 1826 Stengel, Norway 1908 Spielmeyer, Germany 1931 Sjögren, Sweden | Finger print profiles (1963) | 1989 |
| Unverricht-Lundborg disease | 1891 Unverricht, Estonia 1905 Lundborg, Sweden | None | 1991 |
| Lafora's disease | 1911 Lafora Spain/USA 1963 Van Heycop Ten Ham, Netherlands | "Myoclonic corpuscles" (1911) | 1995 |
| Late-infantile NCLF | 1913 Bielschowsky, Germany | Curvilinear profiles (1963) | 1997 |
| Adult NCLF | 1925 Kufs, Germany | Various | 2011 |
| Sialidosis | 1978 Rapin, USA | Enzyme defect (1978) | 1996 |
| MERRF | 1980 Fukuhara, Japan | Ragged red fibres in muscle (1980) | 1990 |
| DRPLA | 1982 Naito and Oyanagi, Japan | None | 1995 |
| Action myoclonus-renal failure syndrome | 1986 Andermann, Canada | None | 2008 |

NCLF: neuronal ceroidlipofuscinosis.

identifying a disease gene by first mapping its chromosomal location, was first fulfilled with the discovery of the cystic fibrosis gene (*CFTR*) (Rommens *et al.*, 1989). While *CFTR* was mapped using restriction length polymorphisms, that same year the discovery of microsatellite polymorphisms was also first reported (Weber & May, 1989). The microsatellite maps that rapidly followed had just the right density for homozygosity and linkage mapping of autosomal recessive Mendelian diseases, and since the vast majority of PMEs are inherited in this fashion, their genes quickly began to be identified in the years that followed.

PME gene discoveries proceeded in the approximate order in which the diseases themselves had been described, which is likely to be a reflection of the relative frequencies of the various diseases. The *CLN1* (Infantile NCL) and *CLN3* (Batten's disease) genes were identified in 1995 (The International Batten Disease Consortium, 1995; Vesa *et al.*, 1995), the ULD gene in 1996 and 1997 (Pennacchio *et al.*, 1996; Lafrenière *et al.*, 1997; Lalioti *et al.*, 1997; Virtaneva *et al.*, 1997), and the LD genes between 1998 and 2003 (Minassian *et al.*, 1998; Serratosa *et al.*, 1999; Chan *et al.*, 2003). The most "myoclonic" of the NCL genes, *CLN2*, was cloned not through reverse genetics, but by using an elegant

biochemical approach, taking advantage of the realisation that most NCL are lysosomal diseases. The authors isolated lysosomal proteins and looked for a missing spot in two-dimensional gels in patients with late-infantile NCL, in order to identify *CLN2*, a lysosomal dipeptidyl peptidase (Sleat *et al.*, 1997). The remaining childhood NCL genes followed in the first decade of the new millennium, again for the most part through homozygosity and linkage mapping (Nita *et al.*, 2016). The gene for Action Myoclonus Renal Failure Syndrome (Andermann *et al.*, 1989b) was one of the first disease genes to be identified using the more abundant polymorphisms established in the 2000s, namely single nucleotide polymorphisms (SNPs), which made it possible to rely on very few patients in order to identify disease genes (Berkovic *et al.*, 2008). Most recently, disease genes, including PME genes, emerged in larger numbers, through combined use of SNP mapping arrays and next-generation (whole-exome and whole-genome) sequencing. Here, identification of disease genes can be based on as few as one patient. The best example of this technical progress relates to Kufs disease (adult-onset NCL). While this disease has been known for 88 years, it was not until advanced mapping and sequencing techniques became routinely used



Figure 4. Gonzalo Rodriguez Lafora (1886-1971). After studying in Spain (with Santiago Ramón Cajal), France (with Pierre Marie and Joseph Jules Dejerine), Germany (with Alois Alzheimer and Emil Kraepelin) and the USA, Gonzalo Rodriguez Lafora returned to Spain (which he had left for Mexico during the Civil War in 1938; he returned to Madrid in 1947). As a psychiatrist, he introduced the Freudian doctrine to both Spain and Argentina, but mainly dedicated his life to the care of intellectually disabled children. During his tenure as a neuropathologist at the Government Hospital for the Insane in Washington DC, he published his landmark paper on “myoclonic corpuscles”, in German.

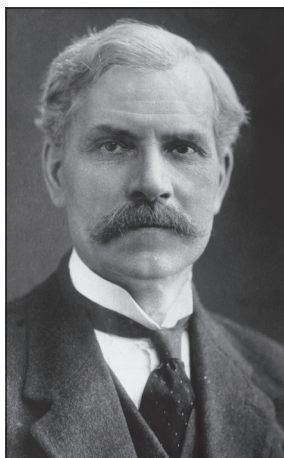


Figure 5. James Ramsay Hunt (1874-1937). After studying in Philadelphia, Paris, Berlin and Vienna, James Ramsay Hunt practised and taught neurology in New York City (Cornell University and Columbia University). His name is associated with a small cutaneous zone innervated by the *ganglion geniculi*. His contribution to the field of PME from 1914 onward was the source of great confusion; from his area of low prevalence, he selected several unrelated cases with myoclonus (and other symptoms). The term “Ramsay Hunt Syndrome”, when applied to a neurological condition with myoclonus, was used to refer to many disparate entities. The term is no longer in use, following the delimitation of discrete PME types.

that its genetic cause was uncovered. This turned out not to be a single gene but, to date, at least four different genetic entities (Nita *et al.*, 2016).

Some PMEs are very rare, caused by private mutations in single families. One example of this is the PME due to mutations in *PRICKLE1* (Bassuk *et al.*, 2008). It is expected that many such PMEs will be identified, as has been the case for other diseases. Mutation for certain genes is limited to allow for viability, but may result in a specific pathology that cannot be replicated by other defects of the same protein. Other PMEs are allelic to previously known PMEs, for example, the most common form of Kufs disease is allelic to the late-infantile variant NCL, CLN6 (Arsov *et al.*, 2011).

As recessively inherited diseases, many PMEs occur fairly frequently in pets and farm animals, due to inbreeding. This includes LD, which is widespread in certain breeds of dog (Lohi *et al.*, 2005), and various forms of NCL in dogs and sheep. In some cases, PME genes were first discovered in animals and then translated to humans, e.g. the severest form of NCL, CLN10, with fatal neonatal disease (Tyynela *et al.*, 2000; Siintola *et al.*, 2006; Steinfeld *et al.*, 2006). PME comparisons between humans and animals has also yielded fascinating insights into genome biology. For example, human ULD is a disease which is not due to the complete absence of the responsible gene (*EPM1*), but to drastic downregulation of the gene's expression caused by expansion of a dodecamer repeat sequence. This repeat is present in the promoter of the human *EPM1* gene but not in the promoter of the orthologous genes in animals. In humans, expansion of this dodecamer leads to significant downregulation but not to the complete absence of *EPM1* mRNA. No patient is reported to have, or probably exists with, a total loss of *EPM1*. Because of the unique genomic particularity within the promoter sequence of the *EPM1* gene, ULD is, therefore, a uniquely human disease and no natural animal model of the disease has been reported. As a second example, the dog genome has a similar dodecamer repeat in the *Epm2b* gene, one of the genes mutated in LD. Recurrent expansion of this repeat in *canine Epm2b* makes LD particularly common in dogs, but this mechanism does not occur in human cases with LD (Lohi *et al.*, 2005).

Conclusions

PMEs comprise a group of rare, heterogeneous genetic (mainly autosomal recessive) disorders, characterised by cortical myoclonus, other types of epileptic seizures, and progressive neurocognitive impairment. PMEs usually present in late childhood or adolescence, which distinguishes them from epileptic encephalopathies that start with polymorphic seizures in early infancy. However, adult-onset PMEs may be

due to rare gene defects or to immune or late degenerative disorders. Recent advances in this area have clarified molecular genetic basis, biological basis, and natural history, and have also provided a rational approach to diagnosis. However, PME's still remain uncommon disorders which are difficult to diagnose in the absence of extensive experience with such conditions, and this severely limits the number of expert groups in the field. Thus, despite the advances in molecular medicine, aetiology remains undetermined in a substantial proportion of patients. In particular, there are still huge areas in medically developing parts of the world, where the diagnosis of PME is probably overlooked. Therefore, the actual prevalence of these conditions is still debatable. The history of PME's shows that international collaboration and sharing experience is the right way to proceed. The Marseille conference occurred at a perfectly opportune moment, serving to clarify and classify the many PME syndromes known at that time. This was the springboard from which scientists, armed with the genetic and genomic tools that were then being invented, were able to rapidly identify causative defects. It is probably safe to say that we have now identified most PME genes, but it is equally safe to expect that many others remain to be found. Each one, however unique, will fill one of the gaps in the great PME puzzle. This will enable us to better understand this severe brain disease, and to move forward towards grasping some of the mysteries of the human brain. At the same time, the emerging picture and biological insights will allow us to find ways to provide our patients with meaningful treatment. □

Disclosures

The authors have no conflict of interest to disclose.

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Neurophysiology of myoclonus and progressive myoclonus epilepsies

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ABSTRACT – The high temporal resolution of neurophysiological recordings makes them particularly suited to faithfully describing the time course of rapid events such as myoclonus and to precisely measure its time relationship with other related activities. In progressive myoclonus epilepsies (PMEs) polygraphy with simultaneous EMG-EEG recordings is a crucial tool for defining the characteristic of myoclonic jerks their topography over different muscles (namely antagonists), their time course and relationship with vigilance muscle activation and stimulations. Moreover on polygraphic recordings it is possible to detect EEG activities associated to myoclonic jerks and define their time relationship with myoclonus thus differentiating cortical types of myoclonus from subcortically generated ones. Thanks to the back averaging technique non obvious time-locked EEG potentials can be detected on polygraphy, furthermore in stimulus sensitive myoclonus the analysis can include the potential evoked by the somatosensory stimulus (SEP). The polygraphic recording also gives information on muscle activity suppression occurring after jerk or as pure negative myoclonus. Besides the time domain analysis, techniques based on frequency analysis have been developed to evaluate EEG-EMG coherence. The neurophysiological techniques provide investigators and clinicians with an invaluable information to define the type of myoclonus and its generating circuitry thus substantially contributing in the diagnosis and management of PMEs.

Key words: progressive myoclonus epilepsies, neurophysiology, EEG-EMG polygraphy, cortical myoclonus, coherence analysis

Neurophysiological features associated with myoclonus in progressive myoclonus epilepsies

Neurophysiological recordings may be conducted over a relatively long period of time, making

them particularly suited to faithfully describing the time course of the shock-like muscle contractions which characterize myoclonus. Moreover, the combination of electroencephalographic (EEG) and electromyographic (EMG) recordings allows detection of any EEG correlates of myoclonus and high

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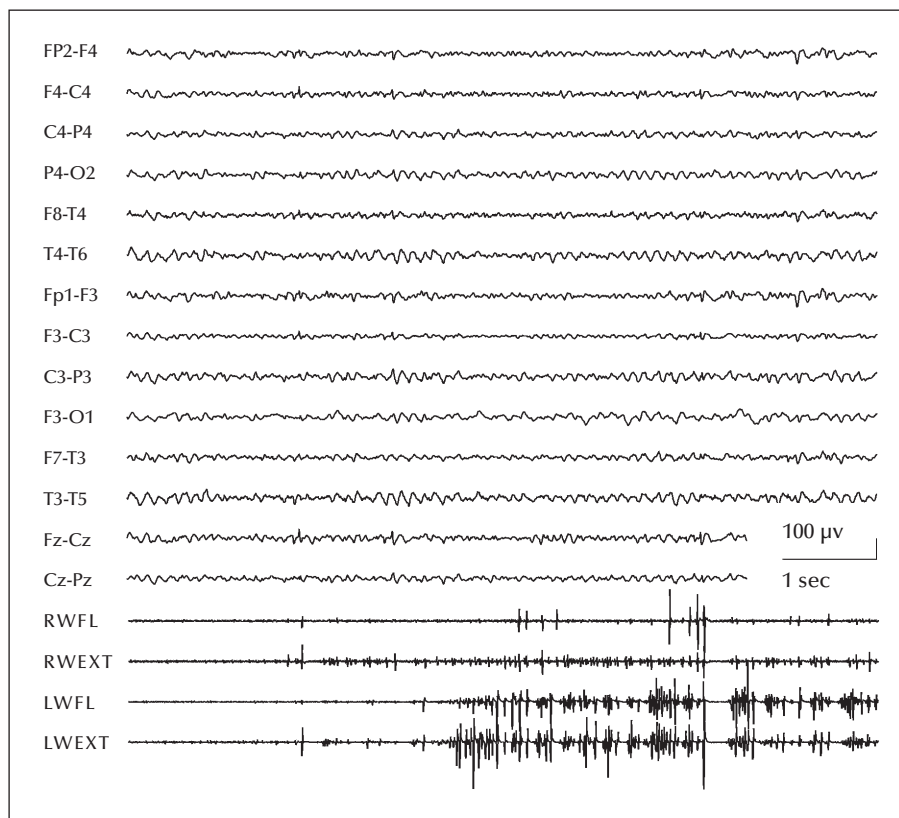


Figure 1. Patient with Unverricht-Lundborg disease. Polygraphic recording with the patient at rest showing fragmentary multifocal myoclonus without overt EEG correlate (EMG artefacts due to myoclonic jerks involving the face are superimposed onto the EEG trace).

precision measurement of their time relationship to muscle jerks. For these reasons, neurophysiological analysis is a first-line approach to myoclonic syndromes, both in terms of clinical characterization and pathophysiological investigation.

The first section of this article deals with the neurophysiological techniques suitable for characterizing different types of myoclonus, while the second section addresses the value of neurophysiology in defining the clinical presentation of some progressive myoclonus epilepsies (PMEs) (Minassian *et al.*, 2016).

Neurophysiological analysis of myoclonus

The correlation between EEG and EMG activities associated with myoclonus is the basis for investigating the pathophysiology of myoclonus as well as the clinical diagnosis of PMEs. Several signal analysis techniques relating to time and frequency domains, which are currently employed to detect EEG correlated with myoclonus and used to investigate its pathophysiology, will be highlighted in this first section.

Polygraphic recordings and EEG-EMG correlations in progressive myoclonus epilepsy

In epileptic disorders, polygraphy with simultaneous recording of EEG-EMG activity can provide relevant information for defining the characteristics of a motor manifestation and the relationship with concomitant EEG activity. Moreover, it can be useful to identify subtle and apparently subclinical manifestations, and is necessary for precise investigation of the temporal relationship between EEG and EMG phenomena (Tassinari and Rubboli, 2008).

In PMEs, polygraphic recording can be a crucial tool for the investigation and definition of the characteristics of myoclonic phenomena, which represent one of the cardinal features of this vast group of diseases. EMG is usually recorded using surface electrodes placed on the skin overlying the muscles involved in myoclonic activity, which should be clearly identified by clinical examination (*figure 1* figure 1). The cortical correlates of myoclonus have also been analyzed using magnetoencephalography (MEG), which can complement the

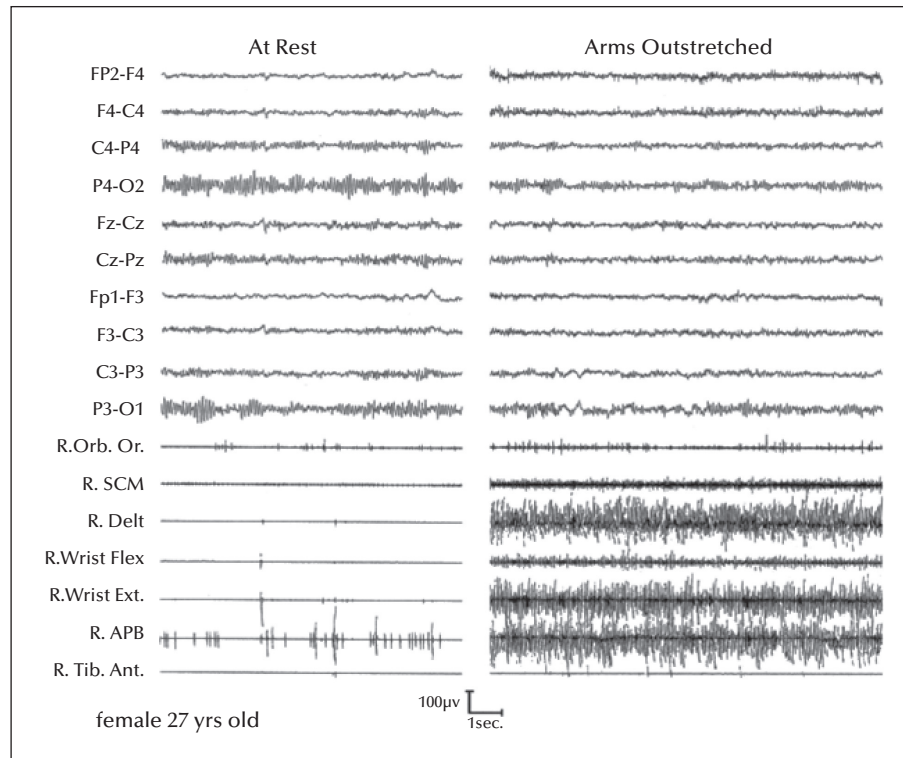


Figure 2. Patient with PME associated with *SCARB2* mutation, at disease onset. Left panel: the patient is at rest; EEG shows preserved background activity without epileptiform abnormalities; on the EMG channels, erratic multifocal myoclonic jerks without EEG correlate are evident. Right panel: the patient keeps her arms outstretched; the EMG channels show continuous rhythmic cortical myoclonic activity at a frequency around 12-20 Hz.

EEG information in terms of the cortical generators of myoclonus (Mima *et al.*, 1998).

Myoclonus is an essential and defining feature of PMEs. It can occur spontaneously or be induced or exacerbated by a variety of stimuli (such as light, sound, touch and emotional strain) and active movement or posture maintenance. At rest, in PMEs, myoclonus is commonly fragmentary and multifocal, and is particularly apparent in the musculature of the face and distal limbs (figure 2, left panel). Action myoclonus, in which movement (as an attempt or intention to move) initiates jerking, is a common feature in almost all the conditions underlying PMEs, and can be extremely disabling. At rest, the EMG expression of myoclonus is a burst of myoclonic potentials of brief (100 ± 50 ms) duration, typically occurring synchronously in agonist and antagonist muscles. If myoclonus occurs in a contracting muscle, then after the myoclonus there is a brief (50-100 ms) suppression of muscle activity.

A period of suppression of muscle activity without a preceding myoclonus can also produce a negative jerk, due to a sudden interruption and resumption of ongoing muscular activity. This latter phenomenon is referred to as 'negative myoclonus' and is related to a mechanism of supraspinal inhibition lasting from 100 to 500 ms. In PMEs, a mixture of positive and

negative myoclonus is common in the same patient. The EMG correlate of a single action myoclonus is an EMG potential of short duration (20 to 30 ms), which appears synchronously in agonist and antagonist muscles (figure 3). It is usually followed by an EMG-silent period lasting 40 to 120 ms (in rare cases up to 300 ms). The myoclonic bursts and silent periods

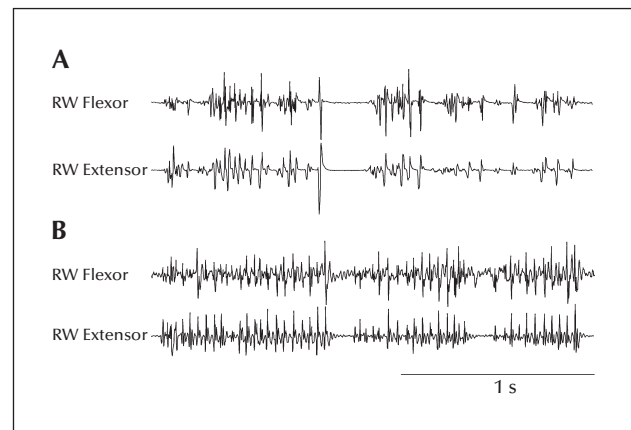


Figure 3. EMG recording from two antagonist muscles of the right wrist (RW) showing the presence of synchronous EMG bursts with an irregular (A) and quasi-rhythmic (B) course.

are seldom related to EEG spike and waves or polyspike and waves (Tassinari *et al.*, 1974). More often, the EEG correlate is small in transient amplitude, merging with spontaneous EEG activity. The cortical nature of the myoclonic event can be assumed when a time-locked spike or other EEG paroxysmal event is detected on EEG-EMG polygraphic recordings either by visual inspection (*figure 1*) or by the back-averaging techniques that will be described in detail below. Cortical myoclonus, regardless of whether it is positive or negative, is attributed to a pathologically enhanced excitability of the primary sensorimotor cortex (Obeso *et al.*, 1985), although negative myoclonus may also originate from the pre-motor or supplementary motor area (Rubboli *et al.*, 2006). Additionally, increased excitability of subcortical circuits is considered to be the causative mechanism of subcortical myoclonus which may coexist with cortical myoclonus in PME. Additional findings that support the cortical or subcortical origin of myoclonus and their pathophysiological implications will be discussed in detail below.

Polygraphic recordings of PMEs can be complemented with stimulation procedures aimed at revealing their stimulus sensitivity. Photic stimulation should always be applied due to the high frequency of photic reflex myoclonus and/or EEG photosensitivity. Intermittent photic stimulation (IPS) may induce bursts of polyspike-wave discharges associated with massive myoclonic jerks. If the triggering stimulus is prolonged, the clinical response may progress to a generalized convulsion. The mechanism of photic reflex myoclonus involves both occipital and motor cortices, with bilateral spread, presumably mediated by transcallosal connections and propagation down the spinal cord via fast-conducting cortico-spinal pathways (Rubboli *et al.*, 1999).

Although cortical positive myoclonus usually occurs irregularly, it may appear fairly rhythmical, and may even resemble a tremor (hence the term 'cortical tremor') in some cases of PME (Toro *et al.*, 1993). Brown & Marsden (1996) reported repetitive myoclonus, both spontaneous and stimulus-evoked, at short intervals of around 20 ms in patients with cortical reflex myoclonus. This was associated with repetitive EEG discharges of the same frequency. Enhanced motor cortex hyperexcitability, as the result of loss of intracortical inhibition, possibly due to abnormal GABA-B mediated inhibitory circuits, has been postulated to underlie the susceptibility to generate rhythmic myoclonic activity (Valzania *et al.*, 1999). Rhythmic cortical myoclonus, at a frequency of around 12-20 Hz has also been reported in the recently described PME associated with SCARB2 mutations (Berkovic *et al.*, 2008; Rubboli *et al.*, 2011) (*figure 2*, right panel). This rhythmic myoclonus, reminiscent of postural tremor, can be evident at disease onset. Its cortical origin has been demonstrated mainly

by coherence and phase analysis of EEG-EMG signals, indicating a significant EEG-EMG coupling and a direct corticospinal transfer (Rubboli *et al.*, 2011). Rhythmic jerks in the beta band have also been described in some PMEs associated with rare storage diseases (Brown *et al.*, 1999; Panzica *et al.*, 2003; Canafoglia *et al.*, 2006).

Additional information can be drawn from sleep recordings which should be performed in the PME work-up whenever possible. The results of sleep studies in different PMEs will be discussed in the second section of this article.

Coherence study of EEG-EMG relationship in progressive myoclonus epilepsies

Since synchronization between muscles and cortical activities was demonstrated in the 1990s (McLachlan & Leung, 1991; Farmer *et al.*, 1993), methods of analysis relating to the frequency domain have become an important tool for investigating the human motor system, particularly in terms of studying whether specific patterns of neuronal synchrony may be of diagnostic value (Brown *et al.*, 1999; Grosse *et al.*, 2003).

In the last decade, spectral analysis, namely coherence and phase analysis, has been increasingly applied to investigations of the relationship between rhythmic or quasi-rhythmic myoclonic events and EEG oscillations. Indeed, the relationship between EEG or MEG activity and voluntary or involuntary muscle contraction can be studied by calculating the linear cross-correlation over certain frequency bands (coherence) during sustained muscle contraction (cortico-muscular coherence) (Conway *et al.*, 1995; Salenius *et al.*, 1997; Halliday *et al.*, 1998; Mima & Hallett, 1999).

Spectral analysis appears to be a powerful method for detecting EEG-EMG coherence (Mima & Hallett, 1999) and MEG-EMG (Salenius *et al.*, 1997; Silen *et al.*, 2000) or EMG-EMG relationships in cortical myoclonus, and has several advantages over the more commonly used jerk-locked back-averaging technique (see below) for a number of reasons. High-frequency myoclonic discharges do not prevent the analysis, no arbitrary trigger level has to be chosen, results can be evaluated from a statistical point of view, and the technique can be automated, such that long sections of signal traces can be analyzed over a short epoch. However, the estimation of EEG-EMG coherence requires relatively artefact-free EEG/MEG epochs, and the recording itself, particularly in children with myoclonus or involuntary jerks, may be difficult and time-consuming.

Brown *et al.* (1999) demonstrated cortical activity related to myoclonic jerking through frequency analysis in five patients in whom jerk-locked back-averaging

failed to show any clear EEG transient associated with myoclonic jerks. To estimate coherence and phase spectra, couples of channels are usually investigated by means of cross-spectral analysis based on traditional Fast Fourier Transform (FFT). Selected data are usually divided into consecutive non-overlapping segments, transformed in the frequency domain and then averaged. A trade-off should be considered in the FFT approach between frequency resolution and spectral variance. As window length decreases, variance also decreases, but spectral resolution becomes poorer.

An alternative approach is based on parametric autoregressive (AR) models. The main advantages of spectral AR estimates over FFT-based methods are that they significantly improve frequency resolution since parametric spectra can be evaluated numerically at any number of frequencies and do not require any averaging to obtain a smoothed spectrum (Gath *et al.*, 1992; Pardey *et al.*, 1996; Spyers-Ashby *et al.*, 1998). Conversely, the FFT-based spectra can be evaluated only on the number of samples (N) with harmonically related frequencies. This advantage is particularly important for the analysis of short sequence lengths or epochs characterized by rapid dynamic changes. Moreover, AR spectra can be obtained without windowing the data since no assumptions about samples outside the data sequence are needed. In addition, the inclusion of a noise term in the AR model means that the estimated spectrum is smooth, since its shape depends only on the values of the coefficients used to model the signal. In contrast, in the FFT-based analysis, random fluctuations due to noise can be reduced only by the averaging procedure. The improvement is related to the number of degrees of freedom of the AR model, which is given by N/p where N is the number of samples and p is the model order (Gath *et al.*, 1992). Using the AR model, the number of AR parameters needed to model a time series is typically much lower than the total number of data points composing the signal, and this therefore gives a statistically desirable compact representation of the signal. For FFT methods, by comparison, it is necessary to determine as many coefficients as there are points in a particular data segment. In itself, this is statistically undesirable and this is the reason why one needs to average over a large number of data segments in order to obtain an appropriate spectrum. As a result, it is commonly claimed that AR spectral estimates tend to be more robust than FFT estimates when working with a small data set. These characteristics make it possible to estimate the myoclonic bursts that need to be isolated from periods of normal muscle contractions (as is the case with Unverricht-Lundborg patients), or from the spontaneous 'epileptic' myoclonus associated with diffuse spike-wave discharges (as is the case with Lafora patients) (Panzica *et al.*, 2003). The main problem with AR models is the choice of model

order. It is important to stress that the model order determines the number of frequency components contained in the spectra (in a univariate model, the maximum number of peaks in the power spectrum is half of that of the model order and thus determines the "frequency resolution of the spectrum" (Schlogl & Supp, 2006). The main advantage of the FFT over AR spectral estimation is its computational efficiency.

Using a parametric approach, multivariate AR models can be used to provide a multivariate representation of the signals, from which appropriate measures of coupling can be estimated. In 1991, Kaminski and Blinowska proposed the Directed Transfer Function (DTF) (Kaminski & Blinowska, 1991), a multichannel estimator of the intensity and direction of activity flow, based on a multichannel autoregressive model between couples of channels as a function of the frequency (Mima *et al.*, 2001; Cassidy & Brown, 2003).

In 2001, Baccalá and Sameshima proposed a different multichannel approach, the partial directed coherence (PDC), which allows the direction of information flow between any of the two channels to be estimated by subtracting the interactions and possible common influences due to other remaining simultaneously observed time series (Baccalá & Sameshima, 2001; Meng *et al.*, 2008). By applying this approach, Panzica *et al.* (2014) were recently able to demonstrate, in patients with cortical myoclonus, a significant increase in cortical outflow towards activated muscles, in comparison to healthy controls. Moreover, they showed a more robust EMG outflow toward ipsi and contralateral cortical areas which could maintain jerk recurrence.

In addition, non-stationary or time-varying multivariate AR models have recently been developed and can be applied to study dynamical changes associated with cortico-muscular coupling in patients with myoclonus, when the statistical properties of the signals change substantially over time. Panzica *et al.* (2010) studied myoclonus-related EEG changes in patients with two forms of progressive myoclonus (Unverricht-Lundborg disease and sialidosis) using bivariate time-varying autoregressive models (TVAR). The results indicated that it was possible to detect the presence of prominent peaks of EEG-EMG coherence between the EMG and contralateral frontocentral EEG derivation by TVAR analysis in all patients and, most importantly, differences were disclosed relating to time-frequency spectral profiles correlated with the severity of myoclonus (*figure 4*).

In patients with cortical myoclonus, regardless of aetiology, frequency analysis showed the presence of an exaggerated coherence peak in the beta band (mainly at 15-20 Hz) between sensorimotor cortex activity and EMG activity that is normally rectified, recorded from muscles co-activated by myoclonic jerks. In this

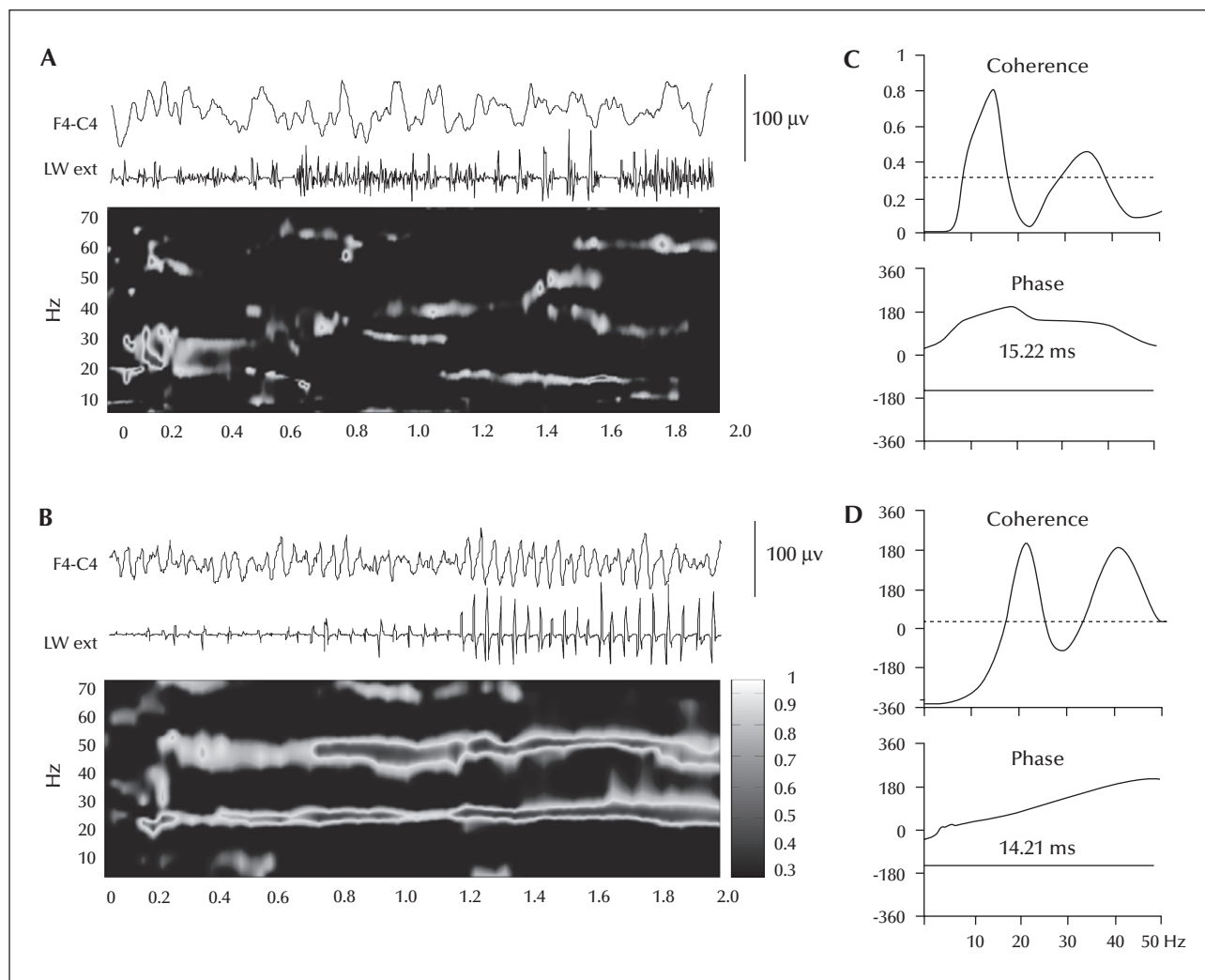


Figure 4. TVAR analysis of a movement-activated myoclonus in a patient with Unverricht-Lundborg disease (A) and sialidosis (B). Note in the coherence spectrum (B), the presence of significant EEG-EMG coherence in the beta and gamma bands remaining consistent throughout the movement, in contrast with the intermittent course in the ULD patient. (C and D) Mean coherence (the dotted line indicates the 95 per cent confidence limit) and phase spectra obtained from TVAR analysis. The estimated time lag was fitted with a cortico-spinal time transfer.

frequency range, often the cortical activity precedes EMG by a time period which is appropriate for conduction in the fast conduction pyramidal pathway (Brown & Marsden, 1996; Brown *et al.*, 1999; Valzania *et al.*, 1999; Silen *et al.*, 2000; Grosse *et al.*, 2003; Panzica *et al.*, 2003). Sometimes, however, the phase difference indicates a time lag that is lower than expected for the fastest conducting pathway (Brown *et al.*, 1998; Ohara *et al.*, 2000). This phenomenon may be due to the presence of multidirectional activities which contribute to the cortico-muscular coherence between muscle and cortex (efferent control from the primary motor cortex to muscle and afferent feedback from the periphery) (Panzica *et al.*, 2012). These findings support the idea that myoclonus may be due to an exaggeration of the

cortical drive to muscle observed during voluntary contractions in healthy subjects (Brown *et al.*, 1999). Unlike coherent beta activity, a coherent gamma peak is not a consistent feature. In healthy subjects, a similarly inconsistent presence of coherent gamma activity has been hypothetically attributed to the variable cortical activation associated with attention or differences in functional cortical-muscular coupling depending on the strength of the muscle contraction (Brown *et al.*, 1998; Mima & Hallett, 1999). Panzica *et al.* (2003), based on a study of patients with different types of PMEs, found constant coherent gamma coherence only in the patients with sialidosis (*figure 4*). In these patients, myoclonic jerks entirely replaced the physiological muscle contractions, which occurred at the

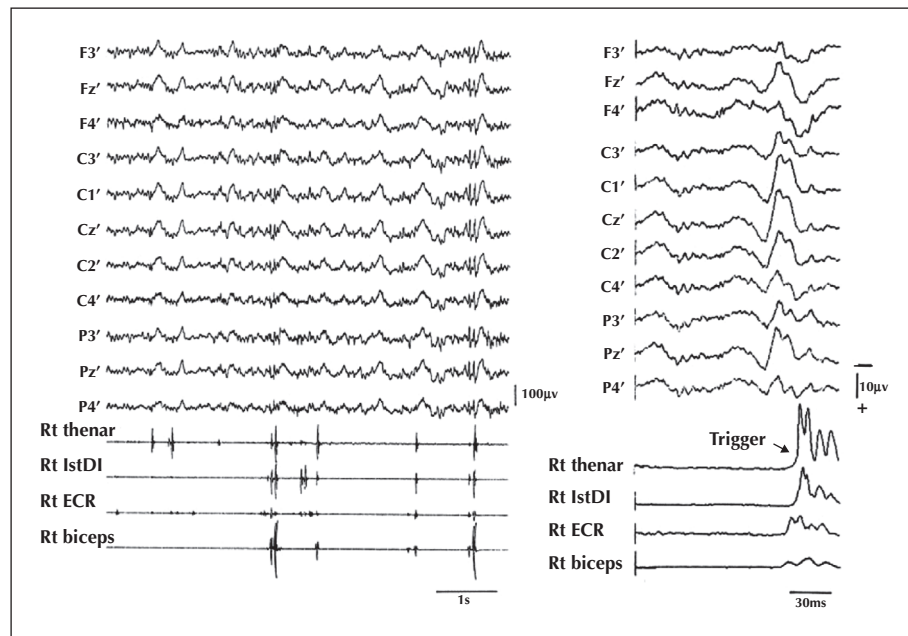


Figure 5. Left panel: EEG-EMG polygraphic recording in a patient with PME manifesting positive myoclonus in the hands at rest. Note that most myoclonic jerks were associated with a spike-and-wave complex on EEG, and the upward negativity recorded from the ipsilateral earlobe electrode. ECR: extensor carpi radialis muscle; 1stDI: first dorsal interosseous muscle; Rt: right. Right panel: Records of jerk-locked back-averaging obtained from the same patient. The onset of the EMG discharge from the right thenar muscle was used as a trigger pulse to back-average multichannel EEGs. EEG was recorded from the ipsilateral earlobe electrode. A positive-negative, biphasic EEG spike is observed maximally near the midline vertex, slightly shifted to the left (C1-Cz), and widespread over the scalp. Note that the myoclonic EMG discharge, which was also averaged with respect to the same fiducial point, spreads rapidly from the proximal muscles to the distal ones (Modified from Shibasaki & Hallett [2005]).

onset of movement and throughout posture maintenance, with a rather rhythmic and protracted course, reminiscent of the rhythmical nature of a tremor.

Brown (2007) suggested that prominent and extensive synchronization in the beta band might limit the ability of sensorimotor neurons to code information in time and space, since these neurons preferentially fire at a frequency locked to the beta rhythm. These synchronized discharges of pyramidal neurons are transferred through the pyramidal tract to the peripheral motor system, resulting in synchronous motor unit discharges that may lead to myoclonic jerks, rather than the sustained muscle contractions or movements normally resulting from the relatively asynchronous activation of motor units.

Neurophysiological investigation of brain circuitry sub-serving myoclonus

In this section, we review some neurophysiological techniques in the time domain that may complement the frequency analysis of EEG-EMG polygraphic recordings to investigate whether muscle jerk is associated with any EEG activity and to define their reciprocal time relationship. The first important point

concerns the existence of myoclonus-associated scalp electrographical events that precede the myoclonic jerk, thus supporting a cortical origin of myoclonus.

Jerk-locked back-averaging

The potential of this technique to detect non-obvious EEG correlates of myoclonus was first reported by Shibasaki and Kuroiwa (1975) and has been widely employed since then to study involuntary movements. The principle upon which it is based is that of averaging the EEG (or MEG) recording preceding the muscle jerk such that all activities which are not consistently time related to the jerk are subtracted from one other, while the time-locked EEG potentials preceding myoclonus will summate, resulting in easily recognizable transients which are well-suited to morphological analysis and time relationship calculation (*figure 5*).

According to Kornhuber and Deecke (1965) in their original study of movement-related EEG activities, the backward analysis could be performed by playing back a magnetic tape where polygraphic the EEG-EMG recording has been stored or by introducing a delay-line analogue circuit into the EEG recording system (Franceschetti *et al.*, 1980). With the increased availability of digitized recordings, analysis of the EEG

preceding the myoclonic jerk can be easily obtained by setting the analysis window at 200 ms before and 200 ms after the myoclonus onset. The trigger pulse is generated by the myoclonus-related EMG potential either in its original or rectified shape. The definition of the true onset of the EMG potential is crucial to ensure reliable results. The method can also be applied to the study of the negative myoclonus of cortical origin, in which the onset of the EMG silent period is used as a trigger for back-averaging EEG (Ugawa *et al.*, 1989).

An EEG potential preceding the muscle jerk by 15–20 ms may express a cortical discharge responsible for the myoclonus and is therefore considered to be compatible with its cortical origin. Shorter intervals or an inverted time relationship with the jerk preceding the EEG potential would rather support a cortical response evoked by the muscle jerk originating in some subcortical structure.

Electrophysiological study of reflex myoclonus

In 1939, Adrian and Moruzzi showed that the stimulus-evoked myoclonus observed in cats anaesthetised with chloralose was due to a discharge travelling in the pyramidal tract and time-locked to a cortical wave (Adrian & Moruzzi, 1939). This “cortical reflex myoclonus” did not fully account for the muscle jerk observed in the anaesthetised animals, as decortication did not eradicate it completely. The origin of the residual component was found to be in the reticular formation. The observation of stimulus-sensitive myoclonus in different human diseases made it possible to characterize cortical versus reticular reflex myoclonus in several pathological disorders, including PMEs. Their main features are summarized here, in line with the work by Shibasaki and Hallett (2005) and Shibasaki (2012).

Cortical reflex myoclonus

Cortical reflex myoclonus, evoked by electrical shocks delivered to the median nerve at the wrist, is associated with a giant somatosensory evoked potential (SEP), first reported by Dawson in 1947 (Dawson, 1947), with an extreme enlargement of P25 and subsequent components (up to 10 times as large as the normal value), whereas the initial components N20 and P20 are normal or only slightly enhanced (*figure 6*).

However, the studies of somatosensory evoked magnetic fields (Karhu *et al.*, 1994; Mima *et al.*, 1998) shows that M20 is also slightly enhanced in some cases, suggesting hyperactivity of the sensorimotor thalamo-cortical loop. Moreover, these studies demonstrate that the hyperactivity involves the primary somatosensory cortex but not the second somatosensory area, which is not hyperexcitable.

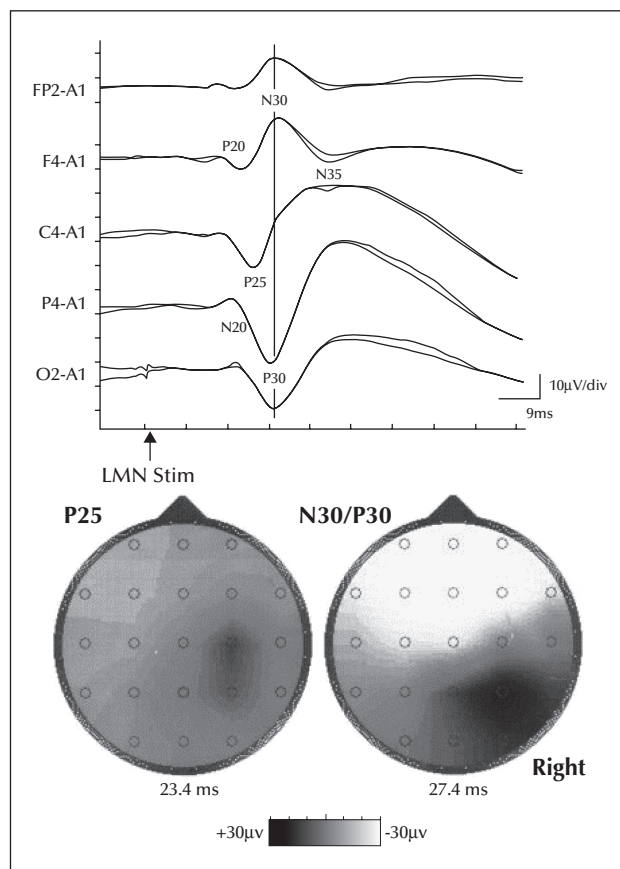


Figure 6. Somatosensory evoked potential (SEP) waveforms following electrical stimulation of the left median nerve (LMN Stim) of the wrist in a patient with Lafora disease presenting with PME (upper panel), with scalp topography consisting of two peaks (lower panel). Four peaks are clearly distinguishable: N20/P20, P25, N30/P30, and N35. N30/P30 shows a similar distribution to N20/P20 (not shown here), although with opposite polarity. A1: left earlobe electrode. Note the upward negativity. (From Ikeda *et al.*, [1995]).

Giant potentials can also be evoked by flash stimulation in photosensitive myoclonus, which is also considered a type of cortical reflex myoclonus (Shibasaki & Neshige, 1987). The areas of enhanced excitability include the occipital and frontal cortices, in which hyperexcitability is considered to account for photo-induced muscle jerks.

The somatosensory stimulus-induced cortical reflex myoclonus is readily evoked in the thenar muscle by stimulating the median nerve at the wrist with a latency of 45 ms, which corresponds to that of the long-loop reflex known as the C reflex (C for ‘cortical’), as reported by Sutton and Mayer (1974). The P25 peak of giant SEP and cortical potential preceding the muscle jerk, as revealed by jerk-locked back-averaging, show a similar topography and time interval relationship to myoclonus and are considered to be related to the same pathophysiological mechanisms, *i.e.*

cortical hyperexcitability (Shibasaki *et al.*, 1978). Analysis of the latencies of the cortical reflex myoclonus recorded from different muscles shows a rostro-caudal order of activation, compatible with a cortical origin of the myoclonus, generating a signal which travels down the brainstem (Hallett *et al.*, 1979).

Reticular reflex myoclonus

Reticular reflex myoclonus differs from cortical reflex myoclonus in terms of SEP, which is not enhanced, and order of activation of muscles; bulbar muscles are activated first and rostral cranial (*i.e.* facial muscles) muscles and caudal muscles (*i.e.* limb muscles) are involved only subsequently (Hallett *et al.*, 1977).

Cortical and reticular reflex myoclonus can coexist in the same patients with PME. The coexistence of subcortical and cortical myoclonus has been demonstrated on the basis of the discrepancies between the latency of reflex myoclonus and the sum of afferent and efferent times to and from the cortex, as evidenced by TMS studies (Cantello *et al.*, 1997).

Neurophysiological findings in different progressive myoclonus epilepsy forms

PMEs share common neurological signs that include progressively worsening cortical myoclonus and epileptic seizures, with classic onset in late childhood and adolescence. Other neurological symptoms, namely dementia and ataxia, are typically associated with myoclonus-epilepsy syndromes, and occasionally further signs and symptoms are due to the specific impairment of nervous or other systems (Marseille Consensus Group, 1990).

The PME phenotype includes a 'core' symptom: multifocal reflex (action-induced) myoclonus. This type of myoclonus is assumed to be cortically generated, since it is typically associated with 'subtle' central EEG changes that can be studied using EEG-EMG relationship analysis (including jerk-locked back-averaging and other techniques). Moreover, cortical myoclonus is coupled with neurophysiological features reflecting neocortical hyperexcitability, such as 'giant' evoked potentials and enhanced long-loop reflexes (Shibasaki, 1988; Shibasaki & Thompson, 2011).

PMEs are derived from heterogeneous genetic disorders (Serratosa *et al.*, 1999; Ramachandran *et al.*, 2009; de Siqueira, 2010), probably with distinct pathological mechanisms, including neural degeneration (Unverricht-Lundborg and dentatorubral-pallidoluysian atrophy), storage disorders (Lafora disease, neuronal-ceroid-lipofuscinoses, sialidoses, Gaucher III, Niemann Pick type C, and action myoclonus-renal failure syndrome), mitochondrial

disorders (myoclonic epilepsy associated with ragged-red fibres), and ion channel dysfunction (Azizieh *et al.*, 2011). Advances in biomolecular research continuously enrich our knowledge of the genetic background of PMEs and their pathogenesis.

Since the causes of PMEs are heterogeneous, they can be expected to affect cortical and subcortical brain structures in varying ways and probably by different mechanisms.

Due to different pathogenetic/neuropathological mechanisms, PME syndromes may present partially different neurophysiological features, which can reflect the prominent dysfunction of distinct cerebral areas or the occurrence of associated symptoms (for instance, the involvement of the peripheral nervous system). Specific neurophysiological findings may be important for diagnostic assessment and phenotypic classification (Kasai *et al.*, 1999; Panzica *et al.*, 2003; Canafoglia *et al.*, 2004; Canafoglia *et al.*, 2010) and may help quantify the severity of the clinical disorder (Garvey *et al.*, 2001) at diagnosis and during follow-up. Indeed, changes in neurophysiological findings relating to the disease course may reflect spontaneous evolution or relate to therapeutic interventions (Kobayashi *et al.*, 2011).

In this section, we address the differences between neurophysiological findings in the main PME forms, Unverricht-Lundborg (EPM1) and Lafora body disease (EPM2), and in some rarer diseases: sialidoses and neuronal-ceroid-lipofuscinoses (NCL) in adults (Kufs disease). With this aim, we report on the EEG-EMG features and neurophysiological findings obtained using somatosensory evoked potentials (SSEPs), long-loop reflexes (LLR) and transcranial magnetic stimulation in these diseases and their changes over time.

EEG-EMG findings may be useful to differentiate between different PMEs

EEG-EMG polygraphy

Unverricht-Lundborg (EPM1)

The EEG-EMG features of EPM1, originally described in patients with Baltic myoclonus from a restricted geographical area (Koskiniemi *et al.*, 1974; Norio & Koskiniemi, 1979), highlight diffuse EEG background slowing with recurrent paroxysms of spike and polyspike and waves, focal spikes in the central region, and marked photosensitivity. A different disease course, probably resulting from more effective treatment (usually including valproate), emerged in more recent observations of large case series. Indeed, EPM1 patients observed in recent years have generally demonstrated normal to slightly slow EEG background and brief and rare epileptic paroxysms of spikes

or polyspikes, occasionally associated with spontaneous isolated myoclonic jerks (Canafoglia *et al.*, 2004; Ferlazzo *et al.*, 2007). Segmental myoclonic jerks occurring with voluntary or passive movements (action myoclonus), unrelated to obvious EEG 'epileptic' paroxysms, remain the prominent symptom. Epileptic paroxysms and photosensitivity tend to disappear over the years, together with a relatively stationary phase of the disease (Magaudda *et al.*, 2006; Kälviäinen *et al.*, 2008; Genton, 2010), while some observations indicate progressive impairment of the physiological EEG sleep pattern (Ferlazzo *et al.*, 2007).

Lafora body disease (EPM2)

The EEG-EMG picture observed in EPM2 patients is strikingly different from that of Unverricht-Lundborg, at least in the intermediate and advanced stages. As described by Tassinari *et al.* (1978), in the first disease period, EEG features may resemble those of primary generalized epilepsy due to preserved background activity and the occurrence of fast spikes, polyspikes, waves and photosensitivity. However, shortly after onset, despite advanced pharmacological treatments, the EEG background markedly slows and posterior or diffuse paroxysms of multiple spikes, associated with atypical (myoclonic-atonic) seizures or absences, sometimes resulting in a non-convulsive status (Fernández-Torre *et al.*, 2012), recur with a high frequency.

Moreover, myoclonus in EPM2 patients shows rather peculiar features, including prominent negative EMG phenomena (Shibasaki, 1995). Voluntary motor activity consistently enhances the occurrence of myoclonic jerks on the activated segment, but it is often difficult to distinguish action myoclonus from the almost incessant spontaneous myoclonus (Canafoglia *et al.*, 2004).

Sialidoses

EEG background activity is substantially preserved, although it may include a moderate amount of theta activity. Brief paroxysms of bilateral spikes and

polyspikes and waves, with maximum amplitude on the central EEG regions, may be present, combined with rare spontaneous bilateral myoclonic jerks. Photosensitivity is mild or absent. Myoclonus mainly occurs during motor activity and typically takes a pseudo-rhythmic course (Rapin *et al.*, 1978; Canafoglia *et al.*, 2011). Rhythmic EEG fast activities are sometimes visible on central and vertex regions, occasionally associated with spikes. The movement-activated central fast rhythm (Kelly *et al.*, 1978), sometimes intermingled with spikes, constitutes a rather typical correlate of the rhythmic myoclonus observed during motor activation.

Neuronal-ceroid-lipofuscinoses (NCL)

Clinical and electrophysiological features of adult-onset NCL (Kufs disease) were critically revised by Berkovic *et al.* (1988). Recently, *CLN6* gene mutations have been found in PME patients with Kufs disease (Arsov *et al.*, 2011). The EEG shows paroxysms of spike-and-slow-wave complexes (Berkovic *et al.*, 1988) or multiple generalized spikes without any slow component (Binelli *et al.*, 2000). Multiple spike discharges may be synchronized with myoclonus and the slow waves with brief EMG, silent in limb muscles (Berkovic *et al.*, 1988). The photoparoxysmal and myoclonic response to photic stimulation is particularly strong both at slow and high (1 to 100 Hz) stimulus frequencies, and low-frequency photoparoxysmal response might be an early clue for diagnosis (Guellerin *et al.*, 2012). Table 1 summarizes the information on EEG-EMG features.

Evolution of EEG-EMG findings during sleep

In Unverricht-Lundborg disease, sleep studies demonstrate:

- (1) a lack of activation of generalized paroxysmal discharges;
- (2) and the appearance of focal multiple fast spikes occurring in repetitive bursts, localized over the

Table 1. EEG-EMG features (in treated patients).

| | Background | Epileptic paroxysms | | PPR | Spontaneous myoclonus | Action myoclonus |
|------------|-------------|---------------------------------|------------|-----|-----------------------|------------------|
| | | Type | Occurrence | | Occurrence | |
| EPM1 | alpha-theta | Diffuse SW/PSW | + | + | + | +++ |
| EPM2 | theta-delta | Diffuse SW/PSW and occipital SW | +++ | ++ | +++ | ++ |
| Sialidoses | alpha-theta | Diffuse SW/PSW | + | +/- | + | +++ |
| Kufs | alpha-theta | Diffuse SW/PSW | ++ | +++ | ++ | +++ |

Table 2. Neurophysiological parameters in different PME.

| | SSEP | LLR | SP | CRT | MEP facilitation | SICI | ICF | LICI |
|------------|----------------------|-----|-----------|---------|---------------------|---------|---------|---------|
| EPM1 | Enlarged N20-P25 | ++ | Prolonged | Reduced | +++ | Reduced | Normal | Normal |
| EPM2 | N20-P25 and N3 giant | + | ND | Reduced | +++ | Reduced | Reduced | Reduced |
| Sialidoses | Normal or giant | ++ | Reduced | Reduced | ND | ND | Normal | ND |
| Kufs | Normal or giant | ND | ND | ND | ND | ND | ND | ND |

midline and centroparietal regions, more frequently during REM sleep, particularly when eye movements are abundant. These fast spikes can be time-locked to myoclonic jerks, particularly in muscles which show a striking action myoclonus during wakefulness. In Lafora disease, sleep organization is radically altered, with the barely recognizable different stages. Paroxysmal activity does not appear to increase during sleep; diffuse multiple fast spikes show variable amplitude and topography and can be intermixed with fast activity, while posterior spikes persist during slow sleep and can appear enhanced during REM sleep (*table 2*).

Somatosensory evoked potentials

Giant potentials

In PME patients, early SEP components are typically enlarged, with major emphasis of the P25-N33 waves, which are thought to be connected to the occurrence of reflex myoclonus (Kakigi & Shibasaki, 1987). In a study performed in EPM1 and EPM2 patients (Canafoglia *et al.*, 2004), the peak-to-peak amplitude of N20-P25 was abnormally enlarged in both groups, but N33 was often poorly defined in EPM2 patients, merging in a broad negative wave (N3, peaking between 43.8 to 66.6 ms). Moreover, the P25-N60 and N60 amplitudes were significantly larger in EPM2 patients in comparison to both controls and EPM1 patients (*figure 7*). Increased SSEP amplitudes may also occur less consistently in patients with sialidoses and Kufs disease, however, they are unlikely to be useful in the differential diagnosis (Berkovic *et al.*, 1988; Canafoglia *et al.*, 2011).

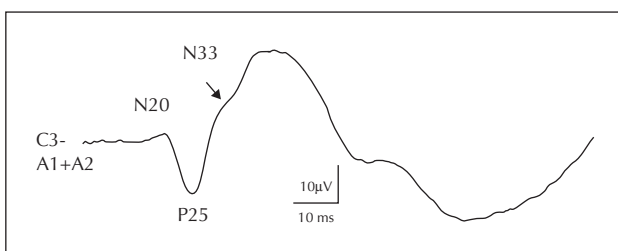


Figure 7. Giant SEP recorded in a patient with Lafora body disease showing an enlarged N20-P25 component and extremely enhanced P25-N33, merged into a very large middle-latency wave.

The amplitude of the SSEP components may change over the course of the disease. A recent study (Kobayashi *et al.*, 2011) suggested that in EPM1 patients, the reduction or disappearance of a middle-latency cortical component (labeled N40) is associated with a decrease in epileptiform discharges at the plateau stage of the disease. Conversely, in EPM2 patients, early and middle-latency SSEP components may increase during the disease, alongside the worsening course of the myoclonus and seizures.

SEP latencies

Various studies indicate that the latencies of the early cortical SSEP components increase (Mervaala *et al.*, 1984; Canafoglia *et al.*, 2004) due to a delay in the central conduction pathway from the thalamus to the somatosensory cortex, while in EPM2 patients, the conduction studies gave normal results (Canafoglia *et al.*, 2004) (*figure 8A*).

In patients with sialidosis, the possible co-occurrence of polyneuropathy is typically associated with prolonged latencies (Canafoglia *et al.*, 2011).

Long latency reflex (LLR)

LLR is the neurophysiological correlate of reflex myoclonus. A comparative study has suggested that facilitated LLRs are more common in EPM1 than EPM2 patients, probably reflecting the prominence of reflex myoclonus in this disorder. LLR latencies were briefer in EPM1 than in EPM2 patients (Canafoglia *et al.*, 2004) (*figure 8B*).

In sialidoses patients, the shape of LLR often includes multiple waves, probably reflecting the rhythmic occurrence of the jerks (Franceschetti *et al.*, 1980; Tobimatsu *et al.*, 1985; Canafoglia *et al.*, 2011).

Cortical relay time (CRT)

According to a comparative study (Canafoglia *et al.*, 2004) the CRT was significantly briefer in both EPM1 and EPM2 patients, in comparison to that of healthy controls. The CRT was particularly brief in EPM1 patients, due to their delayed N20 latency, together with short LLR latency (*figure 8C*).

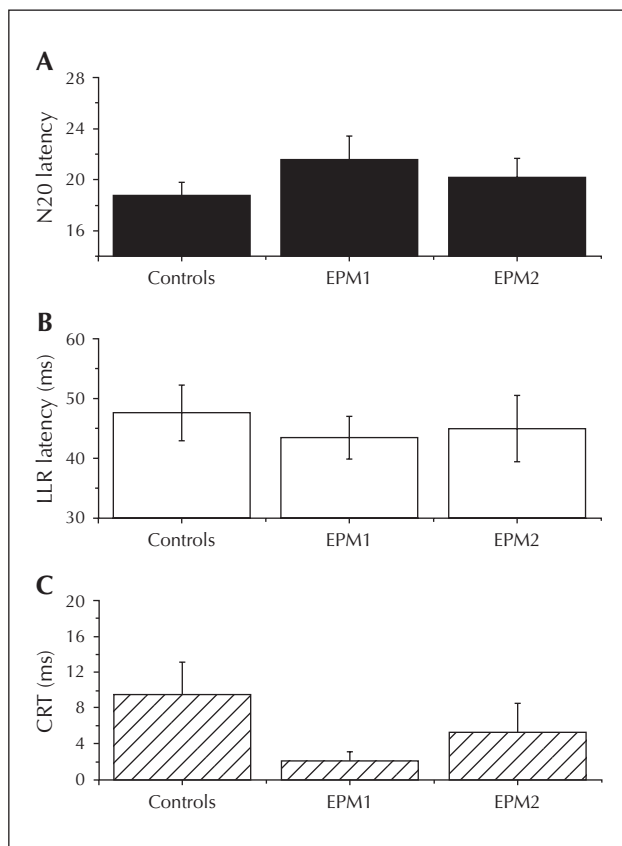


Figure 8. Graphic representation of N20 latency (A), LLR latency (B) and cortical relay time (C), comparing healthy controls with patients with Unverricht-Lundborg disease (EPM1) or Lafora body disease (EPM2).

Transcranial magnetic stimulation

Resting motor threshold is typically high in most PME patients, a finding that can be probably explained by the AED treatment (Reutens *et al.*, 1993; Manganotti *et al.*, 2001; Canafoglia *et al.*, 2004; Danner *et al.*, 2009, 2013). The high threshold means that, in some patients, MEPs cannot be produced. In EPM1 and EPM2 patients, MEP latencies may be either normal or slightly prolonged (Canafoglia *et al.*, 2004; Danner *et al.*, 2009), while for sialidoses they are typically prolonged due to delays in both peripheral and central conduction (Canafoglia *et al.*, 2011). The silent period (SP) tested in EPM1 patients is abnormally prolonged (Danner *et al.*, 2009). In patients with sialidosis type I, the SP was, conversely, reduced (Huang *et al.*, 2008).

Modulation of the motor evoked potentials (MEPs) by afferent somatosensory stimuli

The evaluation of MEP modulation by means of multimodal stimulation protocols suggests some clues about cortical excitability in different PMEs.

Both healthy subjects and PME patients had larger MEP amplitudes after conditioning stimuli (R2) to peripheral mixed nerves, in comparison to basal (non-conditioned) MEP amplitudes (R1); this finding was more evident at inter-stimulus intervals (ISIs), ranging from 30 to 80 ms (Reutens *et al.*, 1993; Cantello *et al.*, 1997; Canafoglia *et al.*, 2004). Digital stimulation markedly facilitated conditioned motor evoked potentials at ISIs ranging from 25 to 40 ms in all patients. This pattern was significantly different from the inhibition observed in controls at the same ISIs (Manganotti *et al.*, 2001).

A comparative analysis of the results obtained in EPM1 and EPM2 patients (Canafoglia *et al.*, 2004) indicated that the degree and time course of MEP facilitation was different for EPM1 compared to EPM2. Indeed, EPM2 patients maintained a significantly higher MEP facilitation at ISIs of between 40 and 60 ms, whereas EPM1 patients had a higher facilitation only at an ISI of between 20 and 40 ms (figure 9).

Intracortical inhibition and facilitation tested with paired magnetic pulses

In patients with cortical myoclonus, short interval intracortical inhibition (SICI) is generally reduced (Brown *et al.*, 1996; Manganotti *et al.*, 2001; Hanajima *et al.*, 2008). A comparative study in patients with EPM1 and EPM2 (Canafoglia *et al.*, 2010) indicated that both the EPM1 and EPM2 patients showed significantly less inhibition than the healthy subjects, with no difference between the two patient groups, with the exception of an ISI of 6 ms; at this ISI there was a significant enhanced inhibition in EPM2 with respect to EPM1 patients. Intracortical facilitation (ICF) was normal in EPM1 patients, while there was a significantly reduced facilitation in EPM2 patients at an ISI of 10 ms.

Data obtained in sporadic patients with sialidosis (Brown *et al.*, 1996; Manganotti *et al.*, 2001) and in a homogeneous series of patients with sialidosis type I (Huang *et al.*, 2008) indicated a reduced SICI. In those patients, ICF was normal.

Long interval intracortical inhibition (LICI) was generally impaired in patients with cortical myoclonus (Valzania *et al.*, 1999). However, a comparative study in patients (Canafoglia *et al.*, 2010) with EPM1 and EPM2 revealed significantly less inhibition in EPM2 patients with afferent stimuli (ISIs: 20 and 40 ms; ISIs: 30-80 ms).

Cortical plasticity

Cortical plasticity may be tested by means of repetitive TMS protocols. A study performed using a paired associative stimulation protocol indicated altered plasticity of the sensorimotor cortex in EPM1 patients. These patients exhibited an average decrease of 15 per cent in motor-evoked potential amplitudes, 30 minutes after

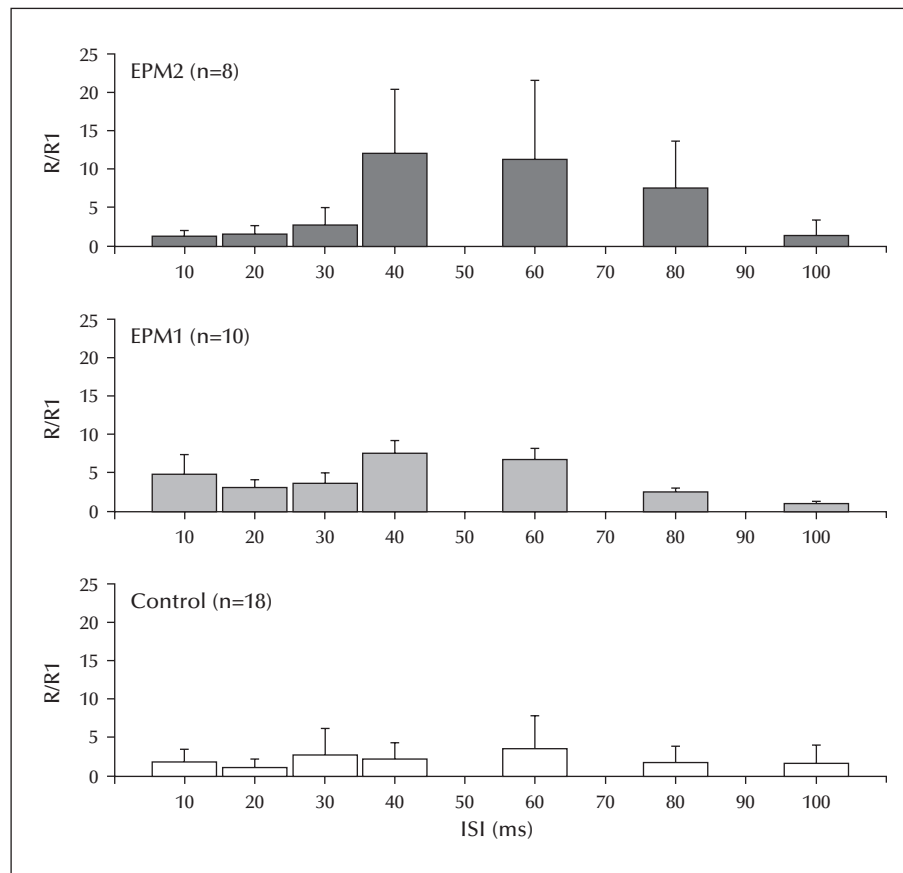


Figure 9. Graphic representation of interaction between peripheral somatosensory stimuli and transcranial magnetic stimulation (TMS). Note the different profiles between controls and both PME groups indicating the amplification of the effect of sensory stimulus on motor cortex excitability. Moreover, Lafora patients show a larger and long-lasting excitatory interaction.

paired associative stimulation, while in the control subjects, there was a significant increase (Danner *et al.*, 2011). Again, AEDs may have an important influence upon these results.

Conclusions

The neurophysiological evaluation of patients with different PMEs demonstrates the presence of peculiar features that, although not strictly distinctive, suggest different mechanisms underlying the generation of myoclonus. Most patients who were included in the study protocols were treated with multiple drugs, mainly AEDs, which may have influenced some results. However, since the treatment was relatively similar in the various PME syndromes, the differences observed actually suggest different dysfunctions affecting the circuitries sustaining myoclonic jerks.

In EPM1 patients, myoclonus was regularly induced by action and was associated with exaggerated LLRs and early facilitation of the motor cortex by afferent

stimuli. These findings, associated with the increased P25-N33 SSEP component, completely fit the definition of cortical reflex myoclonus (Shibasaki & Thompson, 2011). The finding of a short cortical relay time suggests the possibility of an alternative transcortical loop generating reflex myoclonus, passing through the thalamus directly to the motor cortex (Canafoglia *et al.*, 2004). This hypothesis also agrees with the finding in EPM1 patients, of an early facilitation of the motor cortex by conditioning stimuli (possibly following premature invasion of the motor cortex by somatosensory afferent volleys).

The findings obtained with paired TMS stimulation clearly indicated a defective inhibition, probably due to defective GABA circuitry (Canafoglia *et al.*, 2010), in line with the observation made in the CSTB mouse model, suggesting a prominent reduction in GABA-dependent inhibitory function in both the hippocampus and neocortex (Franceschetti *et al.*, 2007; Buzzi *et al.*, 2012).

The finding of abnormal cortical plasticity indicating defective sensorimotor integration may be associated

with the previously reported structural and physiological abnormalities of the primary motor cortex in EPM1 patients (Danner *et al.*, 2011).

In EPM2 patients, neurophysiological findings differ from those found in EPM1 patients, suggesting a more complex circuitry sustaining the severe myoclonic presentation. LLRs are more often within the normal range, corresponding to a less significant reflex myoclonus. Conversely, the prolonged late facilitation of motor cortex found by applying sensory stimuli followed by magnetic stimuli, and the enhanced middle latency SSEP components can, conversely, match a more complex circuitry, generating the high propensity of these patients to show spontaneous epileptic myoclonus. The profile of cortical excitability revealed by paired pulse with TMS, showing multiple abnormalities, not limited to short-time interaction or even late interaction, revealed by ICF and LICI, may support this interpretation, suggesting a more complex, hyperexcitable network and a more extensive defect of the inhibitory mechanisms, possibly related to GABA-B mediation (Ziemann, 2004).

Polyglucosan accumulation in dendrites typically occurring in this disorder (Chan *et al.*, 2004) may directly cause an imbalance between GABAergic and glutamatergic post-synaptic inputs to the dendrite tree or changes in the dendrite electrotonic properties capable of modifying the transfer of inputs to the neuronal soma.

In sialidoses, neurophysiological findings mainly overlap those observed in EPM1, since myoclonus prominently presents as a reflex phenomenon and as action-activated. The most characteristic feature is the rhythmic time course of action myoclonus, which substantially replaces the normal muscle contraction throughout the movement. In accordance with this finding, LLR mostly features repetitive components, probably reflecting pathological loops involving the motor cortex, leading to a reverberating circuitry and recurrence of jerks.

Although sialidoses are lysosomal disorders leading to neuron storage which may in turn lead to cell death, only mild spongiosis and lipofuscin granules have been found in the neocortical structures of patients with sialidosis (Allegranza *et al.*, 1989). Thus, the hyperexcitability sustaining myoclonus in this disorder probably arises from subtle circuitry rearrangements rather than from massive cell loss. The circuitry rearrangement resulting from sialidoses may lead to extreme synchronization of the neuronal pools sustaining action-activated jerks.

In conclusion, although PME presentations always include findings reflecting neocortical hyperexcitability, the recognition of subtle differences in diverse genetic disorders may help in designing the diagnostic

work-up. Moreover, these differences suggest peculiar dysfunctions in the neuronal network responsible for myoclonus. At present, we can only hypothesize that different modulating mechanisms derived from extra cortical regions (possibly subcortical nuclei and cerebellum) or due to complex cortico-cortical interaction, potentially lead to these different presentations.

In this article, we report evidence obtained in the two more common PME forms (EPM1 and EPM2) and for sialidoses, in which different neurophysiological features reflect the different clinical presentation and severity of stimulus reflex vs. spontaneous myoclonus. However, this certainly also occurs for other, rarer PMEs, which are often reported in single cases or minimal case series, preventing an analysis of the comparability of the results obtained by the applied examination protocols. The opportunity to share similar examination procedures could significantly promote better recognition of the specific phenotypes and could better address significant hypotheses to explain genotype-phenotype relationships. □

Disclosures.

The authors have no conflict of interest to disclose.

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Unverricht-Lundborg disease

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ABSTRACT – We first review the clinical presentation and current therapeutic approaches available for treating Unverricht-Lundborg disease (ULD), a progressive myoclonus epilepsy. Next, we describe the identification of disease causing mutations in the gene encoding *cystatin B* (*CSTB*). A *Cstb*-deficient mouse model, which recapitulates the key features of ULD including myoclonic seizures, ataxia, and neuronal loss, was generated to shed light on the mechanisms contributing to disease pathophysiology. Studies with this model have elucidated the diverse biological roles for *Cstb* from functioning as a protease inhibitor, to regulating glial activation, oxidative stress, serotonergic neurotransmission, and hyperexcitability. These findings set the stage for future studies that may open avenues to improved therapeutic approaches.

Key words: Unverricht-Lundborg, EPM1, progressive myoclonus epilepsy

Unverricht-Lundborg disease (ULD) (EPM1) is the 'purest' type of progressive myoclonus epilepsy (PME) (Minassian *et al.*, 2016), with only minor symptoms associated with epileptic seizures and myoclonus (Berkovic *et al.*, 1986 ; Marseille Consensus and Group, 1990). H. Unverricht and H. Lundborg described the condition in 1891 (in Estonia) and 1904 (in Sweden), respectively (Unverricht,

1891; Lundborg, 1903). The recognition of ULD is still lacking in areas of low prevalence, moreover, the distribution of ULD around the world is highly variable which is due to several factors: i) unequal distribution of the genetic defect between different populations; ii) an autosomal recessive mode of inheritance, which implies higher prevalence in areas and cultures with high consanguinity

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or a founder effect; and iii) variable availability of modern diagnostic procedures, including molecular biological techniques. Although ULD remains uncommon, it has been increasingly diagnosed among patients with drug-resistant myoclonic epilepsy, as shown, for instance, in Holland (de Haan *et al.*, 2004). However, whereas the most common mutation in ULD affects the cystatin B gene (*CSTB*), no evidence for a role of this gene was found in sporadic or familial cases of juvenile myoclonic epilepsy (JME) (Mumoli *et al.*, 2015). The availability of molecular genetic diagnostic techniques in developed countries has certainly helped.

ULD is still considered a severe and disabling condition, however, recent years have witnessed a shift for ULD, from a very severe, even lethal entity, towards a comparatively bearable disability with little impact, for instance, on life expectancy. Aggravation by phenytoin (PHT) (Eldridge *et al.*, 1983) has contributed to poor prognosis of ULD in countries where PHT is heavily used and titrated to high doses for resistant seizures. The aim of this article is to review the clinical features, prognosis, management, and pathogenesis of ULD.

Clinical characteristics

Onset occurs in late childhood and early adolescence, peaking at around age 12–13. Sex distribution is equal. *Generalized tonic-clonic seizures* (GTCSs) are usually grounds for an initial referral. They occur typically at awakening or during sleep. At onset, GTCS cannot be easily differentiated from those observed in JME, and may occur without prior myoclonic jerks. However, with disease progression, they may evolve into *cascade seizures* (Kyllerman *et al.*, 1991), characterized by a build-up of increasingly intense and violent myoclonic jerks, culminating into a short GTCS; some patients do not report clear consciousness and more or less retain normal contact during this type of seizure. Often, patients experience GTCS or major seizures after a period of progressive increase in myoclonus and subsequently experience less myoclonus, with a decreased risk of major seizures for a period that can last days to weeks; this periodicity has already been described by Unverricht and reported for other PME (Ferlazzo *et al.*, 2009; Vanni *et al.*, 2014). Besides awakening, there are no clear triggering factors for GTCSs. Of course, modern drug therapy has contributed to the control of these major seizures, which also seem to decrease spontaneously with advancing age (see below).

Myoclonus is already present in the very early stages, with diffuse myoclonic jerks that predominate at

awakening. Over a relatively short period (months to a few years), and in spite of AEDs treatment, myoclonus becomes movement-related and increases with stress. It also becomes physically challenging for patients, e.g. patients may fear using stairs or physical strain, and bilateral, violent myoclonus becomes less apparent (unless the patient is challenged or stressed), and partial, erratic myoclonus predominates. Reflex myoclonus, triggered by sensory stimulation (touch, light, etc.), initiation of movement, and surprise, is a prominent feature in some patients. Remarkably, myoclonus is less severe or even absent at rest or during sleep. In the early stage of ULD and in patients who are insufficiently treated, myoclonus may fluctuate within the same day (with maximal myoclonus in the morning and in the latter part of the day, when patients are tired) or with an interval of a number of days, sometimes with marked periodicity. These features, however, become less prominent over the subsequent years and with effective drug therapy. Contrary to major seizures, myoclonus does not spontaneously abate in the long term, but may even slowly increase in some adult or middle-aged patients. In the more severely affected patients, myoclonus causes major disability; a wheelchair may have to be used and feeding becomes problematic and slow, especially with the intake of liquids (using a straw may be helpful).

Absence, simple motor, or complex focal seizures may occur, but few video-EEG reports of these seizure types have been documented (Kälviäinen *et al.*, 2008).

Photosensitivity, detected in the EEG laboratory for nearly all patients in the early years of the disease, does not pose major problems in daily life, and tends to abate after 5 to 10 years of disease (Ferlazzo *et al.*, 2007).

Associated neurological symptoms are few. Ataxia, impaired walking, and instability upon standing up are associated with the severity of myoclonus. In our experience, patients cease to be ataxic when myoclonus is fully controlled.

Cognitive impairment may be absent or vary from mild to moderate. Many of our patients reached university level and are qualified professionals. However, whenever possible, ULD patients should avoid professions that involve significant physical activity or fine and precise handling of small objects. Neuropsychological impairment may slightly progress over the years. Early evidence points to a 10-point loss of IQ over ten years (Koskiniemi *et al.*, 1974), and the presence of short-term memory, attention, and executive function impairment is particularly apparent, based on recent studies (Ferlazzo *et al.*, 2009; Giovagnoli *et al.*, 2009).

Psychiatric comorbidities are frequent in ULD but have not been systematically studied. Suicidal behaviour, with loss of interest in life and neglect of therapy,

is a common behaviour, however, only one in 60 of our patients committed suicide. Depression is common and was found in six of eight patients by Chew *et al.* (2008).

Prognosis

The long-term evolution is characterized by limited progression after the first five to ten years (Magaudda *et al.*, 2006), with a varying but fairly stable level of disability thereafter; the outcome in adults ranges from independent active life with minimal impairment to severe disability and wheelchair-bound or even bedridden patients. Early death has a comparatively low incidence and may be due to suicide or accidents, but also to SUDEP, the latter mostly in 'undertreated' patients, in relation to persisting convulsive seizures (Khiari *et al.*, 2009).

Management: diagnosis and genetic counselling

Given the absence of specific clinical or pathological markers, a confirmation of diagnosis of ULD based on genotyping is necessary. Diagnosis used to rely on a combination of positive signs and on the absence of the more specific symptoms and markers of other PME's, but nowadays can be considered to depend entirely on molecular biological techniques. It is our opinion that this procedure, which remains costly, should be justified by solid electro-clinical evidence, and should not be performed, together with a variety of tests, to screen all possible genetic aetiologies in poorly assessed subjects with epilepsy and myoclonus. The diagnosis of ULD is based on three levels of evidence:

- 1. *Clinical evidence*: a combination of history-taking (age at onset, familial and ethnic background, circumstances, and aspects and progression of seizures and myoclonus), examination (including cognitive assessment showing the absence of major and progressive cognitive impairment, and exclusion of associated manifestations such as sensory deficits), and video documentation of myoclonus.
- 2. *Complementary evidence*: based on a thorough evaluation of the EEG, polygraphic EEG, and video-EEG recordings (with assessment of changes over time); neurophysiological studies may also help distinguish ULD from other adolescent-onset PME's, e.g. Lafora disease (in which EEG changes are much more spectacular) or juvenile ceroid-lipofuscinosis (with prominent single-flash responses on EEG). A lack of any of the signs and symptoms associated with other

PME's is likely to indicate a diagnosis of ULD, however, other less common PME's that were recently identified among genetically negative 'ULD' patients may complicate the diagnosis (Franceschetti *et al.*, 2014; Muona *et al.*, 2015)

- 3. *Confirmation of diagnosis*: provided nowadays by the demonstration of a pathogenic mutation in both alleles of the *EPM1* gene.

The diagnosis of ULD can be made, or suspected, in various clinical settings. In particular, the most common clinical situation is when idiopathic generalized epilepsy (IGE) or JME has been diagnosed, with a reassessment of the patient's situation because of an unusual evolution or apparent drug resistance; the clinical work-up should help exclude the possibility of aggravation of IGE by inappropriate AEDs, which may result in a pseudo-PME phenotype.

In some patients with a very likely diagnosis of ULD, the genetic testing of *EMP1* remains negative. Such patients should be discussed with clinicians having experience with PME's; if the clinical work-up has been thorough, there is usually no need to screen the genes associated with the very typical PME's which manifest within the same age group (LD, juvenile or adult neuronal ceroid lipofuscinosis, MERRF, DRPLA; etc), although recently described genetic defects in *SCARB2/LIMP2*, *KCNC1* or *PRICKLE* might be considered. When the molecular defect remains elusive, and the general condition is very much compatible with ULD, the patient should be diagnosed with ULD (or 'ULD-like PME') and managed as other patients with ULD, with reassessment of the case at two- to five-year intervals by specialized teams.

The diagnosis of ULD should not be given to the family before a final and definitive confirmation, because of the multiple psychological problems that surround the diagnosis of this genetic disease. However, it should be given to the patient and caregivers when confirmed, together with as much information as possible about the condition and its prognosis. Patient organizations can be contacted, and quality information is available on the web. The patients should be encouraged to learn more about the condition and to follow the scientific progress on ULD.

The consequences of a diagnosis of ULD for the family of the proband should not be underestimated. Regarding the prospects, time should be devoted to the change in the patient's and family's lives (see below), but also to counselling on the following topics:

The understandable, expected feelings of guilt and resentment associated with the diagnosis of a genetically transmitted condition should be alleviated, with the use of a few simple statements: inheritance is bilateral (*i.e.* the disease is not transmitted specifically from either the father or mother, but potentially

from both), the disease results from a very uncommon co-occurrence of abnormal genes (even in consanguineous marriages), and subjects with a single abnormal gene will not have the disease.

One major concern is the possible occurrence of ULD in other family members. The risk can be practically excluded in older, fully asymptomatic siblings, but cannot be excluded in *younger* asymptomatic siblings, *a fortiori* in yet unborn siblings, if the parents are young enough to have other children. Whether siblings, especially younger ones, should be referred for molecular screening is still debatable. For *presymptomatic* ULD cases, there is no reliable, recommended prophylactic treatment to prevent or delay the appearance and progression of symptoms, but this may change in the near future. Having provided all the relevant information, the physician should come to an agreement with the family and obtain their informed consent for all the possible procedures performed for non-affected family members. Concerning future pregnancies for the parents of a patient with ULD, the risks are easy to explain (the risk of ULD is 25%).

The risk of carrying an abnormal gene and transmitting the condition to other generations is also a major concern in families with ULD. Several of our ULD patients have children but none are affected; following medical advice, they had married 'outside the family', an unusual option in some ethnic contexts. Their siblings, or parents (when wanting children with another spouse), as well as other collaterals, may benefit from molecular screening in order to assess the presence or absence of the pathogenic gene found in the proband. An important aspect of diagnosis and genetic counselling is financial and the conditions of insurance and reimbursement (or, more basically, the availability of procedures, tests, and medications) differ greatly between countries and social systems. The families should always be informed about the costs involved; the search for a mutation is expensive, but simple screening for a known mutation is less costly. Similarly, the regulations covering genetic diagnosis, screening, and counselling may differ between countries, and clinicians should always conform to local laws. Obtaining informed consent for the successive steps of diagnostic and screening procedures is a minimum.

Management: medical treatment

This type of PME is characterized by two important features:

- The severity of the condition varies greatly between subjects, even within sibships, and a significant proportion of patients will be able to create a family and lead normal, productive lives with normal or

adapted employment, while a significant minority will be severely disabled and dependent upon their family or have an institutionalized life. In a previously published series of 20 ULD patients followed for more than 20 years (Magaudda *et al.*, 2006), eight lived autonomously, six had a family with children, six were normally employed, and seven were dependent on others, including two who were wheelchair-bound. In the largest cross-sectional series (77 subjects) evaluated using modern methodology in Finland, one third of the patients had mild myoclonus, one third had moderate myoclonus, and one third were wheelchair-bound due to severe myoclonus (Koskenkorva *et al.*, 2009).

- The disease has limited progression. Over several (up to 10) years following clinical onset, major seizures and photosensitivity disappear in most patients, while myoclonus stabilizes or progresses only minimally (Magaudda *et al.*, 2006). Thus, a reasonably accurate prognosis can be made fairly early during the course of the condition, 5 to 10 years after onset.

AEDs and piracetam (PIR), a more specific antemyoclonic agent, alleviate the burden of seizures and myoclonus, and their effect is felt throughout the course of the disease; unfortunately, this effect is partial in some cases, as medication does not influence the natural course of the disease. Patients will usually receive an AED after the first GTCS, typically valproic acid (VPA). VPA is normally effective in suppressing, for some time, most GTCS, photosensitivity, and some of the myoclonus. Other AEDs can be used at this stage (Genton *et al.*, 2012); lamotrigine (LTG) is not an AED of first choice in the context of a myoclonic epilepsy, and has been shown to aggravate myoclonus in some patients with ULD (Genton *et al.*, 2006); phenobarbital (PB) and primidone are effective, but produce cognitive side effects on top of the complications attributed to the condition; levetiracetam (LEV) is increasingly used early on in adolescents with IGE, hence in ULD cases, even before confirmation of the diagnosis. Other useful drugs include topiramate (TPM) and zonisamide (ZNS), both with marked antemyoclonic effects. Additional relief can be obtained, often transiently, with benzodiazepines (BZD). The latter (usually clobazam, clonazepam, or diazepam) should be used with care because of a marked initial effect followed by rapid tolerance.

For ULD, the paradoxical aggravating effect of some AEDs may be difficult to assess. There is no evidence that carbamazepine (CBZ), oxcarbazepine (OXC), phenytoin (PHT), eslicarbazepine, gabapentin, pregabalin, vigabatrin or lacosamide are of any benefit. Often, withdrawal of one of these AEDs (especially CBZ or OXC) will bring some relief.

For established ULD, AED treatment leads to polytherapy with a combination of several of the drugs quoted

above (with the exclusion of LTG); the commonly used combinations are VPA+LEV or TPM or ZNS, with an additional BZD (a three- to five-drug combination is quite usual); one can switch between different BZDs in the event of tolerance. In case of transient worsening, with intense myoclonus and serial seizures, there should be no abrupt change in the usual regimen (except for the interruption of a potentially aggravating AED), and IV BZD should be used, as well as, for a limited period, IV PB or PHT.

In practice, some patients will fare reasonably well with a limited drug regimen, while others will remain severely disabled, especially with intractable myoclonus, and will have a much heavier pharmacological load. In such patients, specific therapeutic approaches can be discussed:

- vagal nerve stimulation has been tried with success in some individuals (Smith *et al.*, 2000), including three in our personal experience, but the benefit is limited;
- deep brain stimulation has also been used in PME cases, including some patients with ULD; combined subthalamic and thalamic high-frequency stimulation has brought some relief, especially in the least severely affected cases (Wille *et al.*, 2011). Our personal experience with three patients with a severe form of ULD, who received bipallidal stimulation (as used for dystonia and myoclonic dystonia), was disappointing (Crespel, personal communication).

Management: social support

For ULD, a lifelong condition, social support is at least as important as medical treatment. Psychological support can be provided by patient organizations; one such organization exists in France specifically devoted to ULD, but epilepsy organizations in other countries may offer support. The individual patient should also receive professional psychological support whenever necessary throughout the course of ULD. Physical therapy aims at maintaining a good overall muscular condition and at preserving the ability to walk for the more severely affected patients.

Small amounts of alcohol may temporarily relieve myoclonus (Genton and Guerrini, 1990), but patients should be warned about tolerance, and chronic abuse of alcohol clearly worsens the condition. Photosensitivity, with increased myoclonic jerks, usually abates after several years and is seldom a problem in daily life (patients may watch television or use a computer). Most patients experience increased myoclonus (and an increased risk of major seizures) in the morning, especially after abrupt/sudden awakening, during the active phase of the condition, and should be advised to take their time before getting up. The effect of sleep

deprivation has not been well documented, but anecdotal evidence shows that it may increase myoclonus and seizures, and should be avoided.

At the onset of seizures, the patient is typically in primary or secondary school and experiencing some difficulties with academic requirements, however, it is usually possible to maintain normal schooling. It is useful to discuss with the parents future professional orientation, in light of the possible disability associated with ULD. In most families, the patient will remain at home, but the environment may need to be adapted, e.g. avoidance of the use of stairs or ensuring the proximity of a bathroom. Specialized institutions are used for the most severely disabled patients, or for periods to promote education about the condition.

Re-evaluation at a specialized neurological department can be organized at 6- or 12-month intervals, with acute admissions in case of complications, often due to intercurrent diseases (e.g. febrile infections). There should be a link between the reference specialized epilepsy team and the local caregiving structure in case of intercurrent health problems, ranging from dental care to surgical procedures. Long-term psychological support is not always necessary, and additional medication (e.g. antidepressants) should be discussed with the reference neurologist. Pregnancies have occurred without major complications in less (or moderately) severe women with ULD.

Adult ULD patients usually reach a level of disability and dependency that will remain fairly stable. As stated above, possibilities range from a fully normal life with minimal impairment and monotherapy (usually with VPA) to institutionalized care.

Pathogenesis of ULD: the role of oxidative stress

Oxidative stress is associated with many neurological conditions, including epilepsies (Chong *et al.*, 2005; Kunz, 2002). Several case reports have suggested that antioxidant therapies, including N-acetylcysteine, may alleviate ULD symptoms (Hurd *et al.*, 1996; Selwa, 1999; Ben-Menachem *et al.*, 2000; Edwards *et al.*, 2002). Consistent with the model in which redox homeostasis may be disrupted in ULD, redox analyses in the *Cstb* knockout mouse have revealed that oxidative damage in the cerebellum contributes to the pathogenesis of murine ULD (Lehtinen *et al.*, 2009). Deregulation of antioxidants, including superoxide dismutase (SOD) and the antioxidant glutathione (GSH), are disrupted in the cerebella of ULD mice. The ratio of oxidized to reduced glutathione (GSSG:GSH), a hallmark of oxidative damage, is also increased in ULD mice. Detailed analyses

have uncovered progressive lipid peroxidation in the cerebella of ULD mice. Lipid peroxidation accelerates with age, beginning at baseline in young adult mice (two months of age) and increasing nearly five-fold in six-month-old mice, compared to controls. Evidence for cellular adaptation to accumulating lipid peroxidation was also observed as the activity of glutathione peroxidase, an enzyme that reduces lipid hydroperoxides, increases in ULD mice, compared to controls (Lehtinen *et al.*, 2009).

A hallmark of the ULD mouse model is a progressive loss of cerebellar granule neurons (Pennacchio *et al.*, 1998). Cerebellar granule neurons can be isolated, cultured, and genetically manipulated *in vitro* (Lehtinen *et al.*, 2006), thus providing a powerful experimental tool for investigating the cellular role of *Cstb*. In these types of *in vitro* studies, healthy neurons upregulate *Cstb* transcription when challenged by peroxide-induced oxidative stress (Lehtinen *et al.*, 2009). Most patients with classic autosomal recessive ULD harbour an unstable dodecamer repeat expansion (5'-CCC-CGC-CCC-GCG-3') in at least one allele in the *Cstb* promoter region (Pennacchio *et al.*, 1996; Lafreniere *et al.*, 1997; Lalioti *et al.*, 1997; Joensuu *et al.*, 2008). Introducing a similar expansion into neurons prevents *Cstb* upregulation during oxidative stress, suggesting that the inability to upregulate *Cstb* expression impairs normal neuronal responses to oxidative stress. Consistent with this model, reducing *Cstb* expression by RNA interference (RNAi) or genetic deletion in knockout mice sensitizes neurons to oxidative stress-induced death (Lehtinen *et al.*, 2009).

As part of its downstream signalling, *Cstb* inhibits the lysosomal protease cathepsin B (Turk and Bode, 1991). Cathepsin B over-expression promotes neuronal death, and cathepsin B activity is upregulated in both *Cstb*-deficient cells, as well as in ULD patients with *CSTB* mutations (Rinne *et al.*, 2002). Importantly, the neuronal cell death that occurs upon *Cstb* loss can be inhibited by a concomitant decrease in cathepsin B both *in vitro* (Lehtinen *et al.*, 2009) and *in vivo* (Houseweart *et al.*, 2003), supporting the model that homeostasis related to *Cstb*-cathepsin B signalling regulates neuronal survival in the mouse model of ULD.

The identification of a role for *Cstb* in oxidative stress responses in neurons, together with reports that antioxidant therapies alleviate some symptoms of ULD (Ben-Menachem *et al.*, 2000; Edwards *et al.*, 2002; Hurd *et al.*, 1996; Selwa, 1999), provide us with a better understanding of ULD disease pathophysiology. However, whether oxidative damage is a primary cause of disease onset and progression remains to be elucidated. It will be important in future studies to further investigate *Cstb*-cathepsin B signalling, as well as the downstream consequences of selective oxidative damage

to cerebellar lipids. These types of studies may shed light on new approaches to selectively tailor therapies for ULD.

Pathogenesis of ULD: disruption in serotonin metabolism

5-hydroxytryptamine (5HT) metabolism may contribute to ULD pathogenesis. Indeed, an early report identified decreased availability of L-tryptophan (TRP) and its metabolites in cerebrospinal fluid and blood of ULD patients (Pranzatelli *et al.*, 1995). These observations supported the hypothesis that insufficient serotonergic neurotransmission contributes to ULD. Studies using *Cstb*-deficient mice failed to identify changes in serum TRP concentrations. However, serum levels of 5HT and 5-hydroxyindole acetic acid (5HIAA), an intermediate metabolite of 5HT, tend to be reduced (Vaarmann *et al.*, 2006). These data suggest that the observed disruption in 5HT metabolism in ULD patients may be causally related to *Cstb* deficiency, rather than being affected by systemic TRP availability or drug therapy. In contrast to TRP concentrations in serum, the brains of *Cstb* knockout mice have increased levels of TRP and its metabolites (Vaarmann *et al.*, 2006). Kynurenine, the first metabolic intermediate in the TRP kynurenine pathway, is also elevated in the *Cstb* knockout cerebellum. The myoclonus in ULD is believed to arise from impaired intracortical inhibition leading to secondary hyperexcitability of the motor cortex (Franceschetti *et al.*, 2007). Interestingly, *Cstb*-deficient mice exhibit significantly higher levels of 5HT and 5HIAA in the cerebral cortex and cerebellum (Vaarmann *et al.*, 2006), the structures where the greatest cellular atrophy and glial appearance have been described (Pennacchio *et al.*, 1998; Shannon *et al.*, 2002). The enhanced serotonergic neurotransmission may result from the loss of GABAergic interneurons, which play a critical role in controlling the serotonergic network in these regions, and are damaged in the *Cstb* knockout cortex (Franceschetti *et al.*, 2007; Buzzi *et al.*, 2012). While a direct relationship between altered serotonergic neurotransmission and ULD remains to be elucidated, these data suggest that disrupted serotonergic transmission is associated with the disease.

Pathogenesis of ULD: loss of *Cstb* contributes to defective inhibitory neurotransmission

In addition to its role in redox homeostasis and 5HT metabolism, *Cstb* protects a cell from endogenous proteases that have the potential to damage neuronal circuitry. *Cstb* deficiency leads to

hyperexcitability and impaired neuronal function of cortical neuronal networks in both ULD patients and in *Cstb* knockout mice. Several studies have investigated the molecular mechanisms underlying neuronal hyperexcitability by pairing analyses of neurodegeneration with network changes occurring in the hippocampus following kainate treatment, a pro-convulsant that triggers excitotoxicity and epileptic events (Arundine *et al.*, 2003). Early *in vitro* hippocampal slice experiments suggested that during kainate perfusion, afferent synaptic activation evokes multiple population spikes in both *Cstb*-deficient and wild-type control mice (Franceschetti *et al.*, 2007). The appearance of such hyperexcitable responses coincides with a rapid decline in the amplitude of the evoked field potentials in slices from *Cstb*-deficient mice. Spontaneous epileptic discharges (SEDs) occur in a subset of slices prepared from wild-type controls, with a delay of about 15 minutes following the onset of kainate perfusion, and persisting until the end of the kainate exposure. In contrast, in *Cstb*-deficient mice, SEDs begin within minutes of kainate perfusion, and progressively decrease in amplitude, ultimately disappearing along with the field responses evoked by electrical stimulation (Franceschetti *et al.*, 2007). To test if increased susceptibility to pro-convulsant agents can be recapitulated *in vivo*, *Cstb*-deficient and control mice received intraperitoneal injections of kainate (30 mg/kg), and their behaviour was recorded for two hours thereafter. Consistent with *in vitro* findings, in this paradigm, *Cstb*-deficient mice display an increased susceptibility to kainate-induced seizures, such that the latency to generalized seizure onset is reduced and the behavioural seizure scores (cumulative seizure score and seizure index) are increased (Franceschetti *et al.*, 2007). To investigate the extent of seizure-induced damage, brain damage was evaluated in *Cstb*-deficient mice and controls one day following kainate administration using several markers of neurodegeneration, including Fluoro-Jade B (Schmued and Hopkins, 2000). The degree of degeneration correlates with the severity of the seizures, and is higher in *Cstb*-deficient mice than in control mice (Franceschetti *et al.*, 2007). In addition, *Cstb*-deficient mice display more neurodegeneration compared to controls with identical seizure scores, suggesting that seizure-induced brain damage is more pronounced in animals lacking *Cstb*. Analyses using immunohistological markers reveal a loss of GABAergic hippocampal neurons, suggesting that the observed hyperexcitability may depend, at least in part, on defective GABAergic inhibition (Franceschetti *et al.*, 2007). To test if ULD is accompanied by a progressive loss of cerebral cortical GABAergic inhibition, cortical GABAergic neurotransmission was analyzed in *Cstb* knockout mice at various ages by histologically visualizing

GABAergic nerve terminals, by examining GABA release from isolated nerve terminals, and electrophysiologically evaluating cortical GABAergic tone. While the overall cell numbers are reduced in the *Cstb* knockout cortex, the loss of GABAergic interneurons is more pronounced compared to the general loss of neurons, indicating that GABA interneurons are selectively more vulnerable to kainate-induced damage compared to other neuronal subtypes (Buzzi *et al.*, 2012). A progressive reduction in the density of GABAergic nerve terminals (marked by VGAT staining) is also observed in the sensorimotor cortex of 4-, 8-, and 12-month-old *Cstb* knockout mice. One post-mortem ULD patient sample has shown a reduction in cortical thickness and a striking loss of VGAT-labelled GABAergic nerve terminals (Buzzi *et al.*, 2012). Experiments performed in mouse sensorimotor cortex using the paired-pulse paradigm, which is a stimulus protocol to test depression of the conditioned stimulus resulting primarily from GABAergic inhibition, show decreased inhibition in *Cstb* knockout mice at interpulse intervals in cortical layers II-III and V, compared to controls (Buzzi *et al.*, 2012). Perfusion with low concentrations of the GABA antagonist bicuculline, at 0.5 μ M, a concentration suitable for slightly reducing GABAergic neurotransmission, results in an amplification of the field responses in cortical layers II-III and V of *Cstb* knockout brain slices (Buzzi *et al.*, 2012). Bicuculline leads to a pronounced decrease in depression profile in the paired-pulse protocol in *Cstb* knockout mice, while only minimally affecting control mice (Buzzi *et al.*, 2012). These effects result from the reduction of early GABAergic inhibition occurring simultaneously with the postsynaptic excitatory potential. Taken together, these data support the model that *Cstb* deficiency increases the susceptibility to seizures and to seizure-induced cell death. *In vitro*, hippocampal slices from *Cstb* knockout mice are hyperexcitable, and when perfused with kainate, display precocious and pronounced epileptic-like responses that couple with an early impairment in cellular function. *In vivo*, *Cstb* knockout mice display increased susceptibility to kainate-induced seizures and develop enhanced seizure-induced cell damage with higher degrees of neurodegeneration. The observation of decreased hippocampal GABAergic interneurons suggests that these neuronal subtypes are especially prone to cell damage resulting from *Cstb* loss. The reduction in GABAergic synaptic transmission observed in the sensorimotor cortex implicates a reduction in GABAergic synaptic transmission in ULD. Together, these findings support the model that one key factor contributing to the pathophysiology of ULD is the progressive loss of cortical GABAergic signalling which, with time, leads to hyperexcitability, myoclonus, and seizures.

ULD pathogenesis: precocious microglial activation

Glial activation, and particularly microglial activation, contributes to the mechanisms underlying brain pathologies including neuronal ceroid lipofuscinoses which also display PME (Cooper, 2010). In an early study using aged mice (16–20 months old), GFAP-positive astrocytes were reported to be more abundant in *Cstb* knockout mice than in controls, especially in the hippocampus (Shannon *et al.*, 2002). A recent study including both pre-symptomatic (P14) and symptomatic mice (>one month old), suggests that precocious glial activation is a key mechanism contributing to the pathogenesis of ULD (Tegelberg *et al.*, 2012; Okuneva *et al.*, 2015). Systematic histological analyses using an unbiased stereological approach revealed early and localized glial activation in the brain, as well as in the thalamocortical system. Microglial activation entailed the expression of p-p38 MAPK, a marker of inflammation. While the proportion of pro-inflammatory M1 and anti-inflammatory M2 microglia favours the M2 type earlier in development, the ratio shifts in favour of the M1 type by P30 (Joensuu *et al.*, 2014; Okuneva *et al.*, 2015). The observed microglial activation precedes the onset of myoclonus, and is followed by gliosis and neuronal loss. Interestingly, active microglia undergo morphological changes during ULD disease progression, from that of phagocytic brain macrophages in young animals, to thickened branch processes in older animals (Tegelberg *et al.*, 2012). Consistent with a requirement for microglial activation during disease progression, neuronal loss was not observed in brain regions lacking glial activation (e.g. thalamic relay nuclei). These findings are consistent with previous studies suggesting that ULD is a neurodegenerative disease (Pennacchio *et al.*, 1998; Shannon *et al.*, 2002). Indeed, recent approaches using magnetic resonance imaging and diffusion tensor imaging have uncovered white matter degeneration in *Cstb*-deficient mice (Manninen *et al.*, 2014), as well as in patients (Manninen *et al.*, 2015). Taken together, these findings reveal the timing and progression of pathological events in the *Cstb*-deficient mouse brain, highlighting the potential role of glial activation during the initial stages of ULD.

Conclusion

Although nowadays ULD can be treated effectively (albeit only symptomatically) which has led to reduced severity, patients may experience significant disability. A precise molecular diagnostic technique is available. Major progress can be expected in the near future, as elucidation of the mechanisms causing

seizures, myoclonus, and associated symptoms (which are mild and mainly cognitive) are likely to bring about pathogenetically-oriented treatment for ULD.

Genetic deletion of *Cstb* in the mouse has provided a powerful tool for modelling ULD in the laboratory. These mice display the triad of symptoms associated with ULD, including myoclonus, ataxia, and neuronal loss. Because *Cstb* is ubiquitously expressed and functions in healthy cells as an inhibitor of the cathepsin family of proteases, it is not surprising that loss of *Cstb* affects a broad range of cellular biological functions, including neuronal death, redox homeostasis, hyperexcitability, and glial activation. The studies reviewed in this article suggest that glial activation may be one of the earliest events contributing to *Cstb*-deficient brain pathology and is accompanied by oxidative stress, neuronal death, aberrant serotonin regulation, and hyperexcitability. Because *Cstb* is ubiquitously expressed, systemic knockout of *Cstb* results in the loss of *Cstb* in all cells of the body. Therefore, a limitation of the present ULD mouse model is the inability to reveal whether the observed phenotypes arise from primary defects in a specific cell type (i.e. neurons vs. glial cells). It is possible that some observed phenotypes are secondary to defects originating in neighbouring cells.

Collectively, these findings lay the foundation for future studies, which, by harnessing the potential of the ULD mouse model, should improve our understanding of the pathophysiology of ULD and open avenues for tailoring new therapeutic approaches. □

Disclosures.

None of the authors have any conflict of interest to disclose.

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Lafora disease

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ABSTRACT – Lafora disease (LD) is an autosomal recessive progressive myoclonus epilepsy due to mutations in the EPM2A (laforin) and EPM2B (malin) genes, with no substantial genotype-phenotype differences between the two. Founder effects and recurrent mutations are common, and mostly isolated to specific ethnic groups and/or geographical locations. Pathologically, LD is characterized by distinctive polyglucosans, which are formations of abnormal glycogen. Polyglucosans, or Lafora bodies (LB) are typically found in the brain, periportal hepatocytes of the liver, skeletal and cardiac myocytes, and in the eccrine duct and apocrine myoepithelial cells of sweat glands. Mouse models of the disease and other naturally occurring animal models have similar pathology and phenotype. Hypotheses of LB formation remain controversial, with compelling evidence and caveats for each hypothesis. However, it is clear that the laforin and malin functions regulating glycogen structure are key.

With the exception of a few missense mutations LD is clinically homogeneous, with onset in adolescence. Symptoms begin with seizures, and neurological decline follows soon after. The disease course is progressive and fatal, with death occurring within 10 years of onset. Antiepileptic drugs are mostly non-effective, with none having a major influence on the progression of cognitive and behavioral symptoms. Diagnosis and genetic counseling are important aspects of LD, and social support is essential in disease management.

Future therapeutics for LD will revolve around the pathogenesis of the disease. Currently, efforts at identifying compounds or approaches to reduce brain glycogen synthesis appear to be highly promising.

Key words: Lafora, laforin, malin, glycogen phosphatase, ubiquitin, EPM2A, EPM2B, progressive myoclonus epilepsies

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Lafora disease (LD, OMIM# 254780) is an autosomal recessive progressive myoclonus epilepsy (PME) (Minassian *et al.*, 2016), first described in 1911. It is particularly

frequent in Mediterranean countries (Spain, Italy, France), Northern Africa, the Middle East, and in some regions of Southern India where a high rate of consanguinity is present

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(Minassian, 2001; Striano *et al.*, 2008). It can, nevertheless, be found in any population (Traoré *et al.*, 2009), and particularly, as expected, with consanguinity.

LD classically starts in adolescence in otherwise neurologically normal individuals, usually with action and stimulus-sensitive myoclonus, as well as tonic-clonic, absence, atonic, and visual seizures. Neuropsychiatric symptoms, such as behavioural changes, depression and apathy, are also often present. Initial symptoms are followed by rapidly progressing dementia, refractory status epilepticus, psychosis, cerebellar ataxia, dysarthria, mutism, and respiratory failure which lead to death within about a decade (Minassian, 2001; Striano *et al.*, 2008). LD is caused by mutations in the *EPM2A* or *EPM2B* (*NHLRC1*) genes, encoding the laforin dual specificity phosphatase and the malin ubiquitin E3 ligase, respectively, both involved in a complex and yet very incompletely understood pathway regulating glycogen metabolism (Minassian *et al.*, 1998; Chan *et al.*, 2003a, 2003b). To date, an additional gene, *PRDM8*, the mutation of which causes a variant of early childhood-onset phenotype in a single family, has been reported (Turnbull *et al.*, 2012).

A distinctive pathology characterizes LD. Cells of various types exhibit dense accumulations of malformed and insoluble glycogen molecules, known as polyglucosans, which differ from normal glycogen due to the fact that they lack the symmetric branching that allows glycogen to be soluble. These polyglucosan accumulations are called Lafora bodies (LBs) and are profuse in all brain regions and in the majority of neurons, specifically in their cell bodies and dendrites (Minassian, 2001; Striano *et al.*, 2008). Neuronal LBs localize in perikarya and dendrites but not in axons, possibly explaining the cortical hyperexcitability seen in LD. The neuropathology of LD patients and LD animal models is described in greater detail below.

Clinical features and diagnosis

The first symptoms of LD appear during late childhood or adolescence (range: 8-19 years; peak: 14-16 years). Characteristically, focal visual seizures are early manifestations and present as transient blindness, or simple or complex visual hallucinations. However, generalized seizure types, e.g. tonic-clonic, absence, or drop attacks, as well as myoclonus, occur soon after. The latter typically occurs at rest and increases with emotion, action, or photic stimulation. In many cases, the disease shows an insidious near-simultaneous, or closely consecutive, appearance of headaches, difficulties at school, myoclonic jerks, generalized seizures, and visual hallucinations (Minassian, 2001; Franceschetti *et al.*, 2006; Striano *et al.*, 2008). It is notable that not all visual hallucinations are epileptic in Lafora patients,

as some respond initially to antipsychotic, rather than antiepileptic, medications (Andrade *et al.*, 2005).

EEG abnormalities often precede clinical symptoms and initially consist of almost normal or slowed background (*figure 1A*) and generalized or focal paroxysmal activity (*figure 1B*), typically not accentuated by sleep. In particular, the occipital discharges on EEG, arising from a slowed posterior dominant rhythm are, in the proper clinical context, highly suggestive of the disease. Within a few years, a slowing of background activity becomes evident with frequent, superimposed bursts of diffuse epileptic discharges (*figure 1C*). In addition, positive or negative myoclonus (*figure 1D*) and marked photosensitivity (*figure 1E*) are prominent features.

Brain MRI is usually unremarkable at onset. In two reported cases (Jennesson *et al.*, 2010), [18]fluorodeoxyglucose positron emission tomography (FDG-PET) revealed posterior hypometabolism early during the evolution of the disease. Electrophysiological investigations (to examine jerk-locked averaging, somatosensory evoked potentials, C-reflex, and visual evoked potentials) can reveal aberrant integration of somatosensory stimuli and cortical hyperexcitability (i.e. giant evoked potentials). Visual evoked potentials may show increased latencies or absence of response. Other findings obtained by transcranial magnetic stimulation indicate a complex circuitry dysfunction, possibly involving both excitatory and inhibitory systems (Canafoglia *et al.*, 2010). A skin biopsy shows the presence of the characteristic periodic acid-Schiff (PAS)-positive glycogen-like intracellular inclusion bodies in the myoepithelial cells of the secretory acini of the apocrine sweat glands and in the eccrine and apocrine sweat duct cells (Andrade *et al.*, 2003; Lohi *et al.*, 2007). Electron microscopy confirms the presence of fibrillary accumulations, typical of polyglucosans. This diagnostic approach offers limited invasiveness and high sensitivity. Interpretation of the biopsy, however, requires expertise in distinguishing LB from normal polysaccharide contents of apocrine sweat glands, without which false-positive diagnosis is common. Genetic testing is crucial to confirm the diagnosis as it reveals mutations in the *EPM2A* or *EPM2B* gene in more than 95 per cent of patients (Minassian, 2001; Ganesh *et al.*, 2006; Striano *et al.*, 2008).

In the years following onset, symptoms of LD progress towards intractable action-sensitive and stimulus-sensitive myoclonus, refractory seizures, psychosis, ataxia, and dysarthria. As the disease progresses, the myoclonus remains asymmetric and segmental but becomes almost constant, and massive myoclonic jerks appear. At this stage, brain MRI may reveal mild cerebellar or cortical atrophy. Moreover, brain ¹H MR spectroscopy shows metabolic changes of the cerebellum, basal ganglia, and frontal cerebral

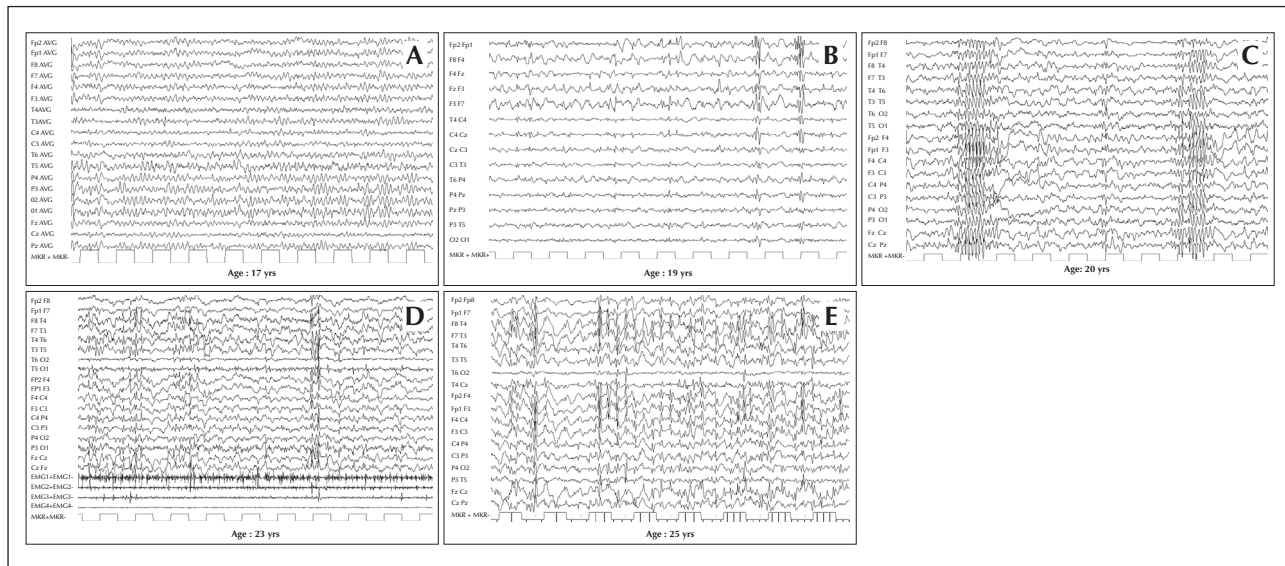


Figure 1. Progression of electroencephalographic (EEG) changes in a patient with Lafora disease. (A) At the time of disease onset (age 17 years), normal to slightly slowed background activity. (B) Two years later (age 19 years) EEG demonstrates asymmetric generalized spikes and polyspikes, maximum over the anterior regions on a slowed background. (C) At age 20 years, the occurrence of fast (4-6 cycles per second) spike-waves was concomitant with head drops. During the final stages of the disease, EEG recordings show long bursts of diffuse spike-waves and fast polyspikes associated with major volleys or massive myoclonic jerks (D), dramatically enhanced by photic stimulation at low frequency (E). *Modified from Striano et al., 2008.*

cortex (Villanueva *et al.*, 2006; Pichiecchio *et al.*, 2008). Subsequently, a rapidly progressive dementia with apraxia and visual loss soon appears. Speech becomes extremely difficult, and ataxia makes walking impossible. For many years, the patient struggles to maintain normal contact, with communication interrupted by extremely frequent myoclonic absence seizures (Minassian, 2001; Striano *et al.*, 2008). Patients finally become totally disabled and bed-bound. Death usually occurs within 10 years from the onset, often during status epilepticus with aspiration pneumonia.

Differential diagnosis

At onset, LD may present with a clinical picture resembling an idiopathic generalized epilepsy, such as juvenile myoclonic epilepsy (Janz syndrome). In the early stages, drug resistance and a slow background with early disrupted sleep patterns should lead to a suspicion of LD. In some patients, the diagnosis is reached only after substantial follow-up of the patient. However, the main differential diagnosis concerns four other forms of PME: Unverricht-Lundborg disease (EPM1), the neuronal ceroid lipofuscinoses, myoclonic epilepsy with ragged red fibers (MERRF), and sialidosis (Berkovic *et al.*, 1986; Minassian, 2001; Striano *et al.*, 2008) (table 1). PMEs are a group of inherited neurodegenerative disorders characterized by progressively worsening myoclonus and epilepsy,

variable neurological dysfunction (ataxia, dementia), and possible associated signs and symptoms. LD is one of the main teenage-onset PMEs. Age at onset, presenting symptoms, occurrence of occipital seizures, and the progressive and rapid course, together with the EEG features, are suggestive of LD, but the diagnosis is definitively confirmed by skin biopsy or genetic analysis. Unverricht-Lundborg disease (EPM1), caused by mutations of the cystatin B gene, is the closest differential. The age at onset is similar to that of LD. However, characteristically, action myoclonus and tonic-clonic seizures are more easily controlled. There is no specific pathology. Finally, cognitive impairment is not a distinctive symptom of Unverricht-Lundborg disease and is usually mild (Minassian, 2001; Striano *et al.*, 2008). Neuronal ceroid lipofuscinoses are heterogeneous conditions characterized by ceroid-lipofuscin lysosomal storage occurring in different organs, including the central nervous system and skin (Nascimento *et al.*, 2016). All NCLs exhibit ceroid lipofuscin accumulation, apart from LB, in various organs, including the skin. Most NCLs present in early childhood. A juvenile form of NCL (Batten's disease, involving the *CLN3* gene) may have onset late enough to overlap with that of LD, but this form has a prolonged initial period of visual loss, resulting from retinal degeneration and mild seizures, which LD patients typically do not have. Some cases of the infantile form (Santavuori-Haltia disease, involving the *CLN1* gene) present late as a result of particular mutations, and there is a very rare form of

Table 1. Distinguishing features of some of the more common inherited progressive myoclonus epilepsies.

| Progressive myoclonic epilepsy | Inheritance | Onset (years) | Suggestive clinical signs | Pathologic features | Gene(s) |
|---|-------------|---------------|--|--|--|
| Unverricht-Lundborg disease (EPM1) | AR | 6-15 | Slow progression; mild and late cerebellar impairment; late or absent dementia | None | <i>CSTB</i> |
| Lafora disease (EPM2) | AR | 6-19 | Visual symptoms | Polyglucosan inclusions (Lafora bodies) | <i>EPM2A</i> <i>EPM2B</i> |
| Myoclonic epilepsy with red ragged fibres (MERRF) | Maternal | Any age | Lactic acidosis | Ragged red fibres | <i>MTTK</i> (<i>tRNA^{Lys}</i>) |
| Neuronal ceroid lipofuscinoses (NCLs) | AR, AD | Variable | Macular degeneration and visual impairment (except adult form) | Lipopigment deposits; granular osmiophilic, curvilinear or fingerprints inclusions | <i>CLN1-CLN9</i> |
| Sialidoses | AR | 8-15 | Gradual cerebellar impairment; cherry-red spot maculopathy | Urinary oligosaccharides, fibroblast neuraminidase deficit | <i>NEU</i> <i>PPGB</i> |

AD: autosomal dominant; AR: autosomal recessive. Modified from Striano *et al.* (2008).

adult-onset NCL (Kufs disease; gene unknown), however, these diseases are excluded by the presence of LB. Mitochondriopathies, such as MERRF and MELAS, usually exhibit maternal mtDNA transmission. Clinical presentation may be heterogeneous and includes a wide range of associated symptoms, such as deafness, low stature, myopathy, lactic acidosis and optic atrophy, with variable prognosis. Sialidosis is an extremely rare lysosomal disease with cherry red spot maculopathy and elevated urine oligosaccharides (Berkovic *et al.*, 1986).

Prognosis and evolution

The prognosis of LD is invariably progressive and fatal, leading to death 5-10 years after clinical onset. Genotype-phenotype correlations do not reveal substantial differences between patients carrying *EPM2A* and *EPM2B* mutations, but a few specific *EPM2B* mutations appear to correlate with a late onset and slow progressing LD (Minassian, 2001; Boccella *et al.*, 2003; Franceschetti *et al.*, 2006; Striano *et al.*, 2008). At present, LD treatment remains palliative, with the best current therapies having limited success in the modulation of symptoms. Commonly used antiepileptic therapy for the management of myoclonus may improve symptoms during the early stages of the

disease (Minassian, 2001; Striano *et al.*, 2008). Hopefully, the work of many investigators, who are putting a massive effort into elucidating the pathophysiology, will result in focused treatments to control the deterioration of this otherwise devastating condition.

Epidemiology of Lafora disease and its genetic correlations

LD is a rare orphan disease. Based on all published reports of LD mutations, we estimate an overall frequency of ~four cases per million individuals in the world. Over 250 patients and/or families have been described with LD (*figure 2A*). Of these, 42 per cent are caused by mutations in *EPM2A* and 58 per cent by *EPM2B* mutations (*figure 2B*). The ratio of *EPM2A* to *EPM2B* cases varies with population, with some regions having many more *EPM2A* cases than *EPM2B*, and vice versa, and this is, remarkably, not solely due to founder mutations (Gomez-Garre *et al.*, 2000; Franceschetti *et al.*, 2006). The most common *EPM2A* mutation is the R241X mutation, which accounts for approximately 17 per cent of *EPM2A*-mediated LD. Large deletions make up 10-15 per cent of *EPM2A* mutations, with the remainder ranging from those causing approximately eight per cent of *EPM2A*-mediated cases of LD (*i.e.* R171H) to orphan

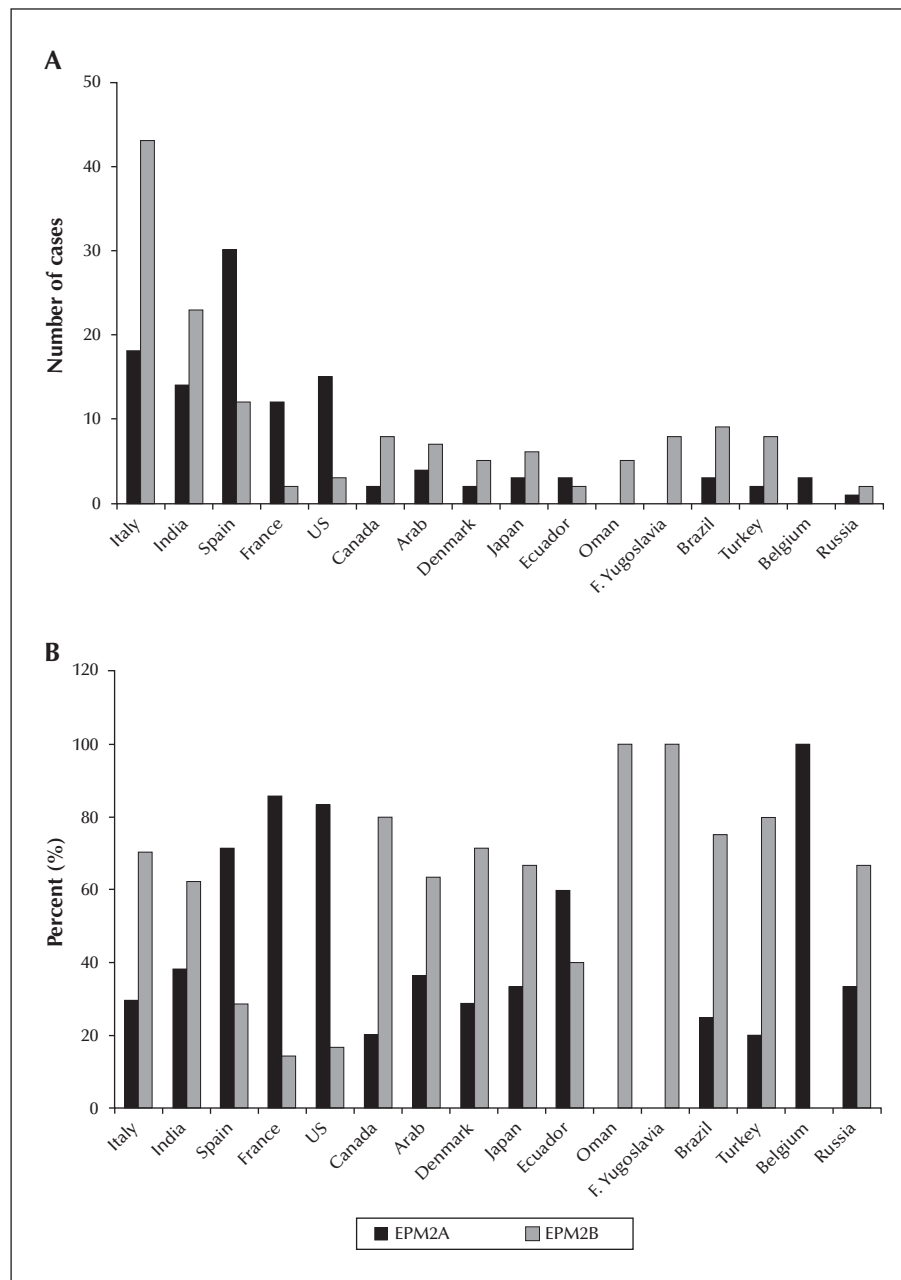


Figure 2. Number (A) and percentage (B) of EPM2A and EPM2B cases according to ethnicity/country, known to us at the time this article was prepared. Only ethnicities/countries with more than one case are shown here.

mutations spread across the gene (figure 3A). For *EPM2B*, the two most common mutations are the missense mutation P69A and the frameshift mutation G158fs16 which affect ~15 and ~eight per cent of *EPM2B*-mediated LD, respectively. As with *EPM2A*, the remaining *EPM2B* mutations span the gene (figure 3B) and are rare, though some mutations (i.e. C26S and D146N) are more frequently observed. Because deletions can be overlooked using conventional sequencing techniques, it is critical to consider

deletion/duplication analysis in any suspected LD patients in whom initial sequencing of *EPM2A* and *EPM2B* reveals no change.

Founder effects and recurrent mutations

Certain mutations appear to be specific to particular ethnic groups and/or geographical locations. For example, LD affects French Canadians from a geographically isolated area of eastern Quebec with an

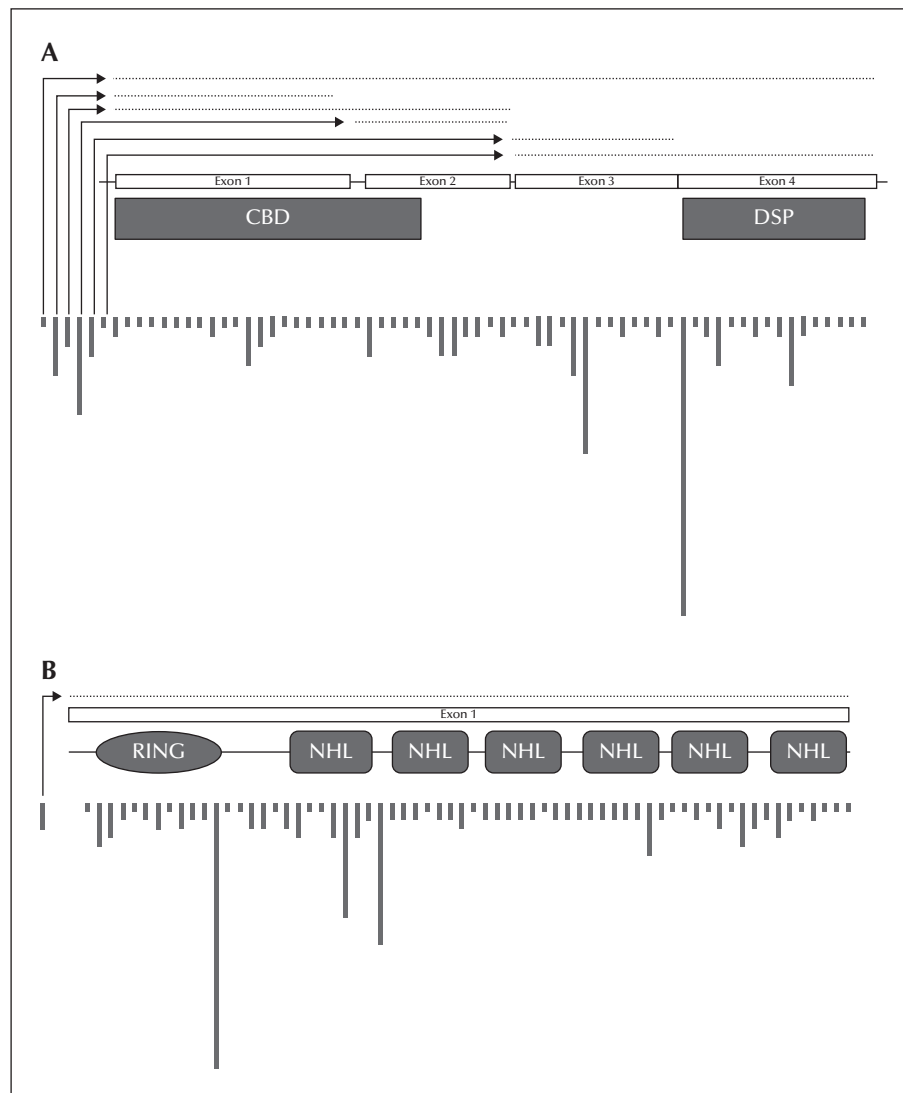


Figure 3. Location and relative frequency of *EPM2A* (A) and *EPM2B* (B) mutations; dotted lines indicate deletion mutations. Locations are approximated across each gene.

unusually high frequency, which is likely to be due to the ancestral *EPM2B* C26S mutation (Chan *et al.*, 2003a, 2003b). One study has been published on LD in Oman, in which all cases in five separate, unrelated families resulted from a single ancestral mutational event in *EPM2B* (Turnbull *et al.*, 2008). Interestingly, the *EPM2A* R241X mutation commonly found in individuals of Spanish descent resulted from both recurrent events and founder effects (Gomez-Garre *et al.*, 2000; Ganesh *et al.*, 2002a). At least five unique haplotypes are associated with this mutation, indicating that a minimum of five separate mutational events have led to the prevalence of the R241X mutation. Accordingly, *EPM2A*-mediated LD is more common than *EPM2B*-mediated LD in Spain. The second most common mutation in *EPM2B*, the missense P69A mutation,

appears to have occurred from multiple mutational events similar to the *EPM2A* R241X mutation (Gomez-Abad *et al.*, 2005). Only one common haplotype was found among eight patients analyzed with P69A mutations, strongly indicative of a recurrent event.

Phenotypic hetero- and homogeneity

Clinically, LD is a fairly homogenous disease with onset in adolescence and neurological decline soon after, but the timing and severity of symptoms can be variable, even within families. Both phenotypic heterogeneity and homogeneity have been noted in LD (see below) and with the large number of mutations in both *EPM2A* and *EPM2B*, genotype-phenotype correlations have been difficult to ascertain. Nonetheless,

some associations between the two have been made. Gomez-Abad *et al.* identified three Arabic families with a complete deletion of the *EPM2A* gene (Gomez-Abad *et al.*, 2007). Haplotype analysis and breakpoint mapping established a shared haplotype among the patients, suggesting a common ancestral origin for this mutation. Of the seven confirmed LD patients in the pedigrees, there was great variability in both disease onset and severity of symptoms, which was unexpected, due to the founder effect of the mutation. Clearly, even with the same ancestral mutation, phenotypic heterogeneity is common. This has been shown with other ancestral founder mutations, including the recurrent R241X *EPM2A* mutation (Gomez-Garre *et al.*, 2000; Ganesh *et al.*, 2002a). However, in a genetic isolate, the same ancestral founder mutation within a population resulted in phenotypic homogeneity (Turnbull *et al.*, 2008). The uniformity of environmental and genetic background reinforces the idea that other genetic and/or environmental modifiers may influence the phenotypic spectrum in LD. Indeed, this has been shown within one family; an *EPM2B* patient harbouring a coding variant for the *PPP1R3C* gene, which encodes protein targeting to glycogen (PTG), exhibited a milder course of LD (Guerrero *et al.*, 2011). The resulting conclusion from reports of both clinical homogeneity and heterogeneity in LD is that even among families, the disease course may or may not be identical and each LD patient is just as likely to have a clinically diverse, rather than classic, disease course.

Genotype-phenotype correlations

'Atypical' LD with early-onset childhood learning disabilities has been reported (Ganesh *et al.*, 2002a; Annesi *et al.*, 2004). Here, the authors showed a significant association with exon 1 mutations in *EPM2A*, linked to the development of learning disabilities prior to the onset of classic neurological symptoms of LD. Exon 4 mutations were mainly associated with classic LD with no childhood-onset educational difficulties. The authors suggest that exon 1 mutations can result in the complete loss of laforin function, whereas exon 4 mutations may preserve some of its functionality. However, other studies have not shown the same association, even with patients with the same or similar mutations in exon 1. Atypical LD with childhood learning difficulties, as observed by Ganesh *et al.* and Annesi *et al.*, may prove to be a sub-syndrome of LD or could be due to as-yet-unknown genetic and/or environmental modifying factors (Lesca *et al.*, 2010).

One study identified a family with two siblings with LD, one of whom presented with severe liver failure as an initial symptom (Gomez-Garre *et al.*, 2007). Liver function in the sibling was abnormal, although the patient remained asymptomatic. Upon further

follow-up, both developed classic LD, and mutation screening revealed a homozygous *EPM2A* R241X mutation in both siblings. Interestingly, the liver disease resembled type IV glycogen storage disease (GSD; due to GBE1 deficiency). The two diseases (type IV GSD and LD) share very similar features, namely the presence of polyglucosan bodies. This suggests a role for modifier genes in LD and the authors propose that these defects may lie in the same metabolic pathway, *i.e.* the glycogen metabolic pathway. The *EPM2A* R241X mutation is a prevalent mutation, and no other cases of hepatic failure have been identified in patients with this mutation. It is likely that other modifying factors led to this presentation, but the similarity to type IV GSD is intriguing and, as the authors of the study mention, certain modifiers may influence the severity of non-neurological symptoms in LD.

Some studies have indicated that patients with malin-mediated LD have a slightly milder disease course than patients with laforin-mediated LD (Baykan *et al.*, 2005; Gomez-Abad *et al.*, 2005; Singh *et al.*, 2006). However, others found no change in severity between the two (Franceschetti *et al.*, 2006; Lohi *et al.*, 2007; Traore, 2009; Lesca *et al.*, 2010; Brackmann *et al.*, 2011). Conclusive evidence involving large-scale comparisons is lacking, but it is clear that there exists one relatively common, milder, *EPM2B* mutation, which may be skewing the comparative results in some analyses. Patients with either heterozygous or homozygous D146N mutation in *EPM2B*, in all cases in our experience and in the literature, have an atypical milder LD consisting of a later onset of symptoms, longer disease course, and extended preservation of daily living activities (Chan *et al.*, 2003b; Baykan *et al.*, 2005; Gomez-Abad *et al.*, 2005; Franceschetti, 2006; Couarch *et al.*, 2011). It is likely that at least some of the functionality of malin is preserved in patients with D146N mutations, as it has been demonstrated that the mutation preserves both the E3 ubiquitin ligase activity and a weak interaction with laforin (Solaz-Fuster *et al.*, 2008). Clinical heterogeneity is common in LD, even among patients with allelic homogeneity and while it is true that certain mutations, such as D146N, cause a milder disease course, most cases of malin-mediated LD have a devastating disease course which is comparable to that of laforin-mediated LD.

The pathology of Lafora disease and Lafora disease animal models

The primary morphological change in LD is the deposition of polyglucosans, which consists of discrete deposits of fibrillary polysaccharides composed of poorly-branched glucose polymers (LBs). They are typically found in the brain, in the periportal hepatocytes

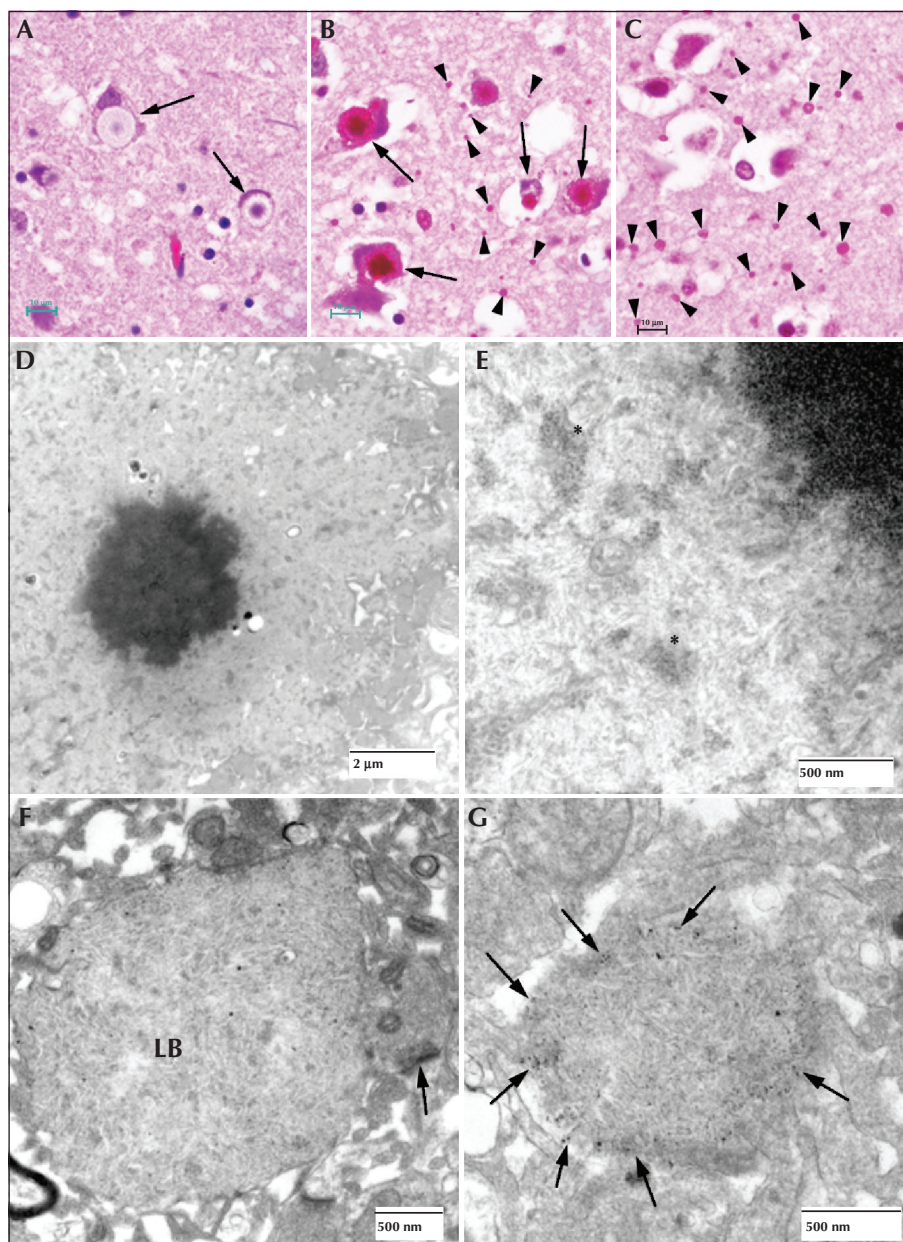


Figure 4. Lafora disease in human brain. (A) Haematoxylin and eosin (H&E)-stained section of brain obtained from an LD patient at autopsy. Arrows indicate perikaryal LBs. Note the outer pale layer and the basophilic core. (B) Diastase-digested periodic acid Schiff (PASD) section of the substantia nigra from the same patient. Note the intense staining of the perikaryal LBs (arrows) and the 'dust-like' LBs (arrowheads). (C) PASD-stained cerebral cortex. Note the presence of numerous 'dust-like' LBs (arrowheads). (D) Low-power electron micrograph of a perikaryal LB. Electron dense core is the equivalent of the basophilic core seen in H&E-stained material. (E) Higher power of (D). As well as the filamentous polyglucosans, aggregates of 'glycogen-like' material were frequently seen asterisks. (F) Electron micrograph of a dendritic LB. Note the synaptic density (arrow). (G) Electron micrograph of a tannic acid-stained LB. Note the presence of glycogen particles at the periphery (arrows).

of the liver, skeletal and cardiac myocytes, and in the eccrine duct and apocrine myoepithelial cells of the sweat glands (Sakai *et al.*, 1970; Carpenter & Karpatis, 1981a; Shirozu *et al.*, 1985; Barbieri *et al.*, 1987).

In the brain, the largest LBs tend to have two layers of various proportions in haematoxylin and eosin (H & E)-stained preparations, the outer layer being

pale and the core basophilic (*figure 4A*). These large bodies are usually found in neuronal perikarya. The ratio of perikaryal LBs to neurons is highest in the substantia nigra, followed by the dentate nucleus and some thalamic nuclei (*figure 4B*). They are rare or absent in the anterior part of the spinal cord. In the cerebral cortex, perikaryal LBs are rather

scant, but a periodic acid Schiff (PAS) stain reveals numerous small LBs in the cortical neuropil (figure 4C), the majority being found in dendrites. These are commonly referred to as 'dust-like' in appearance. The PAS-positive material found in both perikaryal and 'dust-like' LBs, as well as LBs found in other tissues, are diastase-resistant, hence for all diagnoses of suspect LD patients, diastase-treated PAS (PASD) staining plays an integral role in the identification of the disease in both surgical biopsy and autopsy materials. Some patchy neuronal loss may be present in the later stages of the disease, but this is not a prominent feature (Sakai et al., 1970; Carpenter & Karpati, 1981a).

Electron microscopy confirms the presence of fibrillary accumulations typical of polyglucosans within the LBs. Using electron tomography, we were recently able to identify an ordered structure where individual polyglucosan fibrils bifurcate at a regular periodicity, ranging from 50-125 nm, depending on the tissue of origin (unpublished observation). This is especially true of the perikaryal and 'dust-like' LBs found in the brain, in skeletal muscle myocytes, and all of the LBs in affected sweat glands. Along with the filaments, there is often poorly-defined granular material within the LB itself. In two extremely well preserved brain biopsy specimens, we observed "glycogen-like" particles in the dendritic cytoplasm and the perikaryon of neurons in both perikaryal and 'dust-like' LBs (unpublished data) (figure 4D-G). Glycogen is rarely found in neurons because the sugars are rapidly metabolized. In skeletal muscle and cardiac myocytes, the fibrillary polysaccharide accumulates in membrane-bound spaces which are not lysosomal (Yokoi et al., 1975; Carpenter & Karpati, 1981a; Barbieri et al., 1987). There is variation in the morphology of the enclosed material, including some granular material, but the majority is fibrillar. Skeletal muscle can be used as a source for morphological diagnosis, but it is far from ideal, since the degree of involvement varies, and not all muscle fibre types are involved (Neville et al., 1974; Turnbull et al., 2011a) (figure 5A, B). This has been clearly demonstrated in a laforin knockout mouse model, and retrospective studies on human muscle biopsies are currently underway. Similar to the mouse, preliminary data suggest that humans do in fact produce LBs in type II fibres (Turnbull et al., 2011a).

Hepatic insufficiency is an early event in rare LD patients (Nishimura et al., 1980; Tomimatsu et al., 1985; Gomez-Garre et al., 2007). In the liver, PASD staining shows deposits of large portions of PASD-positive material in many of the periportal hepatocytes, confining the nucleus and other organelles to one side of the cell. Liver biopsy has been used for diagnosis, but the pathology is not specific to LD alone (Nishimura et al., 1980). Electron microscopy of periportal hepatocytes with LBs reveals lakes of loosely packed fibrillar

material with some glycogen particles and/or rosettes within the LB. Although a few organelles such as mitochondria and peroxisomes can be found in the LB, the majority of the smaller organelles are confined to a thin layer of cytoplasm which is continuous with the thicker side of the cell which contains the nucleus, the majority of the endoplasmic reticulum, and the Golgi (figure 5C, D).

The occurrence of LBs in the sweat glands has become the gold standard in the pathological diagnosis of LD (Carpenter & Karpati, 1981b; Andrade et al., 2003). Skin is the most accessible of the affected tissues and biopsy procedures are the least invasive. LBs can be found in the eccrine duct cells, close to the secretory coil or close to the surface. They are extremely PASD-positive and are roughly the size of the cell nucleus. Under the electron microscope, LBs are almost 'starch-like' in appearance, however, this is not a diagnostic feature. The eccrine myoepithelial cells rarely contain LBs. The apocrine myoepithelial cells, in contrast, contain LBs in almost all cases. The apocrine secretory (luminal) cells never contain LBs, but do contain PASD-positive inclusions which can be misleading (Andrade et al., 2003) (figure 6).

A third gene (*PRDM8*) has recently been discovered in a single family. As with *EPM2A* and *EPM2B* mutations, the patients have a progressive myoclonus epilepsy which is similar to typical LD, but also exhibit significant differences (Turnbull et al., 2012). The sweat glands in these patients contain no LBs. At present, we have not been able to study the brain of these patients for lack of material. Absence of LBs in the sweat glands of these patients could influence the current diagnostic algorithm. If a patient has all the clinical manifestations of LD, a new gene defect is detected, and there are no LBs found in the sweat glands in the skin biopsy, the physician should consider an open muscle biopsy, as LBs are readily found in the muscle.

In order to further our knowledge of LD, it is necessary to study animal models of this disease which could lead to breakthroughs in the understanding of the pathogenesis, which may subsequently lead to potential therapies. Polyglucosan storage disease with myoclonic epilepsy has been known to exist in animals since it was reported in a beagle in the early 1920s. Although it has only been reported in several breeds of dogs and other species of the Canidae class, as well as cockatiels, pigs and cattle, it has probably been overlooked in many other species, since it is rare for animals to be subjected to post mortem examination, especially non-domesticated animals. The most widely studied of these naturally occurring animal models is the pure bred dog. As these animals are bred from a limited gene pool, the potential for consanguinity is greatly increased. Fortunately, breeders are regulated by kennel clubs and in order for these dogs to be

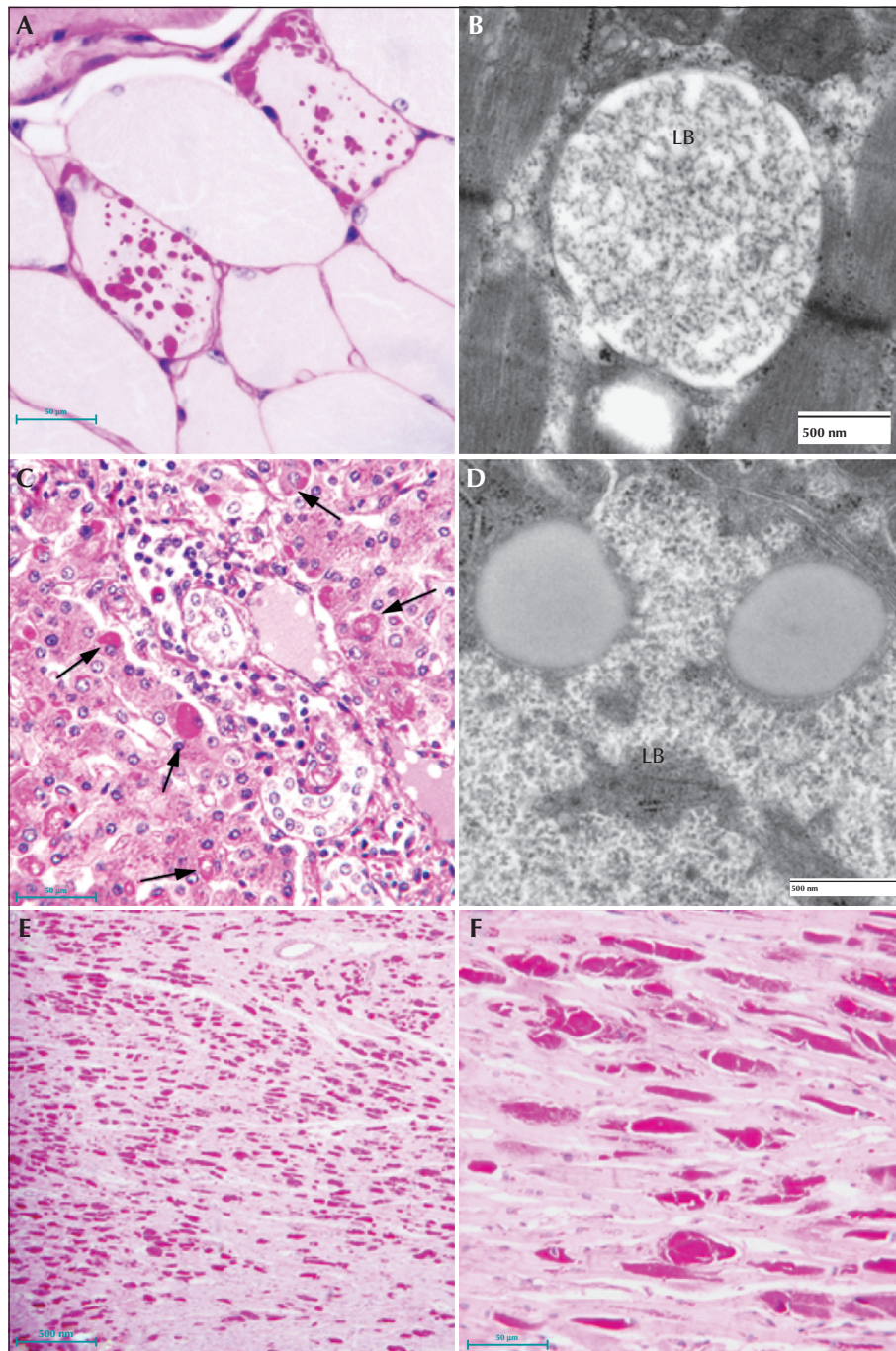


Figure 5. Lafora disease in human muscle, liver, and heart. (A) PASD-stained formalin-fixed muscle biopsy from a patient. Two of the fibres contain LBs. (B) Electron micrograph of an LB in muscle. Note that it is membrane-bound and contains both fibrillar and granular material. (C) PASD-stained hepatic periportal region of an LD patient at autopsy. Arrows indicate hepatocytes containing LBs. (D) Electron micrograph of a LB in liver from biopsy material. Note the thin cytoplasm and the predominately granular, with some filamentous, material forming the LB. (E) Low power of a PASD-stained heart from a LD patient at autopsy. Note the numerous LDs. (F) Higher magnification of (E). LBs occupied a high percentage of the sarcoplasm of the cardiac myocyte.

sold as pure bred, evidence of a known lineage and a stringent medical examination must first be submitted to these organizations, prior to a certificate being issued. As the majority of these animals are

sold as companion animals, the dogs are routinely examined by qualified veterinarians and sometimes, although rarely, LD is detected in these dogs. LD in dogs can occur spontaneously in any breed of

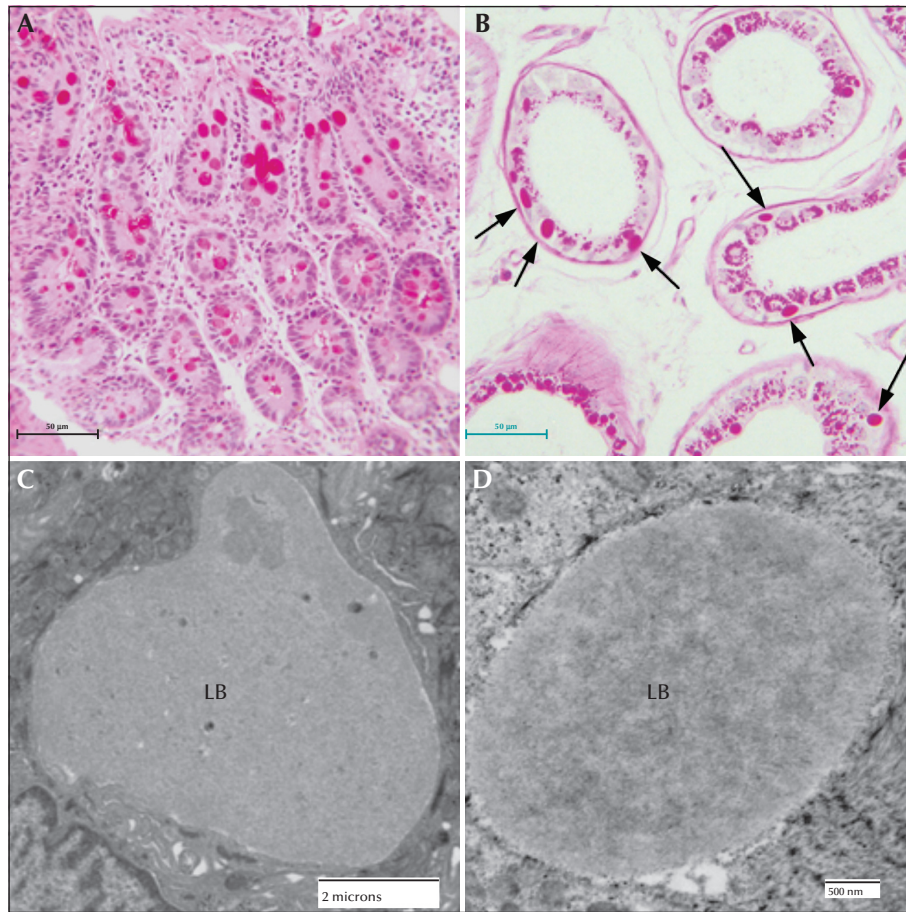


Figure 6. Lafora disease in human sweat glands. (A) PASD staining of LBs in ductile cells in eccrine sweat glands. (B) PASD staining of LBs in the myoepithelial cells of apocrine sweat glands (arrows). (C) Electron micrograph of an LB from a ductile cell in an eccrine sweat gland. (D) Electron micrograph of an LB in a myoepithelial cell from an eccrine sweat gland.

dog, but it particularly affects miniature wire-haired dachshunds, basset hounds, and beagles (Kaiser *et al.*, 1991; Gredal *et al.*, 2003; Lohi *et al.*, 2005). The disease does not present in dogs until at least five years age (35 in human years).

Myoclonus is the dominant feature of the canine disease and this can be induced by flashing lights, sudden sounds, and movement. Generalized or complex partial seizures can be seen in some dogs. The disease progresses over many years and gradually other neurological deficits, such as ataxia, blindness, and dementia, occur. Unlike humans, affected animals have a near-normal life expectancy, though as quality of life diminishes, the owner may be forced to euthanize the animal.

Pathological examination of these dogs revealed distribution and frequency of LBs in the brain identical to that found in the human disease (Lohi *et al.*, 2005). As with humans, the liver contains LBs only in periportal hepatocytes. Both cardiac and skeletal myocytes contain LBs. Dogs do not have sweat glands in the

skin, except in the footpads, where both apocrine and merocrine sweat glands are present; the LBs are identical in size and structure to those of the human disease. They are found in the apocrine myoepithelial cells and in the merocrine ductile and myoepithelial cells (figure 7).

With the identification of two genes (*EPM2A* [laforin] and *EPM2B* [malin]) responsible for LD, it was only logical that transgenic mice technology be utilized to develop murine models of the disease, in order to further advance our knowledge of LD.

The first of the animal models developed was a laforin-deficient murine mutant (Ganesh *et al.*, 2002b). This was achieved by deleting the dual-specificity phosphatase domain of the *EPM2A* gene. These null mutants developed LBs in the brain, specifically in the hippocampus, cerebellum, cortex, and brainstem. Neuronal degeneration was observed in younger mice, but the same phenomenon was observed in age-matched wild-type littermates, which is suggestive of neuronal remodelling during development.

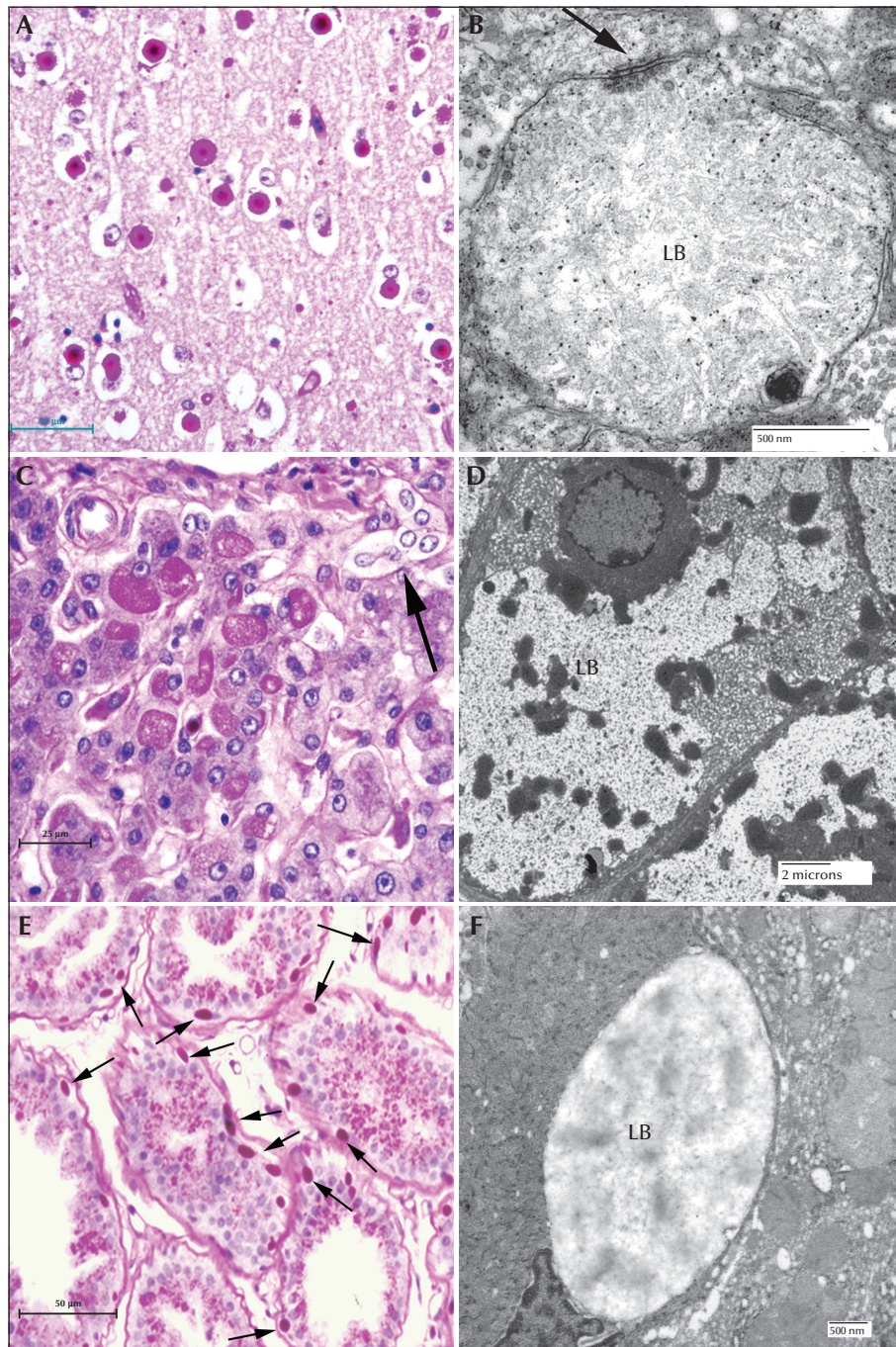


Figure 7. Canine Lafora disease. (A) PASD-stained brain from a miniature wire-haired dachshund. Numerous perikaryal and 'dust-like' LBs are seen. (B) Dendritic LB. Note the filamentous polyglucosans. Arrow indicates the synaptic density. (C) PASD staining of a liver from an affected dog. A cluster of hepatocytes containing LBs is seen in the portal tract. Arrow indicates a bile duct. (D) Low-power electron micrograph of a hepatocyte containing an LB (LB). Note how the majority of the cytoplasm has been pressed to the periphery of the cell. (E) PASD-stained sweat gland from the footpad of an affected dog. Note (arrows) the numerous LBs confined to the myoepithelial cells. (F) Electron micrograph of an LB in a myoepithelial cell.

The onset of LB accumulation occurred at two months. The highest number of perikaryal LBs was found in the molecular layer of the cerebral cortex and, as with humans, perikaryal LBs had two layers of

various proportions; the outer layer being pale and the core basophilic. 'Ground glass' LBs were found throughout. Mild focal neuronal degenerative changes were detected primarily in the Purkinje cells

of the cerebellum and were particularly present in the later stages of the disease. Ultrastructural examination of these tissues revealed prominent LBs in many of the neurons in all of the affected areas. As with dogs, mice have no sweat glands in the skin, except in the footpads where LBs were found. LBs were also found in skeletal and cardiac myocytes. No LBs were detected in the liver using either PASD staining or electron microscopy, although they were reported to be present using an antibody against polyglucosan bodies.

A transgenic mouse was generated which overexpressed myc-tagged inactivated laforin, in order to trap the substrate of laforin (Chan *et al.*, 2004). Myc-laforin was expressed 150-fold higher than endogenous laforin. LBs were formed in the neuronal perikarya and dendrites, liver, sweat glands in the footpad, and skeletal and cardiac myocytes. In the liver, hepatocytes with LBs were found in discrete clusters throughout zones 2 and 3. Using an antibody against myc with immunoperoxidase staining or immunogold labelling, we were able to determine both the cellular and subcellular distribution of myc-laforin. Particularly striking was the staining of the LBs in cerebellar Purkinje cell somas and dendrites and stellate neurons, as well as the presence of discrete punctate structures consistent with LBs throughout the molecular layer of the cortex, and in some hippocampal and cerebral neurons. Overall, however, LBs in the brain in this model were much lower in number than in the knockout mouse mentioned above.

In addition, a malin knockout mouse was developed. The malin exon was removed by *in vivo* recombination and an embryonic stem cell line (ESL) was created. The ESL was aggregated and a chimeric mouse was generated. These were bred into a C57B6 background and the malin null mouse line was established from the resultant heterozygous mice (Turnbull *et al.*, 2010). These mice, like the laforin knockout mice, developed LBs in the brain at around two months. These null mutants developed LBs in the brain in the hippocampus, cerebellum, cortex, and brain stem. As was the case for the laforin-deficient mouse, the LBs consisted of two populations. The perikaryal LBs consisted of two layers; the outer layer being pale and mildly eosinophilic and the core basophilic. These were found primarily in the cell bodies of neurons in the cerebral cortex molecular layer. "Dust-like" LBs were found throughout the grey matter. LBs were also found in the sweat glands of the footpads and in skeletal muscle and cardiac myocytes. Unlike the laforin knockout or mutant laforin mice, malin null mice had LBs in some of the periportal hepatocytes. Not unlike the human or canine liver LBs, they occupied the centre of the cells, pressing the nuclei and the majority of cytoplasm to one side, as well as the periphery of the cell. Under the electron microscope,

the LBs formed lakes which consisted of clusters of 'glycogen-like' particles and fibrillar material. Other groups have also generated malin knockout animals with similar results. Pathological sample images from the third mouse model mentioned above are presented in *figures 8, 9 and 10*.

One of the most exciting developments in the study of LD using transgenic mice is the use of double knockouts. In LD, glycogen synthase (GS) over-activity is considered one of the primary culprits for the formation of polyglucosans (Vilchez, 2007). It has been hypothesized that the reduction in GS activity might prevent polyglucosan formation. Laforin-deficient mice were bred with mice deficient in PTG, a protein involved in activating GS, resulting in LD mice lacking the GS-activating effect of PTG. The resultant double knockout (DKO) mice have almost no polyglucosan or neurodegeneration, and no seizures (Turnbull *et al.*, 2011b). The development of this DKO mouse demonstrates the importance of using this approach to create strategies for therapeutic interventions which ultimately may result in a cure for this devastating disease. Due to the inability to use this approach in humans, results from this and knockouts of other glycolytic pathway constituents in other DKO mice are likely to reveal molecules which, when inactivated or eliminated, lead to the control or cure of LD. Once these molecules have been identified, the development of small molecule antagonists against GS, its activators, and potential GS up-regulators will be developed as potential therapies for human LD.

Pathogenesis of Lafora disease

Normal glycogen remains soluble in the cell due to its highly organized structure, whereas polyglucosans precipitate in the cell due to disturbances in the structure of the carbohydrate, and aggregate into LB (Tagliabracci *et al.*, 2008). LBs are very similar in morphology to polyglucosan bodies seen in glycogen-branching enzyme deficiency, differing only in their cellular location. In both LD and glycogen-branching enzyme deficiency, polyglucosans form in neurons. In LD, they are exclusively seen in the cell bodies and dendrites. In glycogen-branching enzyme deficiency, they are seen in axons. The two diseases have markedly dissimilar clinical symptoms, which is most likely to be due to this difference in polyglucosan localization. LD is a progressive myoclonus epilepsy, whereas glycogen-branching enzyme deficiency patients suffer from an adult-onset motor neuron disease, similar to that seen in amyotrophic lateral sclerosis (ALS) (Bruno *et al.*, 1993).

EPM2A encodes a protein named laforin which contains a dual specificity phosphatase domain and a

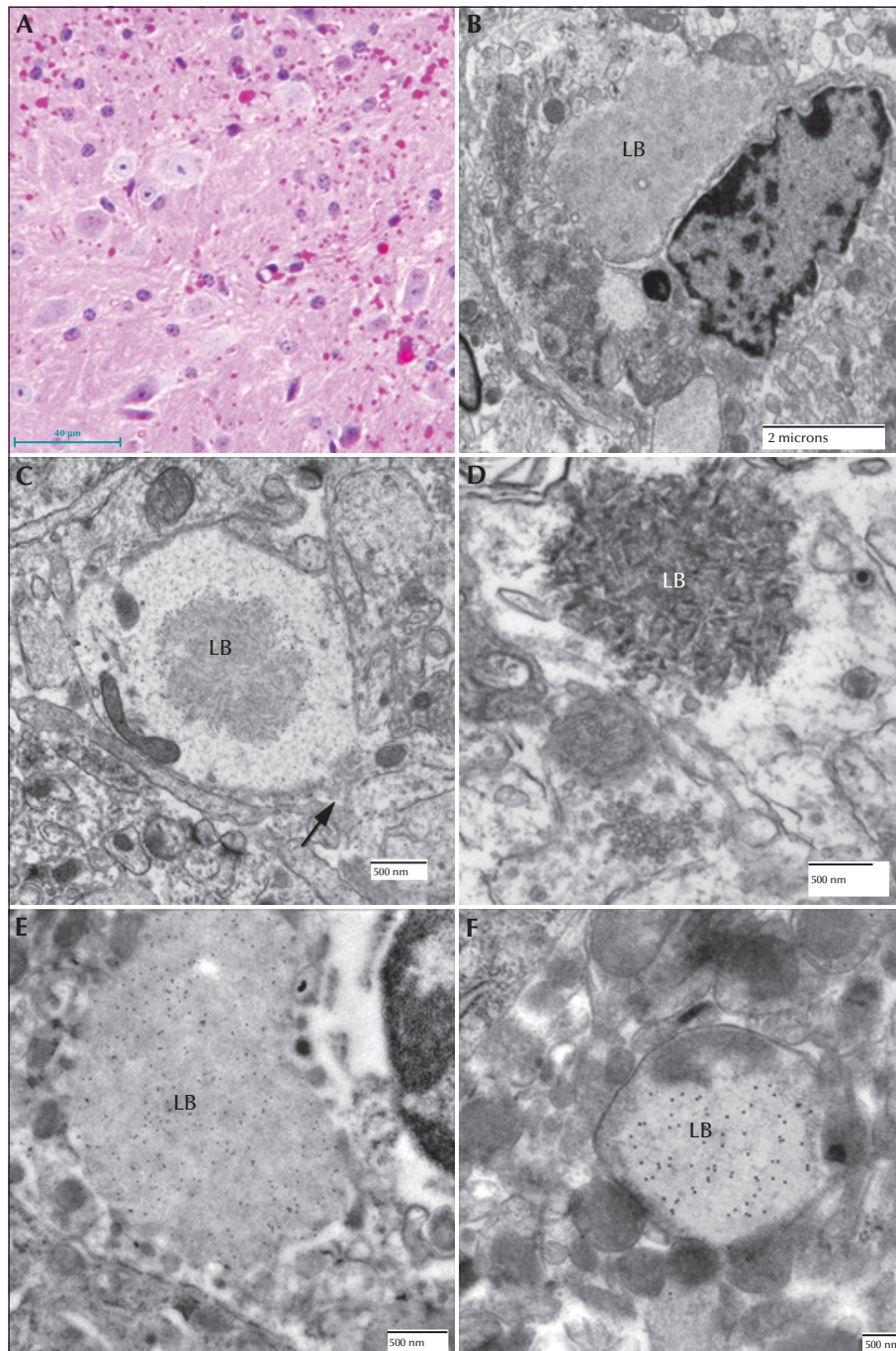


Figure 8. Lafora disease in the brain of a mouse model. (A) PASD-stained cortex from a malin-deficient mouse. Both perikaryal and 'dust-like' LBs are seen. (B) Electron micrograph of a juxtanuclear LB (LB) from a laforin knockout mouse. (C) Electron micrograph of a dendritic LB (LB) from a laforin-deficient mouse. Note central location of the polyglucosans within the dendrite. Arrow indicates the synaptic density. (D) Electron micrograph of an LB from a malin-deficient mouse. Polyglucosans appear thicker and more electron opaque. (E) Electron micrograph of a perikaryal LB from a myc-tagged mutant laforin over-expressing mouse, which has been immunogold labelled for myc. Gold label is confined to the LB. (F) Electron micrograph of an LB from the same mouse line as (D). Gold label is confined to the LB.

carbohydrate binding motif (Minassian *et al.*, 1998). The second LD gene, *EPM2B*, encodes malin, an E3 ubiquitin ligase (Chan *et al.*, 2004). Over a decade has passed since the second causative LD gene was

identified. Since then, a number of hypotheses have been put forward as to the function of both malin and laforin in the pathogenesis of LD. The next decade will, most likely, unravel the complete pathway leading to

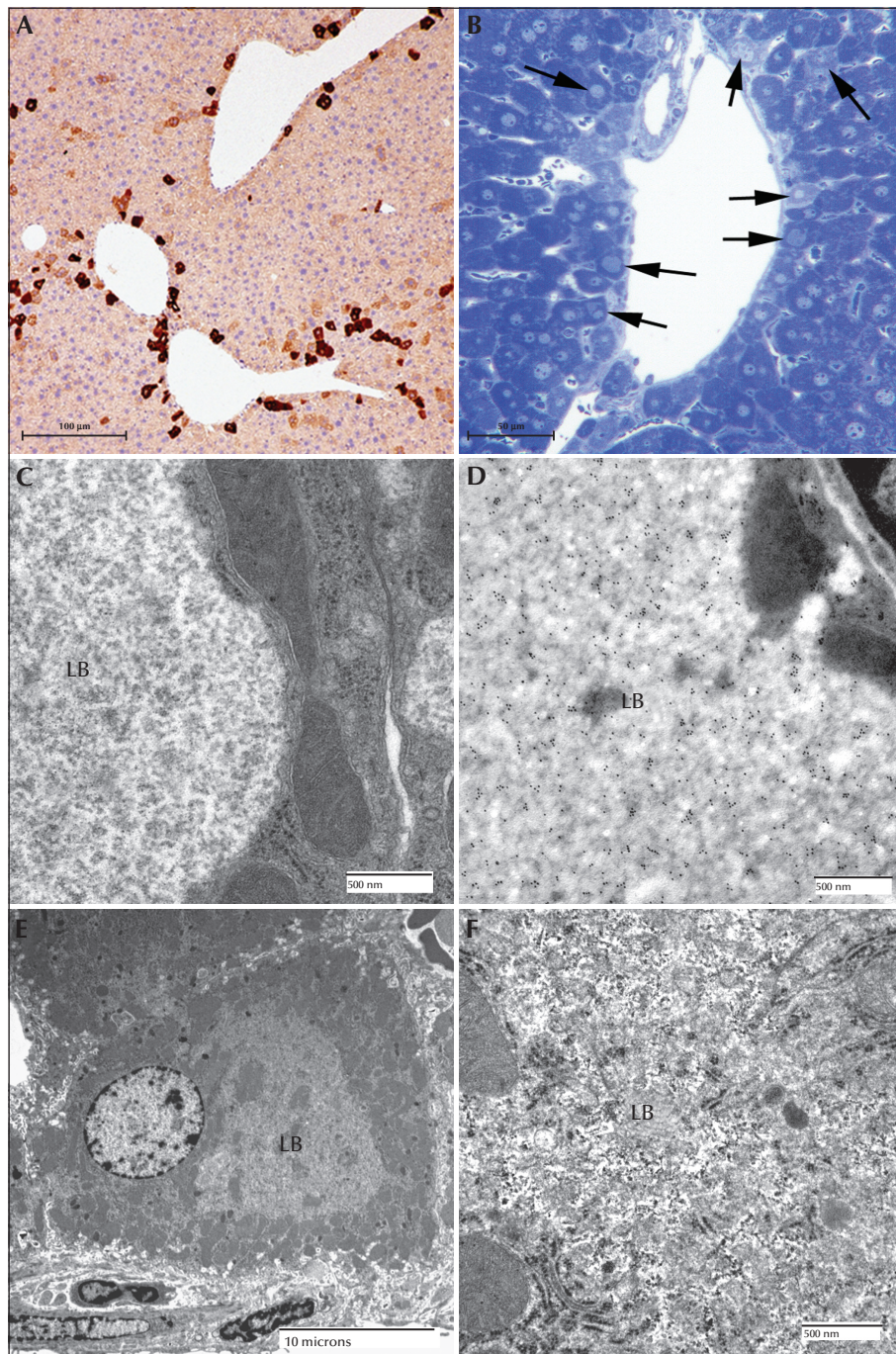


Figure 9. Lafora disease in the liver of a mouse model. (A) Low power of myc immunoperoxidase-stained liver from a myc-tagged mutant laforin over-expressing mouse. Hepatocytes with LBs were only found in zone 2 or 3. (B) Toluidine blue stained section of a portal tract from a malin-deficient mouse. Arrows indicate hepatocytes containing LBs. (C) Electron micrograph of a LB (LB) from a myc-tagged mutant laforin over-expressing mouse hepatocyte. Note how the cytoplasm has been pressed to one side. Note the predominately granular nature of the LB. (D) Electron micrograph of a LB from a myc-tagged mutant laforin over-expressing mouse hepatocyte that has been immunogold labelled with an antibody against myc. Label is confined to the LB. (E) Low-power electron micrograph of a malin-deficient periportal hepatocyte containing an LB. (F) Higher power of (E) showing that the LB contains both granular and filamentous material (LB).

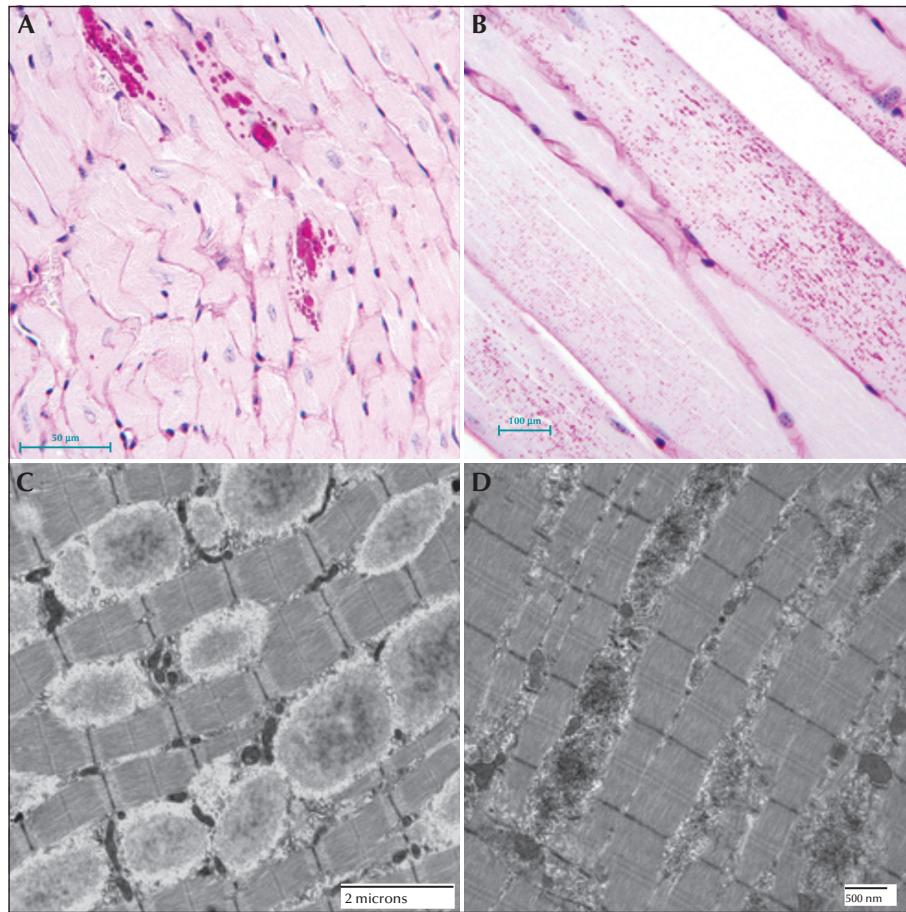


Figure 10. Lafora disease in the heart and skeletal muscle of a mouse model. (A) PASD-staining of a laforin-deficient mouse heart. LBs were detected in some of the cardiac myocytes. (B) PASD-staining of malin-deficient mouse skeletal muscle. Numerous LBs were detected in some of the muscle cells. (C) Electron micrograph of LBs in laforin-deficient mouse skeletal muscle. (D) Equivalent field in a malin-deficient mouse muscle. Note the overall electron opaque filamentous appearance of the LBs.

LD and provide the knowledge which is necessary for finding a cure for this devastating disease.

LD has been speculated to be caused by defects in the clearance system(s) of the cell, similar to diseases such as Parkinson's and Alzheimer's. The presence of large inclusions and LBs, along with the finding that LBs contain a minor component of protein(s), suggested a defect in either autophagic processes and/or protein clearance. In keeping with this line of thought, defects in both autophagy and protein clearance have been reported in both *Epm2a*^{-/-} and *Epm2b*^{-/-} mice (Garyali *et al.*, 2009; Aguado *et al.*, 2010; Knecht *et al.*, 2010; Puri & Ganesh, 2010; Rao *et al.*, 2010; Criado *et al.*, 2011; Puri *et al.*, 2012). Unexpectedly, the autophagic dysfunction in *Epm2b*^{-/-} mice is different to that of *Epm2a*^{-/-} mice, resulting from an mTor-independent pathway (Criado *et al.*, 2011). This suggests that if autophagic dysfunction is, indeed, at least partly causative of LD, mutations in malin and laforin may have separate and dissimilar disease mechanisms, which would be surprising, given the highly

similar phenotypic outcomes in both laforin- and malin-deficient LD.

Malin has been reported to ubiquitinate a number of proteins, including laforin (Gentry *et al.*, 2005), glycogen-debranching enzyme (Cheng *et al.*, 2007), PTG/GS (Vilchez, 2007; Solaz-Fuster *et al.*, 2008; Worby *et al.*, 2008), neuronatin (Sharma *et al.*, 2011), AMPK (Moreno *et al.*, 2010), and dishevelled2 (Sharma *et al.*, 2012). However, only laforin and GS have been clearly shown to be increased in *Epm2b*^{-/-} mice (Tagliabracci *et al.*, 2007, 2008; DePaoli-Roach *et al.*, 2010; Turnbull *et al.*, 2010; Valles-Ortega *et al.*, 2011), indicative of *in vivo* action of malin on laforin. Additionally, no changes affecting the ubiquitin-proteasomal system were seen in *Epm2b*^{-/-} mice (Criado, 2011), while these defects have been seen in a number of laforin-deficient models. It therefore appears that these protein clearance defects are a secondary consequence of LB formation, which is supported by recent studies showing that removal of the major polyglucosan component of LB by downregulation of glycogen synthesis results in

near-complete disease resolution (Turnbull *et al.*, 2011; Duran *et al.*, 2014).

A major prevailing hypothesis over the years in LD research has focused on the balance between glycogen synthesis and glycogen branching (Lohi *et al.*, 2006). Normally, GS synthesizes glycogen while the branching enzyme promotes the extension of the growing chain. This balance results in a soluble, highly ordered glycogen molecule. When this balance is disturbed in the direction of synthesis (*i.e.* when branching enzyme is deficient), polyglucosans form, as is seen in patients with glycogen-branching enzyme deficiency. The similarities between LBs and the accumulations seen in glycogen-branching enzyme deficiency, as well as data showing that over-expression of GS can also cause polyglucosan accumulation (Raben *et al.*, 2001), make this an appealing and logical hypothesis. More substance was added to this proposal that there is a misbalance between synthesis and branching, when Fernández-Sánchez and colleagues demonstrated an interaction between laforin and PTG, a glycogen-targeting subunit of protein phosphatase-1 (PP1) (Fernandez-Sanchez *et al.*, 2003). PTG directs PP1 to the glycogen-metabolizing enzymes, activating GS while inhibiting glycogen phosphorylase, and thus leading to a net increase in glycogen synthesis (Printen *et al.*, 1997). In addition to its interaction with PTG, laforin has also been shown to interact with other regulatory subunits of PP1, including GL and R6, members of the same family as PTG. Subsequent studies have confirmed the PTG interaction using cell over-expression systems *in vitro*, but no *in vivo* interaction studies have been reported, which is likely to be due to a lack of a suitable antibody to PTG. Interestingly, some disease-causing mutations in laforin specifically affect its interaction with PTG, indicating that disruption of this interaction is pathogenic. Dysregulation of PTG has been hypothesized to cause LD, primarily based on results from *in vitro* studies. Elegant work by Vilchez and colleagues showed that malin and laforin act together to control levels of glycogen synthesis by targeting PTG and/or GS for proteasomal degradation (Vilchez, 2007; Solar-Fuster, 2008; Worby *et al.*, 2008). When either laforin or malin is missing, levels of PTG, an indirect activator of GS, increase, causing GS over-activation. In this hypothesis, over-active GS would cause an imbalance between glycogen branching and extension, leading to the formation of LBs. Results from studies using animal models of LD showed no increases in PTG levels or GS activity, indicating that dysregulation of PTG may not be causative of LD (DePaoli-Roach *et al.*, 2010; Turnbull *et al.*, 2010; Valles-Ortega *et al.*, 2011). Nonetheless, the large amount of *in vitro* data, along with a study showing that removal of PTG from laforin-deficient mice resulted in a dramatic reduction in LB and a cure for

LD in the mouse, strongly supports the conclusion that PTG and laforin interact, and that both are involved in the same metabolic pathway.

The beginnings of an attractive hypothesis emerged from the laboratory of Jack Dixon when they reported that laforin was a carbohydrate phosphatase which could dephosphorylate amylopectin, a plant carbohydrate (Worby *et al.*, 2006). Following this, Tagliabracci and colleagues expanded on this work in a series of elegant experiments. First, they demonstrated that laforin could indeed dephosphorylate glycogen and, most importantly, that glycogen phosphate levels were present at a much higher level than normal in mice lacking laforin (Tagliabracci *et al.*, 2007, 2008). The covalently bound phosphate on glycogen was shown to be present at all available glucose carbons of glycogen, *i.e.* C2, C3 and C6 (Tagliabracci *et al.*, 2011; Nitschke *et al.*, 2013). Precisely where the phosphate on glycogen originates and how it results in glycogen becoming polyglucosan remains unknown. It was shown that laforin removes these phosphates during glycogen degradation (Irmia, 2015). This emerging phosphate hypothesis is compelling, but fails to identify a role for malin in LD. At 12 months of age, laforin-deficient mice have a 4.2-fold increase in muscle glycogen phosphate, whereas malin-deficient mice present a more modest increase of 2.8-fold over normal wild-type levels (Tagliabracci *et al.*, 2008; Tiberia *et al.*, 2012). The lack of a comparable phosphate increase in malin deficiency when laforin deficiency and malin deficiency are clinically equivalent raises the likelihood of other mechanisms at play in malin-deficient LD. Based on our most recent results from malin-deficient mice, we showed that absence of malin leads to increased laforin, as an initial step prior to any LB formation, and then progressive accumulation of laforin in glycogen. We also found that the gradual accumulation of laforin in glycogen renders the latter progressively less soluble. In related work, we showed that over-expressing laforin in cell culture leads to a conversion of glycogen to polyglucosan masses. This phenomenon continues to occur when laforin mutants, which bind glycogen but are phosphatase-inactive, are used. This no longer occurs when laforin mutants which cannot bind glycogen are used. Collectively, these results suggest that malin functions to regulate the quantity of laforin, and excess laforin on glycogen is detrimental to glycogen, leading to glycogen conversion to polyglucosan and LB (Tiberia *et al.*, 2012).

The last decade has brought about a number of breakthroughs in the understanding of LD pathogenesis, though a clear picture has not yet formed. Each new hypothesis has come with its own weaknesses. The two main hypotheses of LB formation centre round the synthesis of glycogen and its structure. Compelling evidence has been presented for both, though

each has their own caveats. Tiberia and colleagues attempted to propose a unifying hypothesis (Tiberia *et al.*, 2012). Firstly, they considered laforin's role as a glycogen phosphatase, which removes phosphate from glycogen allowing it to remain soluble in the cell. Secondly, malin acts on laforin, and is likely also to act on other glycogen metabolic enzymes, to remove them from the glycogen molecule. If laforin is missing, phosphate accumulates and causes glycogen to become structurally abnormal and precipitate. If malin is missing, laforin remains "stuck" to glycogen, again disturbing the precise structure of glycogen and causing it to precipitate. This hypothesis answers a number of outstanding questions in LD research, foremost addressing the role of malin in LD. Here, malin has two roles; the first to control cytosolic amounts of laforin to keep it from inadvertently 'clogging up' the structure of glycogen, and second, to remove laforin (and probably other proteins, such as GS) from glycogen itself after laforin has removed the specific phosphate. Gentry and colleagues (Gentry *et al.*, 2005) had previously speculated that malin may play a role in the regulation of laforin levels, but the actual mechanism behind this was unknown. In this hypothesis, both laforin deficiency and malin deficiency lead to LB formation by means of foreign matter on the glycogen molecule itself (phosphate and protein, respectively). While this theory appeared to bring together disparate aspects of LD research, most recent results once again raised a new and serious challenge. It was shown that expressing a phosphatase-inactive laforin in laforin knockout mice can fully rescue LD in these mice (Gayarre *et al.*, 2014). This finding puts back into question the role of phosphate in polyglucosan formation, causing the field to be re-examined once more. The field has, however, been very rich in results since the genes for this disease were identified. It is hoped that a few new breakthroughs will lead to a comprehensive understanding of this deadly disease.

Disease management and therapy

Lafora disease (LD) is a devastating condition, with severely limited life expectancy and therapeutic options which continue to be limited. Its striking clinical and EEG features may lead to early diagnosis, but because it remains uncommon, few clinicians gain significant experience of this condition, and diagnosis is often delayed. Moreover, LD is a recessively inherited condition, with founder effects and increased prevalence associated with inbreeding, and thus LD is unevenly distributed across the world, which contributes to 'knowledge gaps' about the condition. We will try here to delineate the practical steps that we recommend in the management of patients with

LD. It is clear, in our eyes, that there is an intermediary stage: the clinical diagnosis of LD is now comparatively easy to confirm. Efficient anticonvulsants can be used to alleviate the burden of the attacks, but better insights into the mechanisms underlying the production of LBs and the progression of clinical symptoms have not yet produced ground-breaking therapeutic advances. Thus, dealing with LD patients and their families is not an easy task, because there is little hope to offer and the genetic nature of the condition raises many questions within the affected families.

Management: diagnosis and genetic counselling

Given the severity of the prognosis, a biological confirmation of the diagnosis of LD is necessary; this used to be made based on typical pathological findings (from a skin, muscle, or liver biopsy), but nowadays relies on genetics. Genetic screening, which remains costly, should be justified by solid clinical and EEG evidence, and should be performed together with a variety of tests, while screening for all possible genetic mechanisms in a poorly-assessed case with epilepsy and myoclonus.

The diagnosis of LD is based on three levels of evidence (figure 11):

- The clinical evidence is a compound of history-taking (family background, circumstances, aspects and progression of seizures and myoclonus, and visual agnosia) and examination (including cognitive and psychological assessment, exclusion of associated symptoms such as sensory deficits, and video documentation of myoclonus and general behaviour). The most common clinical situation is one where idiopathic generalized epilepsy (IGE) or even juvenile myoclonic epilepsy (JME) has been diagnosed, with a re-assessment of the patient's situation because of a very unusual evolution which tends towards worsening; the clinical work-up should help exclude the possibility of aggravation of IGE by inappropriate antiepileptic drugs (AEDs), which may result in a pseudo-PME.

- Complementary evidence is based on a thorough evaluation of the EEG, polygraphic EEG, and video-EEG (with an assessment of progression of changes over time). Neurophysiology may also help distinguish between LD and other adolescent-onset PMEs, e.g. Unverricht-Lundborg disease, in which the EEG changes are less pronounced, or juvenile ceroid-lipofuscinosis, in which prominent single-flash responses on the EEG are found. Other procedures may also help in the differential diagnosis (neuroimaging/MRI is not informative in LD). Pathological demonstration of LBs in sweat gland duct cells on axillary skin biopsy (or, less commonly, on liver or muscle

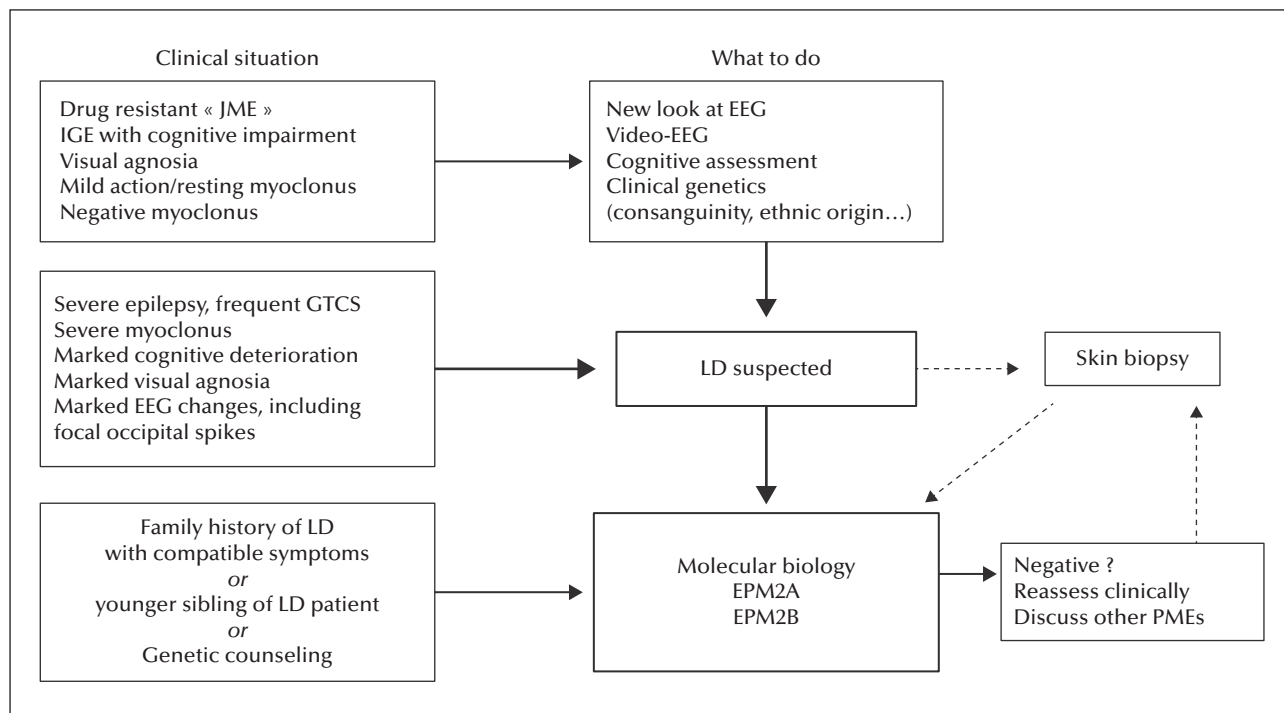


Figure 11. Management of patients with LD: diagnosis.

biopsy) used to be the definitive diagnostic tool, but requires a highly trained pathologist; it may still render services as a supplementary element in favour of a diagnosis of LD when genetic testing is not available, or not entirely conclusive.

– The confirmation of diagnosis is nowadays provided by the demonstration of a pathogenic mutation in both alleles of one of the *EPM2* genes, with presence of heterozygous mutations in each of the clinically unaffected parents.

The diagnosis of LD can be made, or suspected, in various clinical settings, with, in our opinion, three typical situations:

At an advanced stage of the condition, in very typical patients and/or families who were not diagnosed earlier due to a lack of local expertise resulting from the rarity of LD in some settings, or due to the absence of molecular biological tools, e.g. in rural Africa (Traoré *et al.*, 2009). In such cases, genetic testing will show the genetic subtype, and contribute to the worldwide ‘map’ of mutations found in *EPM2A* and *EPM2B*.

A PME or LD should enter the differential diagnosis for any adolescent with a diagnosis of IGE, JME, or photosensitive epilepsy with not only failure to respond to AEDs, but also a condition that appears to worsen. In such patients, a thorough re-evaluation of the clinical data shows that the patient has resting and action myoclonus, often well-controlled/masked

by medication, such as valproate (VPA), or, characteristically, negative myoclonus (Genton *et al.*, 2012), visual agnosia, and prominent, specific EEG changes. The search for a mutation in the known LD genes is warranted to confirm the diagnosis.

In other radically different situations within the context of a family with a confirmed diagnosis of LD in one of its members. Although LD is highly unlikely in asymptomatic adults, such adults will want to know whether they carry a heterozygous mutation; in younger asymptomatic siblings, LD may still manifest, and the emergence of “preventive” treatment strategies may render the identification of the mutation useful in this context. As the pathogenic mutations have already been characterized in the proband, the cost of the investigations for other family members will be lower, and the importance of detecting preclinical cases will also be known.

It is evident that the diagnosis of LD should not be given to the family before a final and definitive confirmation, however, once it is confirmed, it should not be kept from the caregivers. Presenting the diagnosis to patients themselves is not advisable, as there is little hope to offer. Moreover, they may be too young or affected to understand the implications; at an early stage, some may still be able to understand and gain access, e.g. on the internet, to detailed information on LD, which might cause severe depression. In our

experience, a diagnosis of 'severe, drug-resistant epilepsy' will satisfy the patient's curiosity and still offer some hope and a reason for accepting help. However, if significant progress occurs in the near future, this global attitude may have to change, for example, in order to accept pathogenetically-founded therapies.

The consequences of a diagnosis of LD for the family of the proband should not be underestimated. Time should be devoted to the change expected for patients and their families (see below), but also to counselling regarding the following topics:

- The legitimate feelings of guilt and resentment must be alleviated with a few simple statements, that the inheritance is bilateral (*i.e.* the disease does not come specifically from the father or from the mother, but from both sides), that the disease results from a very uncommon co-occurrence of abnormal genes (even in consanguineous marriages), and that persons with a single abnormal gene will not have the disease.

- The main concern is the possible occurrence of LD in other family members. The risk can be practically excluded in older, fully asymptomatic siblings, but cannot be excluded in younger asymptomatic siblings nor, *a fortiori*, in as yet unborn siblings, if the parents are young enough to have other children. Whether siblings, especially younger ones, should be subject to molecular screening is still debatable. There is no reliable, recommended treatment, in pre-symptomatic LD cases, to prevent or delay the appearance and progression of symptoms. However, new perspectives are opening up with new possible treatments. Knowledge of the type of causative mutation that affects the proband allows precise counselling in this matter. Having provided all the relevant information, the clinician in charge of the patient will come to an agreement with the family and obtain their informed consent for all the procedures performed on non-affected family members. If the family shows reluctance towards genetic testing, a simple EEG recording for a younger sibling may provide information on the potential risk of LD, as EEG changes may precede the first clinical symptoms by many years (Van Heycoptenham & De Jager, 1963). Concerning future pregnancies for the parents of a patient with LD, the risks are known (LD: $\frac{1}{4}$; transmission of one pathogenic gene: $\frac{1}{2}$; no risk: $\frac{1}{4}$), and prenatal screening can also be proposed.

- The risk of carrying an abnormal gene and transmitting the condition to other generations is also a major concern in families with LD. We have no experience of patients with LD having children. Their siblings or parents (when they plan to have children with another spouse), as well as other family members, may benefit from molecular screening in order to assess the presence or absence of the pathogenic gene(s) found in the proband.

An important aspect of diagnosis and genetic counselling is financial; insurance and conditions of reimbursement (or, more basically, the availability for procedures, tests, and medications) differ greatly between countries and social systems. Families should always be informed about the costs involved; searching for a mutation is expensive, but the simple screening for a known mutation is less costly. Similarly, the regulations covering genetic diagnosis, screening, and counselling may differ between countries, and clinicians should always conform to local laws. Obtaining informed consent for the successive steps of diagnostic and screening procedures is a minimum requirement.

Management: treatment and social support

In this article, new therapeutic proposals are not discussed since these are covered in detail in the last article (Minassian, 2016). We shall focus on the medical and social measures that are available for the care of present-day LD patients (*figure 12*). It is clear that health systems vary greatly between countries, particularly between affluent and less developed societies. Moreover, in many developed countries, health coverage is not equal; some patients (*e.g.* public sector employees and their families) may benefit from a whole range of possibilities, while others (*e.g.* those informally employed in the private sector) do not. It is also clear that patients with LD should benefit from the maximum possible help provided by the available health system. LD is a rare condition which cannot be considered as a public health problem (as costs are limited by the small number of affected persons), but it has major consequences for patients and caregivers, and society should show solidarity and provide all available support.

With regards to treatment, AEDs only have a partial effect on myoclonus and seizures, and there are none that have a major influence on the progression of cognitive and behavioral symptoms. Patients typically receive an AED, usually valproic acid, after the first generalized tonic-clonic seizure (GTCS). This is usually effective in suppressing, for some time, most GTCS, the symptoms associated with photic sensitivity, and some of the myoclonus. There are two unusual effects, which should lead to an early diagnosis of LD: first, the EEG shows rapidly increasing, permanent interictal changes, including focal occipital spikes, despite the apparent clinical remission; second, the patients develop negative myoclonus, which becomes prominent before the more characteristic myoclonic jerks. Other AEDs are used during this stage: lamotrigine (LTG) is not very advisable in the context of a myoclonic epilepsy, but may help transiently; phenobarbital (PB) and primidone (PRM) are

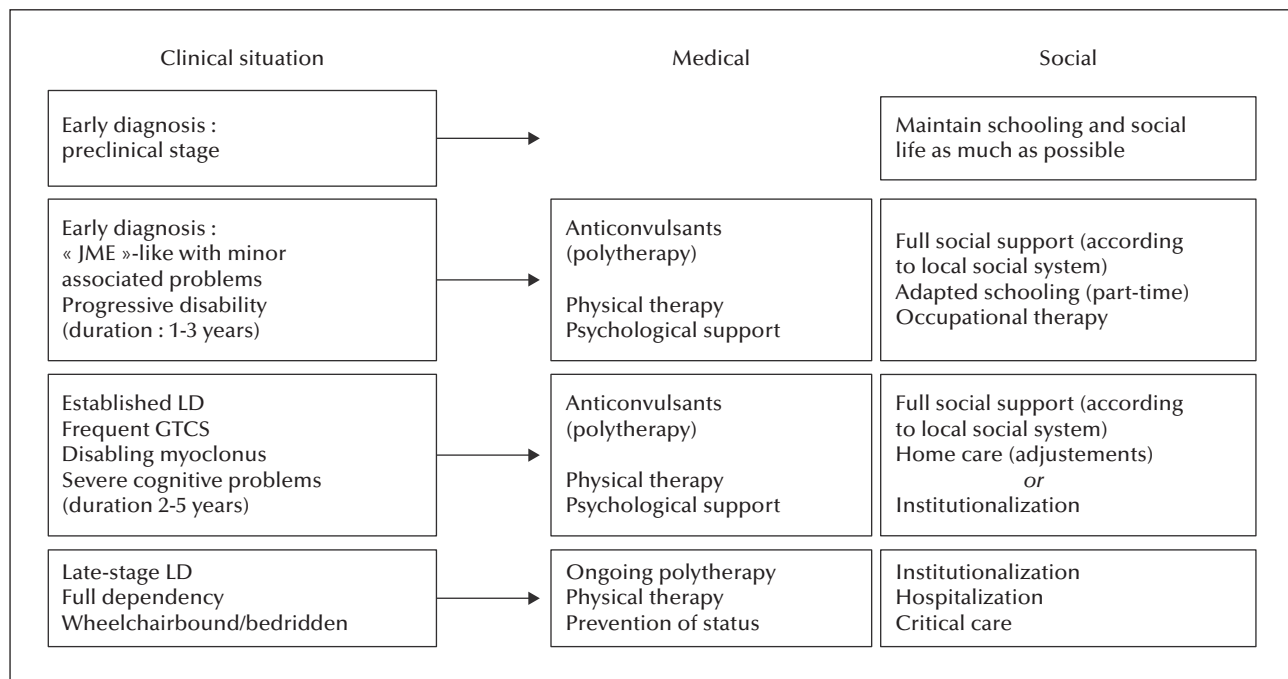


Figure 12. Management of patients with LD: medical and social treatment.

effective, but are often used at high doses and their cognitive effects are added to those of the condition; and levetiracetam (LEV) is increasingly used early for adolescents with IGE, hence in LD cases, even before confirmation of the diagnosis. Other helpful drugs include topiramate (TPM) and zonisamide (ZNS), which both have marked antimyoclonic effects in some patients. Additional relief can be obtained, often transiently, with ethosuximide, felbamate, methsuximide, and benzodiazepines (BZD). The latter (usually clobazam, clonazepam, and diazepam) should be used with care since there is a marked initial effect followed by quick tolerance. Finally, there have been two recent single case reports of rather dramatic beneficial effects of perampanel (Schorlemmer *et al.*, 2013; Dirani *et al.*, 2014), and a larger study is presently underway.

With such a severe condition, the paradoxical aggravating effect of some AEDs may be difficult to pinpoint. There is no evidence that carbamazepine (CBZ), oxcarbazepine (OXC), phenytoin (PHT), eslicarbazepine, gabapentin, pregabalin, vigabatrin, or lacosamide are of any benefit. Often, withdrawal of one of these AEDs (especially CBZ or OXC) will bring some relief. However, we have experienced cases in which status epilepticus responds well to phenytoin loading. Phenytoin should not, however, be kept as maintenance medication subsequent to arresting the status. With the progression of LD, AED treatment progresses to polytherapy, with a combination of several of the drugs quoted above (with the exclusion of LTG);

the commonly used combinations are VPA+TPM or ZNS or LEV, with an additional BZD, a 3- to 5-drug combination being fairly common; one can switch between BZD when tolerance occurs. In case of severe, transient aggravation, with serial seizures or status epilepticus, there should be no abrupt changes to the usual regimen (except for the interruption of a potentially aggravating AED), and IV BZD should be used, as well as, for a limited period, IV PB or PHT. In our recent experience, the final progression of the disease no longer includes refractory status epilepticus, but more commonly involves non-specific complications, infectious or otherwise, in bedridden and demented patients. Thus, despite their lack of influence on the overall evolution of the disease, modern AEDs have partly changed the outcome, which is apparently no longer accompanied by severe, formerly often terminal, episodes of refractory convulsive status epilepticus.

In LD, social support is at least as important as medical treatment. Psychological support can be provided by patient organisations and there are several which are specifically devoted to LD (table 2). Individual patients should also receive professional psychological support during the early stages of the condition. Physical therapy aims at maintaining a good overall muscular condition and at preserving ambulation, for as long as possible.

At the onset of seizures, patients are usually in secondary school and experiencing increasing difficulties

Table 2. Some web-based family support organizations

| Name | Site/address | Service |
|---|---|--|
| Association France-Lafora | http://www.Lafora.org 16, rue Amaudrut F- 53000 Laval | Patient organisation Promotes self-help and research Collects funds for research |
| A.I.L.A. Associazione Italiana Lafora | http://www.Lafora.it | Patient organisation Promotes exchanges between families Identifies centres for diagnosis and treatment Collects funds for research |
| Chelsea's Hope Lafora Children Research Fund | http://www.chelseashope.org | Family-based organisation Connects families with LD worldwide Collects funds for research |

with academic requirements. In order to enable them to maintain social contacts, it is best to maintain schooling for as long as possible, while negotiating with teachers about academic performance. However, this cannot be kept up for extensive periods. Some families will choose to keep patients at home with the best possible environment; this often requires adaptive measures, avoiding the use of stairs, setting up the patient close to the bathroom, and providing 24-hour presence at home with the help of a health professional to check medications. Other families will seek a specialized institution where patients are kept with other epilepsy patients in the same age group, with some amount of education and social activities. Re-evaluation at the specialized neurological department can be organized at 6- or 12-month intervals, with acute admissions in the event of complications, often due to intercurrent diseases (e.g. febrile infections) and/or worsening of epilepsy.

The latter period is characterized by increasing dependency as the patient becomes wheelchair bound and, later, bedridden. According to local availability and the wishes and capacities of the caregivers, the patient is maintained at home, institutionalized or hospitalized. There should always be a connection between the reference specialized epilepsy team and the local caregiving structure.

Conclusion

For patients with LD, the present state of possibilities includes a logical, effective approach to diagnosis, and the rational use of all available tools to help both the patients and their families. We hope that the management of LD patients will undergo a profound change in the near future, when effective, pathogenetically-oriented treatments become available. □

Disclosures.

None of the authors have any conflict of interest to disclose.

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SCARB2/LIMP2 deficiency in action myoclonus-renal failure syndrome

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ABSTRACT – Action myoclonus–renal failure syndrome (AMRF) is an autosomal recessive progressive myoclonus epilepsy (PME) associated with renal dysfunction that appears in the second or third decade of life and that is caused by loss-of-function mutations in the SCARB2 gene encoding lysosomal integral membrane protein type 2 (LIMP2). Recent reports have documented cases with PME associated with SCARB2 mutations without renal compromise. Additional neurological features can be demyelinating peripheral neuropathy, hearing loss and dementia. The course of the disease is relentlessly progressive. In this paper we provide an updated overview of the clinical and genetic features of SCARB2-related PME and on the functions of the LIMP2 protein.

Key words: progressive myoclonus epilepsy, myoclonus, cerebellar syndrome, photosensitivity, SCARB2, LIMP2, lysosome

In 1986, Andermann *et al.* reported four French Canadian patients from three sibships with a condition characterized by the appearance of tremor in the fingers and hands and proteinuria at 17–18 years of age. Severe progressive action myoclonus, dysarthria, ataxia, infrequent generalized seizures, and renal failure ensued between 19 and 23 years of age. Despite severe neurological disability, due mainly to action myoclonus, intelligence remained normal in all four patients. They labelled this condition ‘action

myoclonus-renal failure (AMRF) syndrome’. Since this first description, it has been pointed out that the neurological picture was not caused merely by a metabolic encephalopathy due to renal failure, but rather was the result of a pathophysiological process that appeared to involve primarily both the brain and the kidneys (Andermann *et al.*, 1986). This syndrome was not recognized prior to the advent of dialysis and renal transplantation because of its rapidly fatal course if renal failure is untreated.

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Discovery of *SCARB2* as the causative gene for AMRF

Three unrelated Australian families with a single proband were used to identify *SCARB2* as the causative gene for AMRF. Case A was of Turkish-Cypriot origin; her parents were first cousins. The ancestors of families B and C came from different regions of Britain and no inbreeding loops were known for either family. Case C was deceased, but stored brain tissue in paraffin blocks was available for DNA extraction.

A critical region of 5.3 cM (equivalent to 6.6 Mb) on chromosome 4 (4q13-21) was narrowed down by identifying an overlap of regions in which Case A and Case C were homozygous by descent. The region was reduced slightly in size by excluding a region in family B in which the affected individuals shared a segment identical by descent with an unaffected sibling.

The critical region on chromosome 4 contained 66 annotated genes of which approximately half are expressed in both the brain and kidney. In order to prioritize candidate genes for sequencing, it was hypothesized that the mRNA of the causative gene would be downregulated in affected subjects, possibly because of mutations causing RNA instability or removal by the process of nonsense-mediated decay. RNA from lymphoblastoid cell lines derived from two living affected subjects and from a healthy gender-matched sibling of each (families A and B) was analysed with Affymetrix U133 Plus2 arrays to look for RNAs with decreased abundance in the affected individuals in comparison to their unaffected sibling. RNA analysis was confined to the probe sets in the 6.6 Mb chromosome 4 region, defined by homozygosity mapping. *SCARB2* (*Scavenger Receptor B2*) emerged as a possible candidate gene. The amount of mRNA was reduced approximately 2-fold in the affected subjects compared to their healthy siblings.

The protein-coding regions of the *SCARB2* gene were analysed for mutations by direct Sanger sequencing. A homozygous splice-site mutation (c.1239+1G>T) in Case A was identified. RT-PCR analysis showed that this mutation leads to retention of intron 10 and the subsequent insertion of 20 amino acids, and premature truncation of the protein at residue 433. A homozygous mutation in Case C (c.435_436insAG W146SfsX161) was identified resulting in a frameshift predicted to truncate the *SCARB2* protein to 160 amino acids. Case B was a compound heterozygote, carrying two different mutations in the *SCARB2* gene: a frameshift mutation (c.296 delA N99IfsX34) predicted to shorten the protein to 131 amino acids and a splice site mutation c.704+5G>A. These findings confirmed *SCARB2* as a causative gene for AMRF (Berkovic *et al.*, 2008).

The *SCARB2* gene encodes a 478 amino acid glycoprotein located in lysosomal membranes in a range of tissues including the brain and kidney. The function of *SCARB2* is not well understood, but it is thought to play a role in the biogenesis and maintenance of endosomal and lysosomal compartments. The human and mouse *SCARB2*/Limp2 proteins share 85 per cent amino acid identity.

Founder mutations of *SCARB2* in the French-Canadian and Scottish populations

A number of families were identified from Quebec, Canada, to have probands clinically diagnosed with AMRF. The Quebec population is known to have a high degree of consanguinity which results in a higher incidence of recessive disorders than other parts of the world. Molecular analysis of *SCARB2* found that all but one of the Quebec cases of AMRF were homozygous for the mutation c.862C>T, Q228X. Haplotype analysis using microsatellite markers on affected members and carriers was employed to determine whether *SCARB2* Q228X was a founder mutation inherited from a common ancestor. A shared haplotype spanning 0.6 cM encompassing the *SCARB2* mutation indicated the presence of a founder Q228X mutation for AMRF in the Quebec population. One family from Quebec was found to carry a second *SCARB2* mutation, c.1197+3insT.

Two subjects with AMRF from Scotland were found to be homozygous for the *SCARB2* mutation c.435_436insAG (W146S fs X161). This mutation is the same as that found in an Australian patient (Case C above) and in a Canadian patient (not French-Canadian). Haplotype analysis of the Australian, Canadian and the two Scottish cases, which were not previously known to be related, showed a shared haplotype of 0.6 cM, indicating that the mutation was inherited from a shared ancestor. The Scottish population, therefore, also contains a *SCARB2* founder mutation, W146S fs X161, causing AMRF.

Further cases of AMRF due to *SCARB2* mutations

Given that demyelinating neuropathy is seen in the mouse model deficient in functional *SCARB2*/Limp2 (Gamp *et al.*, 2003), a patient from the USA presenting with PME and asymptomatic neuropathy was hypothesized to carry a *SCARB2* mutation. This patient was found to be a compound heterozygote, carrying two different AMRF-causing mutations: the Q228X (Quebec) mutation on one chromosome and the

second Quebec mutation c.1187+3insT on the other (Dibbens *et al.*, 2011). Since renal dysfunction is usually seen in AMRF, this patient is now undergoing tests for kidney function. Further cases of AMRF from Argentina, Turkey, Portugal and Spain (Balreira *et al.*, 2008; Perandones *et al.*, 2012) have been found to be caused by mutation of SCARB2, suggesting the syndrome is likely to be found worldwide.

Clinical features of action myoclonus renal failure

Following the initial report by Andermann *et al.* (1986), several studies confirmed that the predominant clinical manifestations of AMRF are progressive myoclonus epilepsy (Minassian *et al.*, 2016) and renal failure (Badhwar *et al.*, 2004; Vadlamudi *et al.*, 2006; Balreira *et al.*, 2008; Perandones *et al.*, 2012). Disease onset is typically in the late teens or early twenties, and the neurological features can be seen before, after, or simultaneously with the renal features. The neurological picture may present as a tremor, which is typically first noted in the fingers and hands, present at rest and exacerbated by fine motor activities. The tremor can later involve the head, trunk, lower extremities and sometimes the tongue and voice. As the disease progresses, involuntary spontaneous action-activated myoclonic jerks are seen, as well as asynchronous involuntary spontaneous myoclonic jerks at rest. A reflex myoclonus which is sensitive to touch on the extremities is also present. Action myoclonus, refractory to antimyoclonic drugs, is the most debilitating feature of the disease and, in the final stages, it renders the patients bedridden or wheelchair-bound with lap, trunk and leg belts. Diurnal or nocturnal generalized tonic-clonic seizures occur in the majority of patients. Badhwar *et al.* (2004) reported that the convulsive seizures start with a generalized clonic phase with preserved consciousness proceeding to unconsciousness with tonic-clonic features. Antiepileptic drugs can control convulsive seizures without affecting ongoing active myoclonic jerks. Other common features appearing during the course of the disease include ataxia and dysarthria due to cerebellar dysfunction. Remarkably, despite the progression and severity of the neurological picture, cognitive function is preserved or only slightly affected until the final stages of the disease.

A demyelinating peripheral neuropathy has been reported in a number of patients (Dibbens *et al.*, 2011; Hopfner *et al.*, 2011; Rothdach *et al.*, 2001), while electrophysiological findings indicating a predominantly axonal neuropathy have been observed in a patient without clinical evidence of involvement of the peripheral nervous system (Badhwar *et al.*, 2004). Auditory

defects ranging from abnormal brainstem auditory evoked potentials without clinical expression to severe hearing loss have been reported by Perandones *et al.* (2012, 2014) in a patient and two siblings with SCARB2 mutations and clinical features of AMRF. Interestingly, both these latter neurological manifestations correlate with the phenotype of the *Limp2* knock-out mice whose neurological alterations consist of deafness and peripheral neuropathy, without features of progressive myoclonus epilepsy (Gamp *et al.*, 2003). Finally, a dilated cardiomyopathy has been described in two patients (Hopfner *et al.*, 2011).

Renal involvement in AMRF is heralded by the appearance of proteinuria that can relentlessly progress to a nephrotic syndrome and end-stage renal disease, requiring dialysis or renal transplantation. Detection of proteinuria usually occurs around the age of 20, although onset in childhood has been reported (Badhwar *et al.*, 2004). No correlation has been observed between the ages of onset of proteinuria and tremor, nor between renal failure and onset of myoclonus.

The absence of renal involvement in PME associated with SCARB2 mutations has also been described. In 2009, Dibbens *et al.* reported SCARB2 mutations in 5 of 41 cases considered clinically to be 'Unverricht-Lundborg disease (ULD)-like' (Dibbens *et al.*, 2009). The patients had disease onset between 14 and 26 years of age, with no evidence of renal failure during 5.5 to 15 years of follow-up, although one of them had slight proteinuria in the final stage of the disease. Death ensued in all five patients (the only surviving patient at the time of the report died later). Since this initial report, other cases have been reported and the clinical features of PME without renal failure associated with SCARB2 have been described (Rubboli *et al.*, 2011; Guerrero-López *et al.*, 2012; Higashiyama *et al.*, 2013; Fu *et al.*, 2014; Zeigler *et al.*, 2014). Features seen in these patients included a variable severity of epilepsy: from uncontrolled seizures or status epilepticus with prominent photosensitivity in patients with adolescent onset, to infrequent or no major seizures in patients with a more delayed onset (Rubboli *et al.*, 2011), late onset in adulthood (Higashiyama *et al.*, 2013; Fu *et al.*, 2014), and the occurrence of dementia (Fu *et al.*, 2014). These findings suggest that SCARB2 mutations in patients with PME without renal complications might not be rare and that SCARB2 gene mutations should therefore be evaluated even in the absence of renal involvement.

The course of AMRF is fatal with relentless progression of neurological deterioration and increasing severity of myoclonus and renal impairment leading to death usually within 7 to 15 years after disease onset, due to renal failure, aspiration pneumonia, or septicemia with multiorgan failure.

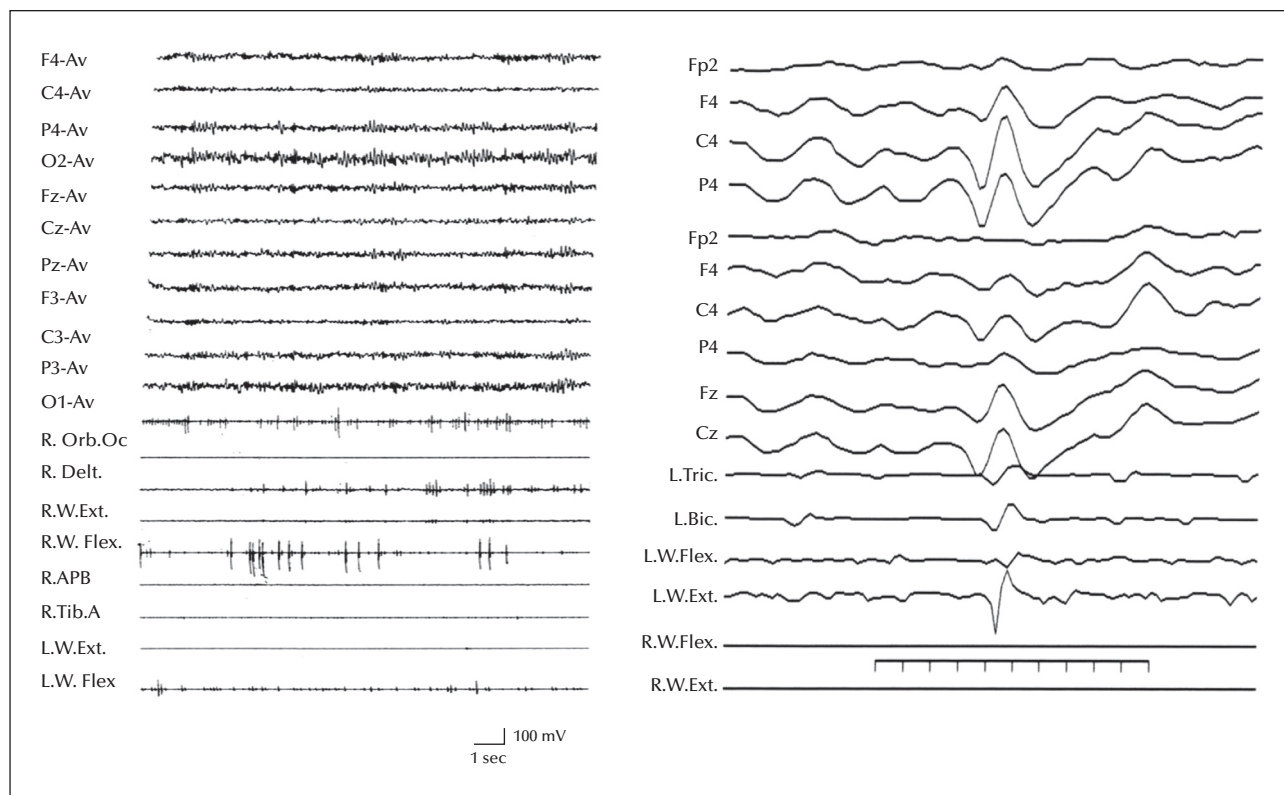


Figure 1. Polygraphic recording in a 32-year-old patient with PME without renal failure associated with SCARB2 mutation. Left panel: the recording shows preserved EEG background activity and erratic myoclonic jerks at rest without overt EEG correlate. Right panel: back-averaging triggered by myoclonia in the left wrist extensor reveals a cortical spike at the contralateral centroparietal electrodes.

EEG and brain imaging

EEG and polygraphic recordings show generalized epileptiform abnormalities that at onset may resemble epileptic activity observed in idiopathic generalized epilepsy (Badhwar *et al.*, 2004; Rubboli *et al.*, 2011). Background activity is preserved at disease onset, slowing progressively over the years. In photosensitive patients, intermittent photic stimulation can trigger bursts of generalized spike-polyspike-wave discharges, often associated with massive myoclonic

jerks that can evolve to myoclonic seizures. Polygraphic recordings show action myoclonus and erratic myoclonic jerks at rest, incessantly associated with contralateral central spikes (*figure 1*). Back-averaging analysis of EEG discharges triggered from myoclonic jerks can reveal a cortical spike at the centroparietal electrodes (*figure 1*). Surface EMG recording of the fine tremor in the upper limbs showed quasirhythmic EMG bursts at a frequency of 12-20 Hz (*figure 2*). Analysis of the EEG-EMG relationship by coherence spectra of the tremor demonstrated a pattern consistent with

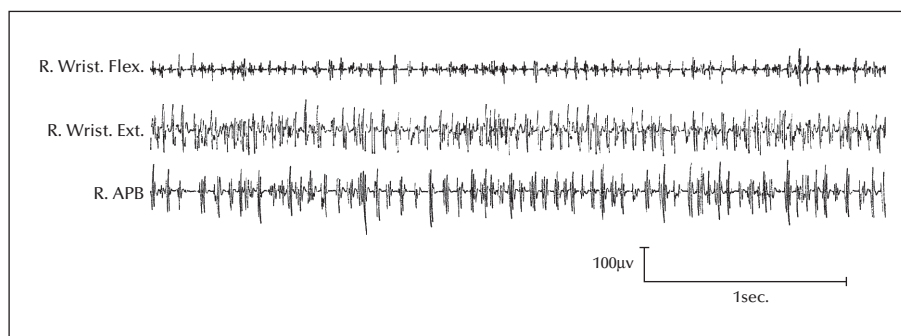


Figure 2. Surface EMG recording of the tremor in the upper limbs shows quasirhythmic EMG bursts at a frequency of 12-20 Hz.

a rhythmic myoclonic phenomenon of cortical origin, as in 'cortical tremor' (Rubboli *et al.*, 2011).

Brain imaging studies are usually unremarkable or display diffuse cerebral atrophy, often associated with cerebellar atrophy.

Histology

Widespread deposition of abnormal, extraneuronal brown pigment in the brain, with no neuronal loss or significant gliosis, has been reported in AMRF patients (Andermann *et al.*, 1986; Badhwar *et al.*, 2004), and more recently in two patients with PME without renal failure associated with SCARB2 mutations (Fu *et al.*, 2014). Based on the staining characteristics, it has been suggested that the pigment consists of lipofuscin-like oxidized lipid or proteolipid (Badhwar *et al.*, 2004). The deposits of pigment granules are extraneuronal, in astrocytes or in the extracellular space, especially in the cerebellar and cerebral cortices without any increase in intraneuronal lipofuscin (Badhwar *et al.*, 2004; Fu *et al.*, 2014). Interestingly, the patients without renal failure reported by Fu *et al.* (2014) also showed neurodegenerative changes, such as neuronal loss and gliosis in the brain, including the pallidoluysian and cerebello-olivary systems and the spinal cord. The authors speculate that the neuronal loss and gliosis in the pallidoluysian and cerebello-olivary systems may be responsible for the patients' involuntary movements and cerebellar dysfunction. Moreover, degenerative changes observed in the upper and lower motor neuronal systems, as well as in the dorsal root ganglia, strongly suggested that both the motor and sensory neuronal systems were also involved in the disease process.

Renal biopsy specimens have shown extensive tubular abnormalities with isometric vacuolization in distal and collecting tubules, the presence of granular material in cortical tubules without inflammatory infiltration (Chaves *et al.*, 2011), and focal glomerulosclerosis, with features of collapsing glomerulopathy (Badhwar *et al.*, 2004; Berkovic *et al.*, 2008).

Differential diagnosis

Differential diagnosis of AMRF primarily concerns other PMEs without dementia. At onset, since they share various clinical features, the correct diagnosis may be difficult on clinical grounds, particularly when renal impairment has not yet appeared or has not been diagnosed. Unverricht-Lundborg disease (ULD) is the paradigmatic example of PME with action myoclonus, ataxia, generalized seizures, and preserved intellect. The clinical suspicion of AMRF should be raised when

a fine postural tremor in the upper limbs, which is generally not present in ULD, is detected on neurological examination at disease onset. Other rare causes of PME without dementia that should be considered in the differential diagnosis include sialidoses, distinguishable by macular cherry red spots, and PME, caused by mutations in PRICKLE1, the onset of which is generally earlier than that in AMRF and associated with early ataxia.

Concluding clinical remarks

The molecular analysis of patients clinically diagnosed with AMRF showed that mutations in SCARB2 have been identified in many patients, confirming that the likelihood of identifying a SCARB2 mutation in a patient presenting with PME and renal complications is fairly high. The French Canadian and Scottish populations have been shown to have founder mutations in SCARB2 causing AMRF. Different SCARB2 mutations have now been found in patients in many different countries, suggesting that SCARB2-associated PME is a world-wide disorder that is presently under recognized. Onset of PME due to mutations in SCARB2 occurs in teenagers or young adults, and diagnosis is important in terms of providing counselling for the patient and family, particularly as the prognosis is worse than for classic ULD. Being informed about the carrier status of family members is particularly relevant in terms of the prevention of future disease in populations of known high consanguinity. In addition, in AMRF patients, an early diagnosis is of utmost importance as renal dysfunction can cause premature death in childhood or adolescence. Treatment with kidney dialysis or renal transplantation has been shown to prolong life by at least 10 years (Badhwar *et al.*, 2004). Furthermore, SCARB2 mutations in patients with PME in the absence of renal complications are probably not rare, therefore mutations of the SCARB2 gene should be considered in undiagnosed PME without renal involvement.

The LIMP-2 protein

The lysosome is the major degradative compartment of the cell. Its limiting membrane fulfils multiple functions, such as acidification of the interior, sequestration of active lysosomal enzymes, and transport of degradation products from the lysosomal lumen to the cytoplasm (Saftig & Klumperman, 2009). The lysosomal membrane contains several highly N-glycosylated proteins, the functions of which remain largely unknown (Eskelinen *et al.*, 2003).

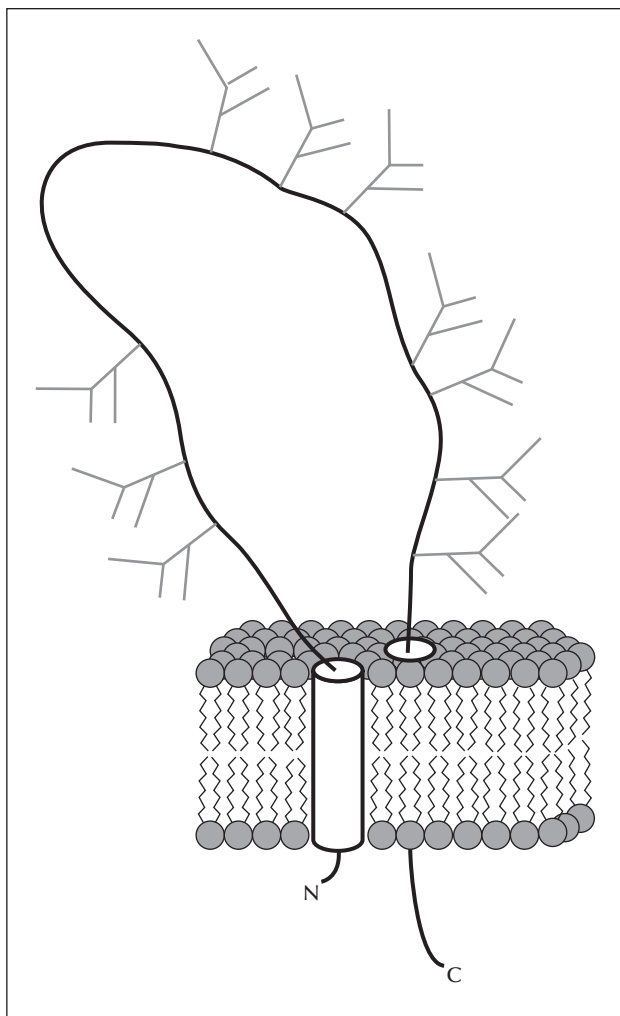


Figure 3. Topology of the lysosomal membrane protein LIMP-2. LIMP-2 is a heavily N-glycosylated type III transmembrane protein with N- and C-termini located in the cytoplasm. The coiled-coil domain, which is necessary for GC binding, is located within the luminal domain of LIMP-2.

LIMP-2, a major component of the lysosomal membrane

LIMP-2 (lysosomal integral membrane protein type 2), also known as SCARB2, is an abundant, highly glycosylated lysosomal membrane protein and belongs to the CD36 family (Calvo *et al.*, 1995). All family members share a common topology, transverse the membrane twice with an N-terminal transmembrane domain, a large luminal domain, and a second membrane-spanning domain preceding a 20-amino-acid cytoplasmic tail at the C-terminus (*figure 3*). Whereas in most tissues LIMP2 is found in lysosomes, other members of the CD36 superfamily are localized and function at the cell surface.

LIMP2 is a ubiquitously expressed protein with highest expression in the liver and spleen (Tabuchi *et al.*, 1997). It has a molecular weight of about 74 kDa, which includes a 54-kDa polypeptide backbone of 478 amino acids. The luminal domain contains 10 to 11 putative glycosylation sites (*figure 3*). The degree of the complex glycosylation of LIMP-2 depends on the species, tissue, and cell type. A leucine-isoleucine motif, within the C-terminal cytoplasmic tail, interacts with the heterotetrameric adaptor-complex 3 (AP3) and has been proposed to be responsible for the lysosomal localization of LIMP-2 (Honing *et al.*, 1998). In addition, based on antibody uptake experiments, a recycling of LIMP-2 between lysosomes and the plasma membrane has been described (Akasaki *et al.*, 1994). Furthermore, it has been demonstrated that distinct phosphatidylinositol 4-kinases influence the trafficking of LIMP-2 (Jović *et al.*, 2012).

Overexpression of LIMP-2 leads to altered cellular membrane trafficking

Overexpression of LIMP-2 causes an enlargement of early and late endosomes/lysosomes, which has not been observed for other abundant lysosomal membrane proteins. LIMP-2 overexpression impairs the endocytic membrane traffic out of these enlarged compartments and leads to an accumulation of cholesterol in these vacuole-like structures. Co-transfection of LIMP-2 and the dominant-negative form of Rab5b inhibits the formation of enlarged vacuoles, suggesting that Rab5b function is necessary for the formation of such vesicles (Kuronita *et al.*, 2002). Mutation experiments suggest that the N-terminal transmembrane and proximal luminal domains of LIMP-2 are essential for the generation of the enlarged vesicular structures (Kuronita *et al.*, 2005). These findings support the idea that LIMP-2 plays a role in the biogenesis and maintenance of the endo-lysosomal system.

LIMP-2 receptor functions

Similar to CD36, LIMP-2 appears to display a role as a multifunctional receptor at the plasma membrane. By using LIMP-2-GST fusion proteins and labelled thrombospondin, an interaction between both proteins was shown (Crombie & Silverstein, 1998).

In addition, LIMP-2 appears to be a cellular receptor for enterovirus 71 (EV71) and the coxsackie virus A16 (CVA16), which are most frequently associated with hand, foot and mouth disease (HFMD) (Yamayoshi *et al.*, 2009). Although HFMD is considered to be a mild infection, it can progress to a severe neurological disease, associated with fatal encephalitis, aseptic meningitis, and acute flaccid paralysis.

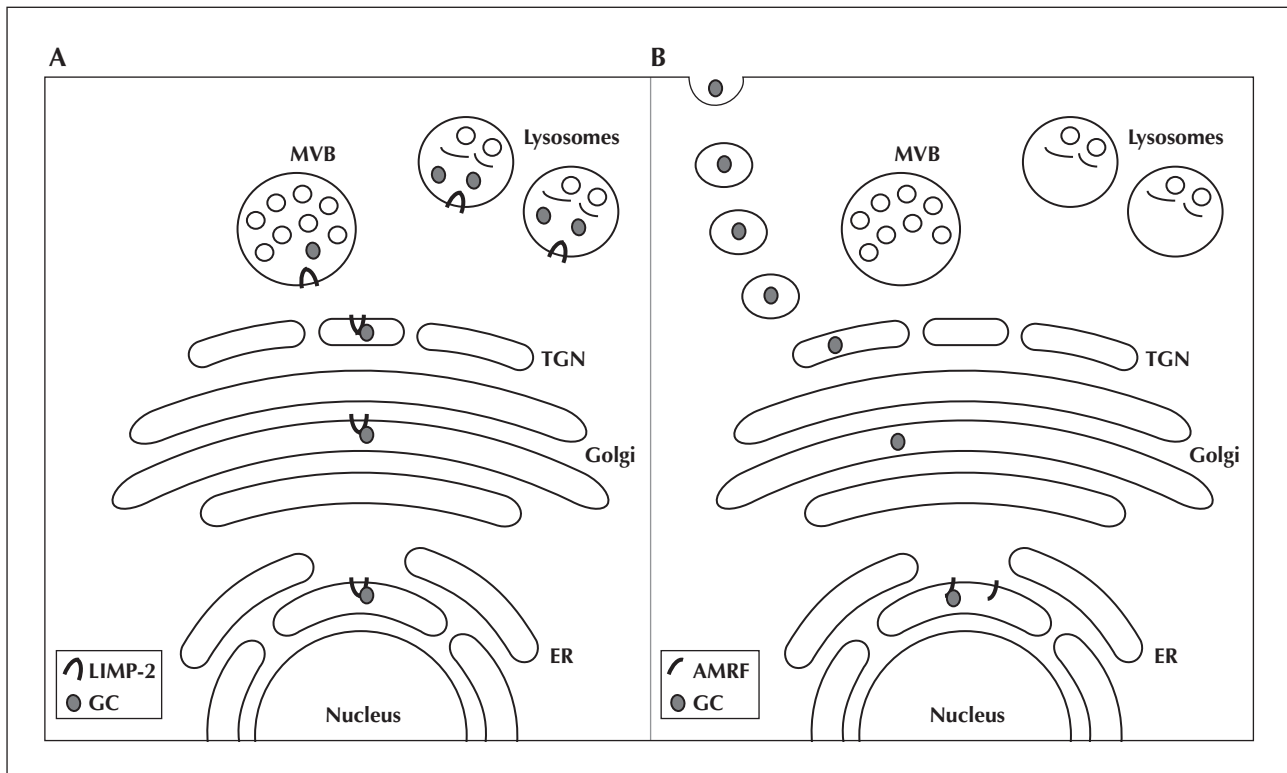


Figure 4. LIMP-2-dependent targeting of GC to the lysosome. (A) In wild-type cells, GC is transported to the lysosome by LIMP-2. After binding between LIMP-2 and GC in the ER, GC leaves the receptor-ligand complex in the ER and moves via the Golgi apparatus to multivesicular bodies and lysosomes, where they dissociate due to the acidic pH. (B) All clinical AMRF-causing mutations described for LIMP-2 thus far have led to retention of the mutated protein in the ER. Thus, in cells derived from AMRF patients, GC is retained in the ER and/or secreted to the extracellular space, depending on the effect of the AMRF-causing LIMP-2 mutation on binding to GC.

It is well established that LIMP-2 has another important receptor function. It acts as a receptor for the lysosomal delivery of acid hydrolase β -glucocerebrosidase (GC) (Reczek *et al.*, 2007). Mutations in the gene encoding this enzyme have been shown to cause Gaucher Disease, the most common lysosomal storage disorder, which is due to lysosomal accumulation of the glycosphingolipid glucosylceramide. LIMP-2 binds the enzyme, involving a helical domain in the luminal domain, early in the endoplasmic reticulum and transports it all the way to the lysosome (Reczek *et al.*, 2007; Neculai *et al.*, 2013; Blanz *et al.*, 2015; Zunke *et al.*, 2016) (figure 4a). Due to the acidic pH of the lysosome, the LIMP-2 receptor and its ligand dissociate (Reczek *et al.*, 2007; Zachos *et al.*, 2012), leading to the active lysosomal enzyme. All analyzed clinical AMRF-causing mutations described for LIMP-2 thus far have led to retention of the mutated protein in the endoplasmic reticulum (figure 4b). However, the binding to GC is differentially affected by the various AMRF-causing mutations (Blanz *et al.*, 2010). It will be interesting to study whether the lack of lysosomal transport of GC contributes to the pathology of AMRF syndrome. Interestingly, the structure of the extracellular domain of

LIMP-2 was recently revealed, showing an exposed helical bundle where GC binds. In addition, a cavity within the protein suggests a possible lipid transport function (Neculai *et al.*, 2013).

LIMP-2-deficiency in mice reveals major roles in the inner ear, kidney and myelinisation of peripheral nerves.

The different roles of LIMP-2 as a receptor may also contribute to the phenotype of mice engineered with a deletion of the mouse LIMP-2 gene. LIMP-2-deficient mice are characterized by the development of deafness, a unilateral or bilateral hydronephrosis, proteinuria, and a peripheral demyelinating neuropathy (Gamp *et al.*, 2003).

The development of deafness was indicated by deficits in acoustic startle responses, in brainstem-evoked auditory potentials, and a reduced endochondral potential. A massive decline of spiral ganglia in the cochlea, concomitant with that of the inner and outer hair cells, and a progressive atrophy of the stria vascularis are typical pathological changes which start

shortly after birth (Gamp *et al.*, 2003). Hearing loss is temporally linked to a loss of the potassium channel subunits KCNQ1/KCNE1 and the endocytic receptor megalin in the luminal surface membrane of marginal cells of the stria vascularis (Knipper *et al.*, 2006). A role of LIMP-2 in the regulation of the correct surface expression of these proteins through vesicular transport can be anticipated. This is also supported by the in-depth analysis of the development of a unilateral or bilateral hydronephrosis caused by an obstruction of the ureteropelvic junction. An impairment of the membrane transport processes is suggested by an abnormal accumulation of lysosomes in the epithelial cells of the ureter, adjacent to the ureteral lumen and a disturbed apical expression of uroplakin (Gamp *et al.*, 2003). It is speculated that the pathology in the urothelium leads to the obstruction of the urinary tract between the renal pelvis and the ureter. In addition to this obstruction, kidney functions are affected. Decreased osmolality and altered urine parameters in LIMP-2-deficient mice point towards renal dysfunction. The high quantity of albumin in the urine of LIMP-2 knockout mice may be explained by glomerular filtration damage. Both the hydronephrosis and subtle glomerular changes may explain the kidney pathology in these mice (Berkovic *et al.*, 2008). Interestingly, renin and LIMP-2 are co-regulated in renin-producing cells, although LIMP-2 does not play a role in the direct regulation of renin synthesis or release (Schmid *et al.*, 2013). The development of a peripheral demyelinating neuropathy in LIMP-2-deficient mice is an additional phenotypic hallmark and is most likely caused by a downregulation of peripheral myelin proteins (Gamp *et al.*, 2003). Interestingly, lysosomal enzymes are upregulated in LIMP-2-deficient Schwann cells, suggesting that peripheral myelin proteins are missorted and degraded in the lysosomal compartment. Finally, Berkovic and colleagues observed in the LIMP-2-deficient mice, intracellular inclusions in cerebral and cerebellar cortex neurons accompanied by hyperactivity and ataxic gait behaviour (Berkovic *et al.*, 2008). A recent study also revealed that the loss of LIMP-2 in the brain led to an almost complete loss of neuronal GC. This caused a substrate accumulation, followed by a secondary accumulation of neurotoxic α -synuclein. In cell-based assays, it was shown that an increased expression of LIMP-2 also led to a reduction in α -synuclein, suggesting a putative therapeutic role of LIMP-2 in synucleopathies (Rothaug *et al.*, 2014).

Miscellaneous functions

Additional studies analysing the LIMP-2 homologue in *Dictyostelium discoideum* (DdLIMP) revealed a role as an effective suppressor of the profilin-minus phenotype (Karakesisoglou *et al.*, 1999; Temesvari *et al.*,

2000). Profilin is a ubiquitous G-actin binding protein which is involved in multiple cellular processes, such as cytokinesis, phagocytosis, and micropinocytosis. Profilin-deficient *Dictyostelium discoideum* cells show defects in pinocytosis, macropinocytosis, exocytosis, secretion of hydrolases, and an increased rate of phagocytosis. Interestingly, the infection of mice deficient in LIMP-2 revealed that phagocytosis is also compromised. When such mice were infected with *Listeria monocytogenes* (LM), it was found that they were highly susceptible to infection and displayed defective macrophage activation (Carrasco-Marin *et al.*, 2011). This defect increased the amount of pathogen, which is able to escape to the cytosol, increased the production of early acute phase pro-inflammatory cytokines, and enhanced the ability of LM phagosomes to interact with MIIC vesicles. These experiments suggested that in concert with active Rab5a, LIMP-2 regulates the phagosomal fusion machinery of the late endosomes-lysosomes and the cytosolic levels of the pathogen (Carrasco-Marin *et al.*, 2011).

Conclusions

Although the exact molecular role of LIMP-2 in the various cellular events is still incompletely understood, a central role for this lysosomal membrane protein in health and disease has emerged. Future studies will help to further understand the role of LIMP-2 in various tissues, including the central nervous system. It will be of particular importance to link the multitude of cellular functions with the phenotypic alterations seen in knockout mice and the clinical presentations in human AMRF patients. This will most likely also shed light onto the hitherto poorly understood role of the endocytic pathway in the central nervous system and the development of progressive myoclonus epilepsy disorders.

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Neuronal ceroid lipofuscinoses

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ABSTRACT – The neuronal ceroid lipofuscinoses (NCL) are neurodegenerative conditions that associate cognitive decline, progressive cerebellar atrophy, retinopathy, and myoclonic epilepsy. NCL result from the excessive accumulation of neuronal and extraneuronal lipopigments, despite having diverse underlying biochemical aetiologies. Here we review the clinical presentation, pathophysiology and genetics of these conditions as well as the approach to diagnosis and management.

Key words: Haltia-Santavuori, Janský-Bielschowsky, Batten, Spielmeyer, progressive myoclonus epilepsies

The neuronal ceroid lipofuscinoses (NCLs) represent a heterogeneous group of genetically-determined neurodegenerative conditions that are characterized by a progressive decline of cognitive and motor capacities, retinopathy evolving into blindness, variable cerebellar atrophy, and myoclonic epilepsy, leading to significantly decreased life expectancy (Mitchison *et al.*, 1998; Santavuori *et al.*, 2000; Jalanko & Braulke, 2009; Warrier *et al.*, 2013). They are the most prevalent neurodegenerative disorders of childhood with an incidence in the USA estimated at 1.6-2.4/100,000, while in Scandinavian countries the incidence varies between 2-2.5/100,000 in Denmark, 2.2/100,000 in Sweden, 3.9/100,000 in Norway, 4.8/100,000 in Finland, and 7/100,000 in Iceland (Uvebrant & Hagberg, 1997; also reviewed in Mole *et al.*, 2011).

The first description in the medical literature was probably by Otto

Christian Stengel (1795-1890), a German physician who served in the mining community of Røros, Norway, between 1821 and 1882. He described a juvenile-onset disorder with blindness and progressive dementia (Stengel, 1826). In 1903, Frederick Eustace Batten (1865-1918), an English neurologist and paediatrician, described a similar clinical disorder and was the first to describe the neuropathology of cerebral degeneration with ocular macular changes in two members of a family (Batten, 1903). In 1905, Walther Spielmeyer (1879-1935), who had succeeded Alois Alzheimer as director of neurology and clinical psychiatry laboratory in Munich and Heinrich Vogt (1875-1936) reported a similar disorder (Spielmeyer, 1905; Vogt, 1905). At this time, juvenile neuronal ceroid lipofuscinosis was referred to as Batten-Spielmeyer-Vogt disease. Later, Jan Janský (1873-1921) and Max Bielschowsky (1869-1940)

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described a similar disorder, but with a “late-infantile” onset (Janský, 1908; Bielschowsky, 1913). This form came to be known as “Janský-Bielschowsky disease” or “late-infantile NCL”.

Hugo Kufs (1871-1955), from Leipzig, Germany, published four reports between 1925 and 1931 in which he described an adult-onset disease with similar pathological characteristics, but without the loss of vision that was so prominent in juvenile NCL and late-infantile NCL (Kufs, 1925). This came to be known as adult-onset NCL or Kufs disease. More recently, Matti Haltia (b. 1939) and Pirkko Santavuori (1933-2004), while investigating a child suspected to have GM1 gangliosidosis type II, concluded by identifying a novel type of NCL with early onset (Haltia et al., 1973a; Haltia et al., 1973b; Santavuori et al., 1973). Classic infantile NCL is also known as Haltia-Santavuori disease.

Traditionally, NCLs were classified according to the age at onset as: infantile (INCL), late-infantile (LINCL), juvenile (JNCL) and adult (ANCL), but were also known by their eponyms Haltia-Santavuori disease, Jansky-Bielschowsky disease, Batten-Spielmeyer-Vogt disease, and Kufs disease, respectively (*table 1* and *figure 1*). Moreover, the term ‘Batten disease’ was used in the literature to designate both the whole group of NCL and JNCL in particular. As less common forms of NCL began to be discovered, these were often referred to by the country of origin of the first described patients (*table 1*).

Today, at least 14 affected genes are implicated, from *CLN1* to *CLN14*, 13 of which have been identified (*CLN1-8* and *CLN10-14*). *CLN9* refers to the predicted locus in a family who do not appear to have mutations in any of the known genetic forms (Schulz et al., 2004). The currently genetically identified types of NCL are listed in *table 2*.

Very recently, a new nomenclature has been discussed internationally and subsequently proposed which

is gene-based and specific to phenotypic variation arising from different mutations. This is an axial diagnostic classification system that includes seven axes:

- affected gene (CLN gene symbol);
- mutation diagnosis;
- biochemical phenotype;
- clinical phenotype;
- ultrastructural features;
- level of functional impairment; and
- other remarks (additional genetic, environmental, or clinical features) (Mole & Williams, 2013).

In reality, NCL classed according to the affected gene, combined with the age at onset, is sufficient for general use (e.g. classic infantile CLN1 disease, or adult CLN1 disease).

Pathophysiology of NCLs

NCL are grouped together on pathological grounds due to the common presence of neuronal and extraneural autofluorescent pigment accumulations, despite diverse underlying biochemical aetiologies. They are considered lysosomal storage diseases, however, NCLs also exhibit characteristics that distinguish them from lysosomal storage diseases (Mink, 2010). Consistent with lysosomal storage disorders (LSD), many of the identified NCL proteins are present in the lysosomes, and lipofuscin-like ceroid lipopigments also accumulate in the lysosomes. Under the electron microscope, the accumulated material takes different forms: granular osmiophilic deposits (GRODs), curvilinear profiles (CLP), fingerprint profiles (FPP), as well as rectilinear complex (RLC) or so called ‘condensed forms’. However, unlike classic lysosomal storage disorders, for the NCL, the ‘stored’ material is not disease-specific. While the major clinical NCL subtypes (CLN1, CLN2 and CLN3

Table 1. Historical NCL classification.

| Disease | Clinical phenotype | Abbreviated name | Eponym |
|---------|---------------------------------|------------------|---|
| CLN1 | Infantile classic | INCL | Haltia-Santavuori |
| CLN2 | Late-infantile classic | LINCL | Janský-Bielschowsky |
| CLN3 | Juvenile | JNCL | Batten-Spielmeyer-Sjögren |
| CLN4 | Adult autosomal dominant | ANCL | Parry |
| CLN5 | Late-infantile variant | vLINCL | Finnish variant late infantile |
| CLN6 | Early-juvenile / late-infantile | vLINCL | Lake-Cavanagh/Indian variant/Kufs (adult) |
| CLN7 | Late-infantile variant | vLINCL | Turkish variant late infantile |
| CLN8 | EPMR late-infantile variant | vLINCL | Northern epilepsy/EPMR |



Figure 1. (A) Otto Christian Stengel; (B) Frederick Eustace Batten (first from the left, bottom row); (C) Walther Spielmeyer; (D) Jan Janský; (E) Max Bielschowsky; (F) Hugo Kufs; (G) Matti Haltia; and (H) Pirkko Santavuori.

Table 2. Genetic classification of NCL.

| Locus name | Gene symbol | Locus | Protein name | Phenotypic spectrum |
|------------|----------------|----------|----------------------------------|---|
| CLN1 | <i>PPT1</i> | 1p34.2 | Palmitoyl protein thioesterase 1 | I, LI, J, A |
| CLN2 | <i>TPP1</i> | 11p15.4 | Tripeptidyl peptidase 1 | LI, J, P |
| CLN3 | <i>CLN3</i> | 16p11.2 | CLN3 | J, P |
| CLN4 | <i>DNAJC5</i> | 20q13.33 | DnaJ homolog | A (Parry disease) subfamily C member 5 |
| CLN5 | <i>CLN5</i> | 13q22.3 | CLN5 | LI, J, P, A |
| CLN6 | <i>CLN6</i> | 15q23 | CLN6 | LI, P, A (Kufs type A) |
| CLN7 | <i>MFSD8</i> | 4q28.2 | Major facilitator superfamily | LI, J domain-containing protein 8 |
| CLN8 | <i>CLN8</i> | 8p23.3 | CLN8 | LI, P |
| CLN9 | <i>n/a</i> | unknown | unknown | |
| CLN10 | <i>CTSD</i> | 11p15.5 | Cathepsin D | C, LI, J, A |
| CLN11 | <i>GRN</i> | 17q21.31 | Granulins | A |
| CLN12 | <i>ATP13A2</i> | 1p36.13 | Probable cation-transporting | J ATPase 13A2 |
| CLN13 | <i>CTSF</i> | 11q13.2 | Cathepsin F | A (Kufs type B) |
| CLN14 | <i>KCTD7</i> | 7q11.21 | BTB/POZ domain-containing | I protein KCTD7 |

A: adult; C: congenital; I: infantile; J: juvenile; LI: late-infantile; P: protracted.

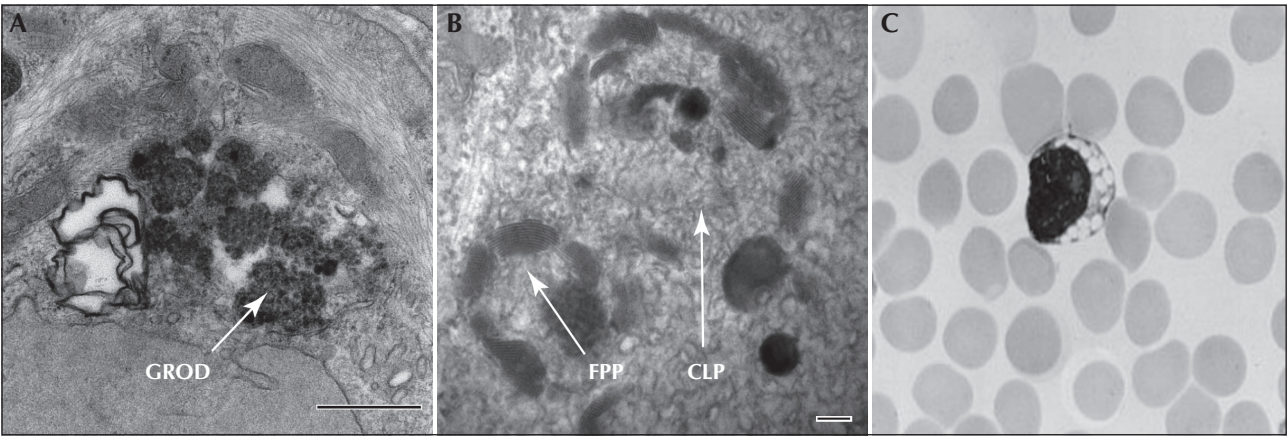


Figure 2. Electron microscopy and pathology findings in NCL. (A) Granular osmophilic deposits (GROD) in a conjunctival biopsy from a patient with *CLN1* mutations (bar is 500 nm); (B) Fingerprint profiles (FPP) and curvilinear profiles (CLP) in a conjunctival biopsy from a patient with *CLN2* mutations (bar is 100 nm). The predominance of curvilinear profiles and the comparative scarcity of fingerprint bodies were consistent with *CLN2*. Should fingerprint profiles predominate, the biopsy would be more consistent with *CLN3*; (C) Vacuolated lymphocytes in a patient with *CLN3* disease.

diseases) were formerly each associated with characteristic inclusions under electron microscopy (GRODs, CLP, and FPP, respectively) (figure 2), the ultrastructural findings do not absolutely correlate with clinical presentation, and the same NCL may contain more

than one pattern of inclusion (table 3). Furthermore, the appearance of the pathological inclusions can depend on the tissue examined. Vacuolated lymphocytes are typically seen in classic juvenile *CLN3* disease.

Table 3. Typical electron microscopy findings and enzyme activity according to NCL genotype.

| Disease | EM | Lymphocytes | Enzyme activity |
|---------|----------------|----------------|-----------------|
| CLN1 | GROD | Non-vacuolated | PPT1 deficiency |
| CLN2 | CLP | Non-vacuolated | TPP1 deficiency |
| CLN3 | FPP | Vacuolated | Not applicable |
| CLN4 | GROD, mixed | Non-vacuolated | Not applicable |
| CLN5 | FPP, CLP, GROD | Non-vacuolated | Not applicable |
| CLN6 | CLP, FPP, RLC | Non-vacuolated | Not applicable |
| CLN7 | CLP, FPP, RLC | Non-vacuolated | Not applicable |
| CLN8 | CLP, FPP, GROD | Non-vacuolated | Not applicable |
| CLN9 | GROD, CLP | Non-vacuolated | Not applicable |
| CLN10 | GROD | Non-vacuolated | CTSD deficiency |
| CLN11 | FPP | Non-vacuolated | Not applicable |
| CLN12 | GROD, mixed | Non-vacuolated | Not applicable |
| CLN13 | FPP or none | Non-vacuolated | CTSF deficiency |
| CLN14 | GROD, FPP | Non-vacuolated | Not applicable |

GROD: granular osmophilic deposits; CLP: curvilinear profiles; FPP: fingerprint profiles; RLC: rectilinear complex.

CLN1

CLN1: Genetics

In CLN1 disease, the underlying defect is the lack of activity of the lysosomal palmitoyl protein thioesterase (PPT1), an enzyme that removes palmitate residues from proteins (Vesa *et al.*, 1995). The main protein component of the storage material is saposins A and D, with a characteristic ultrastructure of GRODs. To date, 64 mutations have been described in *CLN1* (for updated information on NCL mutations see the online database at <http://www.ucl.ac.uk/ncl/>). The exact physiological function and *in vivo* substrates of PPT1 are unknown, but it is proposed that PPT1 is required to maintain various cellular processes, including apoptosis, endocytosis, vesicular trafficking, synaptic function, and intracellular signalling (Greaves & Chamberlain, 2007). In neurons, PPT1 is found also outside the lysosomal compartment in presynaptic terminals (Ahtiainen *et al.*, 2006), suggesting that PPT1 is not exclusively confined to lysosomes, and the disease is not due solely to the abnormal storage.

CLN1: Clinical presentation

The first symptoms of classic infantile CLN1 disease manifest in the second half of the first year of life,

with irritability, followed by rapid psychomotor deterioration, central hypotonia, and deceleration of head growth. These are quickly followed by myoclonic jerks (and other seizures types) and blindness with optic atrophy. The ERG (electroretinogram) is unrecordable by age 4 years. Hand-wringing often develops during the disease course, which, along with the slowing of head growth and developmental regression, raises the differential for Rett syndrome, but unlike the latter, CLN1 disease does not stabilize, continuing instead to deteriorate until death in early childhood (Mole *et al.*, 2005). Most children die at around 10 years of age. Of all the NCLs, CLN1 disease has the widest range of age at onset, determined by the combination of particular mutations. Although the majority of patients have infantile onset, some have late-infantile, juvenile, and even adult-onset, as late as 40 years of age (Van Diggelen *et al.*, 2001, Ramadan *et al.*, 2007).

CLN1: Diagnosis

Historically, the diagnosis was made based on the ultrastructural finding of GRODs, together with the presence of suggestive clinical features. The diagnosis can now be made rapidly and specifically by demonstrating a lack of PPT1 activity, even in adult-onset forms. Brain MRI usually demonstrates a variable degree of cerebral atrophy: signal change in the thalami and

basal ganglia and thin, hyperintense, periventricular high-signal rims of white matter (Riikonen *et al.*, 2000). A progressive diffuse brain atrophy on MRI is seen in children during the first 4 years of life which then usually stabilizes. MR spectroscopy shows a decrease in the N-acetyl aspartate (NAA) peak and increased choline, however, with the rapid progression of the disease, all peaks completely disappear by the age of 6 years (Vanhanen *et al.*, 2004).

Neurophysiological findings in CLN1 disease are non-specific and include decreased reactivity of the posterior dominant rhythm to eye opening and eye closure, loss of sleep spindles by the age of 2 and an evolution towards an isoelectric electroencephalography (EEG) after the age of 3, which parallels the neuronal degeneration and brain atrophy.

CLN1: Differential diagnosis

Differential diagnosis should include other progressive neurodegenerative disorders with onset from birth to age 2 years: Rett syndrome, hexosaminidase A deficiency, leukodystrophies, peroxisomal disorders, Niemann-Pick disease types A and B, and Leigh syndrome. While some of these disorders are associated with cortical blindness, retinal involvement is rarely seen (Mole & Williams, 2013).

CLN1: Treatment

Treatment of CLN1 disease is symptomatic. As the enzyme cleaves fatty acid thioesters in plasma membranes, it was suggested that the drug cysteamine, a simple aminothioliol used for the treatment of cystinosis, may have some effect. A clinical trial in children did not show any significant improvements (Levin *et al.*, 2014) and *in vitro* studies also cast doubt on this concept (Lu & Hofmann, 2006). A phase I trial of intracerebral injection of human foetal neuronal stem cells has been performed (Guillaume *et al.*, 2008; Mole, 2014) and enzyme replacement therapy has been considered (Lu *et al.*, 2010).

CLN2

CLN2: Genetics

CLN2 encodes the lysosomal enzyme tripeptidyl peptidase (TPP1), a member of the serine carboxyl proteinase family (Rawlings & Barrett, 1999). This group of enzymes removes tripeptides from the N termini of small polypeptides such as the subunit c of mitochondrial ATP synthase. To date, more than 109 mutations have been described, the two most

common being the splice site mutation c.509-1G>C and the nonsense mutation p.Arg208* resulting in broadly similar clinical phenotypes (Mole *et al.*, 2005). The majority of the protein component of the storage bodies is subunit c of mitochondrial ATP synthase, as well as low amounts of saposins A and D (Lake & Hall, 1993; Jalanko & Braulke, 2009).

CLN2: Clinical presentation

Classic late-infantile CLN2 disease presents around the third year of life, with intractable epilepsy and an arrest of cognitive development. Myoclonus and ataxia are commonly seen early in the course, followed by progressive cognitive and motor decline. Retinopathy is often not prominent early in the course and may be overlooked after progression to more severe neurological deficits. Spasticity, truncal hypotonia, loss of head control, near-continuous myoclonus, frequent seizures, and an extended vegetative state are characteristic, until death in early adolescence. Death is often due to aspiration pneumonia. A few cases presenting late (8 years) have been reported, exhibiting slow regression with death as late as 40 years of age (Sleat *et al.*, 1999).

CLN2: Diagnosis

Historically, diagnosis was made based on the presence of clinical features and the ultrastructural demonstration of curvilinear bodies. However, the diagnosis can now be reached rapidly and specifically by demonstrating a lack of TPP1 activity using blood, skin biopsy, saliva, or dried blot spot.

Brain MRI in CLN2 disease shows progressive cerebral atrophy that predominates in the infratentorial region. Hypointense thalami on T2-weighted images were also reported (Seitz *et al.*, 1998). MR spectroscopy demonstrates a reduction in the NAA peak and an increase in myo-inositol and glutamate/glutamine in the white matter (Seitz *et al.*, 1998).

EEG includes characteristic occipital spike responses to slow flash (1-2 Hz) stimulation which precede the onset of seizures and which increase as the disease progresses (*figure 3*). Electroretinogram is diminished even before noticeable visual loss (Wisniewski *et al.*, 1998; Wisniewski *et al.*, 2001; Goebel & Wisniewski, 2004). Visual evoked potentials are also enhanced at the onset of the disease.

CLN2: Differential diagnosis

Other progressive neurological diseases with onset from ages 2 to 4 years should be considered, including epileptic encephalopathies, lysosomal storage

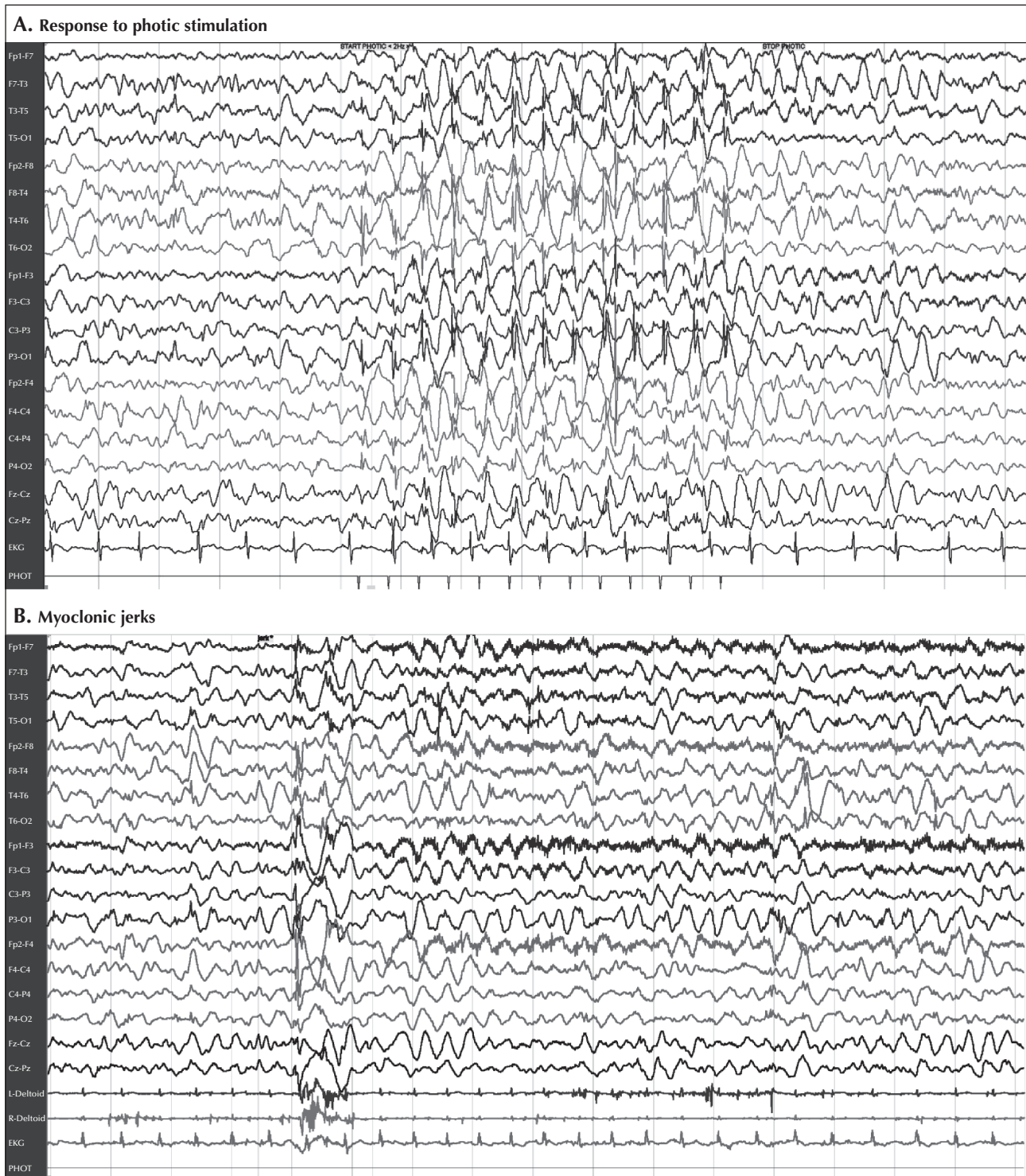


Figure 3. Classic EEG findings in CLN2 disease.

disorders, mitochondrial diseases, and leukodystrophies. Other rarer NCL variants, such as CLN5, CLN6, CLN7 and CLN8 diseases, should also be considered if TPP1 activity is normal.

CLN2: Treatment

Treatment is symptomatic. An experimental treatment approach uses intracerebral injection of viral vectors

containing normal coding segments of the *CLN2* gene. In a mouse model of *CLN2* disease, this procedure resulted in cerebral enzyme expression, reduced brain pathology, and increased survival. A small number of human patients have recently been treated in the same way (Worgall *et al.*, 2008), but the trial was too small to determine efficacy. A phase I trial of intracerebral injection of human foetal neuronal stem cells has been performed (Mink, 2010). Enzyme replacement therapy was efficacious in mice and dogs (Passini *et al.*, 2006; Chang *et al.*, 2008; Whiting *et al.*, 2014) and a phase 1 trial is well underway in Europe (Biomarin).

CLN3

CLN3: Genetics

CLN3 encodes a membrane protein of unknown function. Whilst generally considered to be present predominantly in the endolysosome system, this protein has been reported to localize to membrane lipid rafts in synaptosomes, Golgi, and the cell membrane, as well as in mitochondria (Phillips *et al.*, 2005). Currently more than 57 mutations have been characterized in the *CLN3* gene. Most cases world-wide, which can be traced to a northern European origin, are due to a common ancestral 1-kb deletion founding mutation (Munroe *et al.*, 1997) shown to retain partial activity (Kitzmüller *et al.*, 2008).

Numerous roles have been attributed to the gene product of *CLN3*, and much work is needed to reconcile these disparate functions. In mitochondria, the gene product of *CLN3* was suggested to aid the processing of mitochondrial membrane proteins, such as ATPase subunit c, which accumulates in this condition as a result of synaptic vesicle transport (Margraf *et al.*, 1999). The *CLN3* gene product has been implicated in the regulation of lysosomal pH, transport of basic amino acids into the lysosome, and lysosomal size (Golabek *et al.*, 2000; Holopainen *et al.*, 2001; Ramirez-Montealegre & Pearce, 2005). An antiapoptotic role has been ascribed to the gene product of *CLN3*; the C-terminus appears to participate in cell cycle regulation, and mutations of this region result in slow growth and increased apoptosis (Puranam *et al.*, 1999). *CLN3* knockout mice show neutralizing antibodies against glutamic acid decarboxylase (GAD65), suggesting that an autoimmune response against GAD65 might contribute to preferential loss of GABAergic neurons in this disease. However, it is not understood whether these autoantibodies contribute to the pathogenesis or whether they are secondary entities arising during neurodegeneration, although their presence may influence excitotoxic mechanisms (Chattopadhyay *et al.*, 2002). Recently, an enzymatic

function has been associated with the *CLN3* gene product, namely palmitoyl-protein D-9 desaturase activity (Narayan *et al.*, 2006). Another group showed a correlation between *CLN3* expression and the synthesis of bis monoacylglycerol phosphate (BMP) and suggested that the *CLN3* gene product may play a role in the biosynthesis of BMP (Hobert & Dawson, 2007).

CLN3: Clinical presentation

Classic juvenile *CLN3* disease presents between the ages of 4 to 8 years (mean of 5 years) with a progressive loss of vision due to retinal degeneration, followed by progressive dementia. Ocular pathology is initially a pigmentary retinopathy often misdiagnosed as retinitis pigmentosa or cone dystrophy. During adolescence, epilepsy and extrapyramidal/parkinsonian signs (rigidity, hypokinesia, shuffling gait, impaired balance) are more prominent. Neuropsychiatric symptoms, such as anxiety and aggression, are common (Marshall *et al.*, 2005). The clinical course is variable but inexorably progressive toward death in the second or third decade. *CLN3* disease may present in adulthood with visual failure sometimes later accompanied with heart failure (Eksandh *et al.*, 2000).

CLN3: Diagnosis

Ultrastructurally, *CLN3* disease cases exhibit fingerprint profiles. These may be the only apparent feature within the lysosomal residual body, or may occur in conjunction with curvilinear or rectilinear profiles, or as a small component within large membrane-bound lysosomal vacuoles. The diagnostic hallmark of this frequent NCL type is conspicuous vacuoles in the cytoplasm of lymphocytes which are detectable on a regular blood smear (Anderson *et al.*, 2005). Diagnosis is based on clinical suspicion, the presence of vacuolated lymphocytes, and ultrastructural studies or combined with genetic testing.

Brain MRI shows cerebral and cerebellar atrophy in the later stages (age > 15 years) and is normal before the age of 10 years. MR spectroscopy has not shown specific abnormalities.

The EEG shows non-specific progressive background disorganization and spike-and-slow-wave complexes. The predominant seizure type is generalized tonic-clonic, however, partial complex seizures can occur as well. Enhanced somatosensory evoked potentials may be seen, supporting the presence of a myoclonic component even before myoclonus is clinically present.

CLN3: Differential diagnosis

Differential diagnosis is limited given that juvenile *CLN3* disease has a unique presentation.

Peroxisomal, mitochondrial and other lysosomal disorders that are associated with retinopathy can be considered.

CLN3: Treatment

Treatment is symptomatic. Autoimmunity against GAD65 has been used as the basis for investigation of immunomodulatory treatments (Pearce *et al.*, 2004).

CLN4

CLN4: Genetics

The *CLN4* gene encodes DNAJC5, which underlies the autosomal dominant adult form of NCL, known as “Parry disease”. The gene symbol *CLN4* was also used in the past to account for a heterogeneous group of adult forms of NCL which were recessively inherited (collectively recognised as Kufs disease) without known genetic loci at that time. Some of these forms were later identified as being secondary to mutations in *CLN6* and *CLN13*.

Other NCL genes that may present with adult-onset are *CLN1*, *CLN5*, *CLN10*, *CLN10* and *CLN13* (table 2). While for most NCLs there is a specific phenotype associated with the loss of function of a particular *CLN* gene, for some NCLs that arise from mutations that have an incomplete effect of the gene function, the associated phenotypes are protracted or have a later age of onset (reviewed in Mole & Cotman [2015]).

CLN4: Clinical presentation

Adult-onset NCL can present with two different clinical phenotypes: Kufs type A with marked myoclonus, progressive epilepsy, dementia and ataxia; and Kufs type B, marked by behavioural changes and dementia, as well as peculiar facial dyskinesia. Vision is not impaired in primary adult forms of NCL.

CLN4: Diagnosis

Ultrastructural patterns include granular, curvilinear, or fingerprint profiles in different cell types and organs of the same patient, or a combination of patterns. Vacuolated lymphocytes were not reported.

CLN4: Differential diagnosis

All NCL with possible onset in adulthood should be included in the differential diagnosis. The pattern of inheritance may point towards an autosomal recessive or autosomal dominant form.

CLN4: Treatment

Treatment is symptomatic.

CLN5

CLN5: Genetics

CLN5 encodes a soluble protein that is directed to the lysosome. It is reported to interact with the gene products of *CLN2* and *CLN3* (Vesa *et al.*, 1995). These observations suggest that there may be common molecular pathways or important interactions between pathways in various types of NCLs. Currently, more than 36 different mutations have been described in *CLN5*. The most common mutation, occurring in patients of Finnish origin, is a 2-base pair deletion in exon 4 (c.1175_1176delAT) that results in an early stop codon (p.Tyr392*).

CLN5: Clinical presentation

The first symptoms of the disease typically begin between 4 and 7 years of age (slightly older than the range in *CLN2* disease), although cases are reported that present in adulthood. The usual course is motor clumsiness followed by progressive visual failure and blindness, dementia, motor decline, myoclonus, and seizures. The rate of progression is variable, but ultimately death occurs between the ages of 14 and 36 years (Mink, 2010). First reported in Finland, this type of NCL has recently been observed in many other European countries (UK, Czech Republic, Netherlands, Portugal, Italy), in North America (Canada, USA), South America (Argentina, Colombia), and other countries including Afghanistan and Pakistan. It should be considered in any exhaustive diagnostic approach of a patient with suspected NCL, especially with onset in late infancy but also up to adulthood (Santavuori *et al.*, 1991).

CLN5: Diagnosis

Ultrastructurally, lipopigments are distributed in the central nervous system and extracerebrally, and include fingerprint bodies, curvilinear profiles, lamellar inclusions, and occasionally condensed fingerprint images associated with lipid droplets. The major stored material is subunit c of the mitochondrial ATP synthase (Tyynela *et al.*, 1997). Similar to *CLN3*-defective fibroblasts, *CLN5*-deficient fibroblasts also exhibit elevated intralysosomal pH (Kyttala *et al.*, 2006).

Brain imaging shows prominent cerebellar atrophy and in addition, on T2-weighted images, the thalamic signal intensity is low compared to that of the caudate, while increased signal intensity is seen in the periventricular white matter and the posterior limb of the

internal capsule (Autti *et al.*, 1992). Neurophysiological examination shows giant visual evoked potentials, exaggerated somatosensory potentials, and occipital spikes in response to photic stimulation, similar to CLN2. Ultimately, CLN5 can only be confirmed by DNA analysis.

CLN5: Differential diagnosis

The clinical presentation with dementia, motor clumsiness and visual failure is strongly suggestive of a neurodegenerative disease. The probability of NCL is further supported by the electrophysiological and imaging studies.

CLN5: Treatment

Treatment is symptomatic. The experimental finding that at least a portion of the *CLN5* gene product is trafficked via the manose-6-phosphate pathway (Sleat *et al.*, 2005) means that therapeutic approaches that depend upon cross-correction, including enzyme replacement therapy, gene therapy, and stem cell transplantation are likely to be tested in the near future (Selden *et al.*, 2013).

CLN6

CLN6: Genetics

CLN6 encodes a protein of unknown function with seven transmembrane domains localizing to the endoplasmic reticulum (ER) (Sharp *et al.*, 2003; Heine *et al.*, 2004). Currently, more than 68 different mutations have been described in CLN6 disease. Patients are found all over the world, with particular concentrations in Costa Rica and Portugal, arising from a founder effect, as well as a range of mutations in Turkey and Newfoundland.

CLN6: Clinical presentation

Age at onset of CLN6 disease straddles the ages at onset of CLN1, CLN2, and CLN3 diseases, ranging from 18 months to 8 years, with the majority between 3 and 5 years. Early visual failure occurs in about 50 per cent of patients. The most prominent symptoms are motor impairment, including developmental delay, dysarthria, and ataxia. Seizures occur in the majority of patients, and usually begin before age 5 years. Deterioration is rapid after diagnosis and most children die between the ages of 5 and 12 years.

CLN6: Diagnosis

Diagnosis is based on clinical suspicion and genetic testing. Ultrastructurally, a mix of rectilinear profiles

and fingerprint profiles are seen. The stored material contains subunit c of mitochondrial ATP synthase (Elleder *et al.*, 1997). There is marked neuronal loss in layer V of the cerebral cortex and the extent of cerebral atrophy in CLN6 patients has been shown to be proportionate to the duration of symptoms based on post-mortem data (Elleder *et al.*, 1997).

MR imaging shows progressive cerebral and cerebellar atrophy. As in CLN2 disease, EEG shows progressive background slowing and high-amplitude discharges in the posterior head regions in response to the photic stimulation.

CLN6: Differential diagnosis

Other NCL variants should be considered in the differential diagnosis of CLN6 disease. CLN1, CLN2 and CLN10 diseases can be excluded easily by enzyme analysis of PPT1, TPP1 and CTSD. Lymphocyte vacuolation, a hallmark of CLN3, is not seen in CLN6 disease.

CLN6: Treatment

Treatment is symptomatic. As the function of the CLN6 protein remains unknown, no experimental therapeutic studies have yet been initiated.

CLN7

CLN7: Genetics

The *CLN7* gene product belongs to the large major facilitator superfamily (MFS) that transports specific classes of substrates, including sugars, drugs, inorganic and organic cations, and various metabolites (Jalanko & Braulke, 2009). CLN7 is also referred to as MFSD8. CLN7 is localized to lysosomes (Kousi *et al.*, 2009). At present, more than 31 disease-causing mutations have been reported.

CLN7: Clinical presentation

Age at onset is usually between 2 and 7 years. Psychomotor regression or seizures are the initial presenting signs. Progressive cognitive and motor deterioration, myoclonus, personality changes and blindness occur later. The disease has a rapidly progressing course. A Rett syndrome-like onset has also been reported (Craiu *et al.*, 2015).

CLN7: Diagnosis

Ultrastructural examination reveals both fingerprint patterns and rectilinear patterns. This form can be diagnosed by ultrastructural pathological analysis of

peripheral lymphocytes where dense fingerprint profiles are observed. Vacuolations are not usually present in lymphocytes. Diagnosis is based on clinical suspicion and genetic testing.

Brain MR studies are abnormal from the early stages of the disease and show progressive cerebral and cerebellar atrophy, thinning of the corpus callosum, and hypointensity of the thalami on T2-weighted images. Neurophysiological studies show diffuse background slowing of the EEG with occipital spikes, more prominent during sleep, which may evolve into electrical status epilepticus during slow-wave sleep.

CLN7: Differential diagnosis

Differential diagnosis includes mainly CLN3 and CLN6 diseases. Condensed fingerprint profiles in the lymphocytes and the absence of vacuolation is characteristic for CLN7 disease.

CLN7: Treatment

Clinical management is supportive. No experimental therapeutic trials have been initiated so far.

CLN8

CLN8: Genetics

CLN8 encodes a polytopic membrane protein that is localized to the ER and shuttles between the ER and ER-Golgi intermediate complex. The exact function is unknown, but it belongs to the TRAM-Lag1p-CLN8 (TLC) family of proteins, which are suggested to have roles in biosynthesis, metabolism, transport, and detection of lipids (Jalanko & Braulke, 2009). More than 24 mutations have been described.

CLN8: Clinical presentation

Depending on the mutation, CLN8 disease presents with childhood-onset (5-10 years), intractable epilepsy, followed by progressive cognitive decline or mild developmental delay in late infancy, followed by a florid PME with progressive myoclonus (Minassian *et al.*, 2016), seizures, retinopathy, and psychomotor regression starting between 3 and 6 years. This typical late-infantile NCL phenotype leads to loss of vision (Topcu *et al.*, 2004).

In the very specific subtype of Progressive Epilepsy with Mental Retardation or Northern Epilepsy, caused by a defined missense mutation p.Arg24Gly, age at onset is 5 to 10 years and seizures are the first symptom. All patients have generalized tonic-clonic seizures with frequent episodes of status

epilepticus. As patients pass through puberty, the frequency of seizures decreases but progressive dementia and motor impairment continues (Ranta & Lehesjoki, 2000). Patients with Northern Epilepsy may survive until 50-60 years of age.

CLN8: Diagnosis

Diagnosis is based on clinical suspicion and genetic testing. GROD, curvilinear, and fingerprint profiles have been reported on electron microscopy, in various tissues, including lymphocytes, however, the stored material consists mostly of subunit c of mitochondrial ATP synthase.

MR studies show progressive cerebral and cerebellar atrophy with thinning of the corpus callosum. Neurophysiological testing is similar to other NCLs. Diagnosis can only be confirmed by DNA analysis.

CLN8: Treatment

Treatment is supportive, and no experimental therapeutic trials have been attempted so far.

CLN9

CLN9: Genetics

CLN9 has been proposed as a specific NCL entity, but no gene has yet been identified. Fibroblasts from affected families have a distinctive phenotype (rapid growth, sensitivity to apoptosis, manifestation of a cell adhesion defect, and reduced levels of ceramide, dihydroceramide, and sphingomyelin) (Schulz *et al.*, 2004). Little is known of the function of the unidentified CLN9 protein.

CLN9: Clinical presentation

CLN9 is clinically indistinguishable from juvenile CLN3 disease, but perhaps with a more rapid course.

CLN9: Diagnosis

Ultrastructure is characterized by GRODs and curvilinear bodies. Diagnosis is one of exclusion. For patients presenting with typical features of CLN3 disease who have characteristic ultrastructural abnormalities, but no mutation in *CLN3*, possible CLN9 disease should be considered.

CLN10

CLN10: Genetics

The affected gene encodes cathepsin D (CTSD), a lysosomal enzyme thought to be important for neuronal stability (Siintola *et al.*, 2006; Steinfeld *et al.*, 2006), which is also secreted and exerts effects in the extracellular environment. More than 7 mutations have been described. Alterations in a macroautophagy-lysosomal degradation pathway appear to mediate neurodegeneration in this disease.

CLN10: Clinical presentation

CLN10 disease is characterized (in the congenital form) by primary microcephaly, neonatal (possibly already intrauterine) epilepsy, respiratory insufficiency, and rigidity. Death occurs within hours to weeks after birth. Late-onset forms of this NCL may be seen in juveniles and adults (Steinfeld *et al.*, 2006). In one patient, missense mutations caused a childhood onset neurodegenerative disease with ataxia, retinopathy, severe cognitive decline, and apparently no seizures at age 17. The pathological correlate was GRODs (Steinfeld *et al.*, 2006).

CLN10: Diagnosis

Diagnosis is based on clinical presentation and enzymatic testing for CTSD in fibroblasts or blood. Genetic testing for mutations in *CLN10* is also available. On post-mortem examination, massive loss of neurons in the cerebral cortex, extensive gliosis, absence of myelin, and autofluorescent storage bodies with a GROD ultrastructure have been described.

CLN10: Treatment

Treatment is symptomatic and mainly targets quality of life.

CLN11

CLN11 disease is characterized by rapidly progressive visual loss due to retinal dystrophy, seizures, cerebellar ataxia, and cerebellar atrophy. Two disease-causing mutations, present as compound heterozygous in the *GRN* gene, have been reported in two Italian siblings from nearby villages in Lombardy, Italy. The transmission pattern of adult-onset NCL in the family was consistent with an autosomal recessive inheritance. Electron microscopic examination of a skin biopsy demonstrated numerous fingerprint profiles

in membrane-bound structures in eccrine secretory cells and in endothelium, consistent with NCL. EEG results showed polyspike-wave discharges with a posterior emphasis, and MRI indicated cerebellar atrophy. Heterozygous mutations in this gene were previously known to cause frontotemporal lobe dementia (Smith *et al.*, 2012).

CLN12

CLN12 disease was reported recently in a Belgian family. The index case had unsteady gait, myoclonus, and mood disturbance from age 11 to 13, progressing to clear extrapyramidal involvement with akinesia and rigidity, as well as dysarthric speech. There was no retinal involvement. Exome sequencing identified a single homozygous mutation in *ATP13A2* that fully segregated with the disease within the family. Muscle biopsy showed numerous subsarcolemmal autofluorescence bodies with a fingerprint appearance under electron microscopy, suggestive of neurogenic muscular atrophy. Mutations in *ATP13A2* are better known to cause Kufor-Rakeb syndrome (KRS), a rare parkinsonian phenotype with juvenile onset (Bras *et al.*, 2012).

CLN13

Five disease-causing mutations have been reported in *CLN3* (Smith *et al.*, 2013). CLN13 disease is an adult-onset neuronal ceroid lipofuscinosis with cathepsin F (CTSF) deficiency, without vacuolated lymphocytes. The clinical phenotype (Kufs type B) is characterized by behaviour abnormalities and dementia, which may be associated with motor dysfunction, ataxia, extrapyramidal signs, and bulbar signs. Electron microscopy has showed fingerprint profiles in some cases. The protein product, CTSF, is part of the papain family of cysteine proteinases that represent a major component of the lysosomal proteolytic system.

CLN14

One disease-causing mutation has been reported in a Mexican family with vision loss, cognitive and motor regression, premature death, and prominent NCL-type storage material (Staropoli *et al.*, 2012). The gene product, BTB/POZ domain-containing protein KCTD7, is a potassium channel tetramerization domain-containing protein 7. Other mutations in this protein cause infantile PME or opsoclonus-myoclonus ataxia-like syndromes.

Diagnostic approach of NCLs

NCL have a recognizable phenotype that correlates with the progressive grey matter neurodegenerative process involving the cortex, deep grey nuclei, cerebellum, and retina. The first approach to diagnosis should consider age at onset and type of clinical presentation, and has been well summarised recently (Schulz *et al.*, 2013).

Presentation in a neonate with severe epilepsy and microcephaly should suggest CLN10 disease as a possible diagnosis. Enzyme testing for CTSD (CLN10) should be the first step. If this is negative, further or concurrent enzyme testing for PPT1 and TTP1 should be attempted before more invasive biopsies.

In young children (> 6 months) with otherwise unexplained epilepsy and developmental arrest, CLN1 and CLN2 diseases are the most likely considerations. If enzyme testing for PPT1 and TTP1 are negative and electron microscopy demonstrates typical storage material, genetic testing for *CLN5*, *CLN6*, *CLN7/MFSD8*, *CLN8* and *CLN14/KCTD7* should be considered.

A school-aged child presenting with rapid visual loss between ages 4 and 7 should be tested for CLN3 disease, first looking for lymphocyte vacuoles. If no lymphocyte vacuolization is present and testing for PPT1, TPP1 and CTSD is also negative, skin biopsy is indicated to assess if typical NCL storage material is present. If so, genetic testing for *CLN5*, *CLN6*, *CLN7/MFSD8*, *CLN8*, and *CLN12/ATP13A2* is indicated.

The NCL variants CLN5, CLN6, CLN7, or CLN8 should be considered in a child who phenotypically and ultrastructurally has an NCL, but testing for the more common entities is negative.

In adults with non-specific mental, motor, or behavioural abnormalities in which NCL is suspected, the first line of investigation includes the enzymatic assays for PPT1, TTP1, CTSD and CTSF which, if normal, should prompt ultrastructural examination. If storage material is present, genetic testing for autosomal recessive (*CLN6*, *CLN11/GRN*, *CLN13/CTSF*) and autosomal dominant NCL (*CLN4/DNAJC5*) should be initiated, and if negative, all other remaining NCL genes should be investigated.

Generally speaking, enzyme testing should be performed first. Ultrastructural examination of skin or lymphocytes should be performed prior to gene sequencing, however, gene “chips” or other new DNA approaches that test concurrently for multiple NCL genes in the near future will reverse this approach. If there is no storage material, NCL is highly unlikely, although possible.

Symptomatic treatment for children with NCL

Patients with NCL require symptomatic treatment for a constellation of neurological manifestations, including seizures, sleep problems, extrapyramidal symptoms, behavioural problems, anxiety, and psychosis.

Routine medical management of children and young adults with complex neurological disability is relevant to all those affected by NCL, including clinical surveillance for sialorrhea, swallowing difficulties, gastroesophageal reflux, aspiration pneumonia, and X-ray surveillance of hips and spine.

Seizures may not require early treatment if generalized convulsions are rare and epileptic myoclonus is not obvious. As seizures progress, antiepileptic drugs of choice are valproic acid and lamotrigine, alone or in combination. However, lamotrigine may exacerbate seizures in CLN2 disease. Topiramate and levetiracetam are also effective. Benzodiazepines are useful in combination therapy, but they may cause problems due to sialorrhea. Carbamazepine, phenytoin, and gabapentin should be avoided as they may worsen myoclonic seizures. The ultimate goal must always be to improve the quality of life, and for this disease, the aim is not focussed on becoming seizure-free. Sleep disturbance is common and is likely to worsen with age. In general, a calm environment and set routines before going to bed are helpful. Benzodiazepines and sedatives are commonly used. Melatonin was found to have limited effect. Benzodiazepines may also benefit anxiety and spasticity. Trihexyphenidyl improves dystonia and sialorrhea.

Emotional, behavioural and psychotic problems are common. Delusions and hallucinations may be managed using newer atypical neuroleptics, such as risperidone or olanzapine. If depression is thought to be the underlying problem, selective serotonin reuptake inhibitors may be considered which have been found to be beneficial. In all cases, medication should be kept to a minimum in order to avoid side effects or worsening disease progression.

Genetic counselling and testing of NCLs

The NCLs are inherited in an autosomal recessive manner with the exception of adult NCL, which can be inherited in either an autosomal recessive or an autosomal dominant manner. The parents of an affected child are obligate heterozygotes and are asymptomatic. The siblings of a proband have a 25 per cent chance of being affected, a 50 per cent chance of being an asymptomatic carrier, and a 25 per cent chance of being unaffected and not a carrier.

Prenatal testing is possible in high-risk pregnancies (if biochemical studies in the proband have revealed deficient activity of the enzymes CTSD, PPT1, or TPP1, or mutations defined in any NCL gene). In these instances, testing is performed on foetal cells obtained by chorionic villus sampling at 10-12 weeks of gestation or amniocentesis usually performed between 15-18 weeks of gestation (Mole & Williams, 2013).

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Sialidoses

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ABSTRACT – Sialidoses are autosomal recessive disorders caused by *NEU1* gene mutations and are classified on the basis of their phenotype and onset age. Sialidosis type II, with infantile onset, has a more severe phenotype characterized by coarse facial features, hepatomegaly, dysostosis multiplex, and developmental delay while patients with the late and milder type, known as “cherry red spot-myoclonus syndrome” develop myoclonic epilepsy, visual impairment and ataxia in the second or third decade of life. The diagnosis is usually suggested by increased urinary bound sialic acid excretion. We recently described genetically diagnosed patients with a specially mild phenotype, no retinal abnormalities and normal urinary sialic acid. This observation suggests that genetic analysis or the demonstration of the neuraminidase enzyme deficiency in cultured fibroblasts are needed to detect and diagnose mildest phenotypes.

Key words: cortical myoclonus, neuraminidase, *NEU1*, cortico-muscular coherence, progressive myoclonus epilepsies

Sialidosis was first recognized as a specific neurological disorder in a patient presenting with muscular hypotonia and hypotrophy, ataxia, myoclonus, and seizures, who was later confirmed to have neuraminidase deficiency (Cantz *et al.*, 1977; Spranger *et al.*, 1978). However, it was first recognized as a clear causative factor for progressive myoclonus epilepsy (PME) (Minassian *et al.*, 2016) by Rapin *et al.* (1978) who reported this disorder as ‘cherry-red spot-myoclonus syndrome’ because of the characteristic aspect of the fundus oculi, resulting from storage material in perifoveal ganglionic cells.

Aetiology

The disease presents with variable phenotypes, giving rise to at least two main age-related conditions:

sialidosis type I and II. Both conditions exhibit autosomal recessive inheritance and are caused by mutations of the same gene, *NEU1*, localized on chromosome 6p21.3 (Bonten *et al.*, 1996; Pshezhetsky *et al.*, 1997), which encodes lysosomal neuraminidase (sialidase). Different mutations may account for the variable severity of the disease (Bonten *et al.*, 2000). Indeed, patients with the severe infantile type II disease typically have inactive sialidase, while patients with the milder type I disease have some residual activity. Sialidase is part of a multienzyme complex containing other lysosomal enzymes such as cathepsin A, β -galactosidase, and N-acetyl-galactosamine-6-sulfate sulfatase. The integrity of the multi-enzyme complex ensures the normal catalytic activity of sialidase and protects it against proteolysis.

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NEU1 gene mutations can directly affect the active site or the central core of sialidase, leading to folding defects and retention of sialidase in the endoplasmic reticulum/Golgi compartment, but may also affect the surface region involved in binding to the multienzyme lysosomal complex (Lukong *et al.*, 2001; Pattison *et al.*, 2004).

Sialidase has a central role in removing terminal sialic acid molecules from oligosaccharides and glycoproteins, and its deficiency therefore leads to sialic acid-rich macromolecular storage and urinary sialyl-oligosaccharide excretion.

Neuropathology

Light and electron microscopy reveal cytoplasmic vacuolation involving neurons and perineuronal and interfascicular oligodendroglia, and endothelial and perithelial cells. Vacuolations are associated with diffuse neuronal intracytoplasmic storage of lipofuscin-like pigment which is detectable in the neocortex, basal ganglia, thalamus, brainstem, and spinal cord, as well in extra-nervous organs (Allegranza *et al.*, 1989).

The accumulation of the sialic acid-rich substrates prominently contributes to the pathogenesis of the disease, however, other 'indirect' mechanisms are possibly involved. For instance, it was recently discovered that neuraminidase is a negative regulator of the lysosomal exocytosis of catalytically-active hydrolases (Yogalingam *et al.*, 2008). The resulting increase in extracellular proteolytic activity may lead to premature degradation of other molecules implied in various cellular activities.

Laboratory findings

The laboratory diagnosis is usually supported by increased urinary bound sialic acid excretion and confirmed by genetic analysis or the demonstration of neuraminidase enzyme deficiency in cultured fibroblasts (Lowden & O'Brien, 1979).

Clinical presentation

Sialidosis type I presents with the typical features characterizing PME (Rapin *et al.*, 1978; Lowden & O'Brien, 1979), while the phenotype of *sialidosis type II* includes dysmorphic features (coarse facial features, short trunk, barrel chest, spinal deformity, and skeletal dysplasia), sometimes associated with corneal clouding, hepatomegaly, and inner ear hearing loss. The characteristic macular change found in this metabolic disorder, leading to the definition of 'cherry-red spot', may lead to late visual failure resulting from ganglionic degeneration. The cherry-red spot can, however, be

clinically undetectable for many years and may, moreover, disappear in later stages of the disease (Kivlin *et al.*, 1985). Young-onset cataract formation was also identified in a few patients with type I sialidosis (Thomas *et al.*, 1979).

Both types of sialidosis present with progressively worsening multifocal myoclonus, usually occurring in the second decade of life and variably associated with seizures and ataxia (Lowden & O'Brien, 1979).

Recently, we observed 6 adult patients from 2 different families, presenting high-frequency myoclonus, but no seizures. The disorder progressed slowly and myoclonus was recognized after an interval of many years, although patients displayed a prominent gait disorder with occasional but repeated falls. At the time of our initial diagnostic observation, none of the patients had the cardinal signs suggesting sialidoses, such as macular cherry-red spot or significant urinary sialic acid excretion. Diagnosis resulted from the detection of *NEU1* mutation through genome-wide screening (Canafoglia *et al.*, 2014). Our observation, together with a recent similar report (Schene *et al.*, 2015), points towards the possibility that mild and late forms present with 'cortical myoclonus' and are possibly misdiagnosed.

Most of the patients, with either sialidosis type I or II, become wheelchair-bound within a few years due to severe motor impairment, mainly resulting from severe myoclonus.

Imaging

MRI findings in sialidoses are normal in the early stages, while cerebellar, pontine, and cerebral atrophy can appear during disease progression (Palmeri *et al.*, 2000).

Myoclonus and associated neurophysiological features

In the earliest description of classic PME resulting from type I sialidosis, the authors reported findings similar to those for Unverricht-Lundborg disease, with the exception of photosensitivity that is typically present in Unverricht-Lundborg but not in sialidosis (Engel *et al.*, 1977). As for other types of PME, the subsequent description revealed some degree of phenotypic variability both in terms of the severity of myoclonus and the associated signs of cortical hyperexcitability. In general, the cortical origin of the myoclonus is confirmed by the results of simple back-averaging techniques showing a sharp transient preceding the myoclonus (Franceschetti *et al.*, 1980; Tobimatsu *et al.*, 1985). Since, in patients with sialidosis, myoclonus is often subtle but highly rhythmic and the EEG correlate

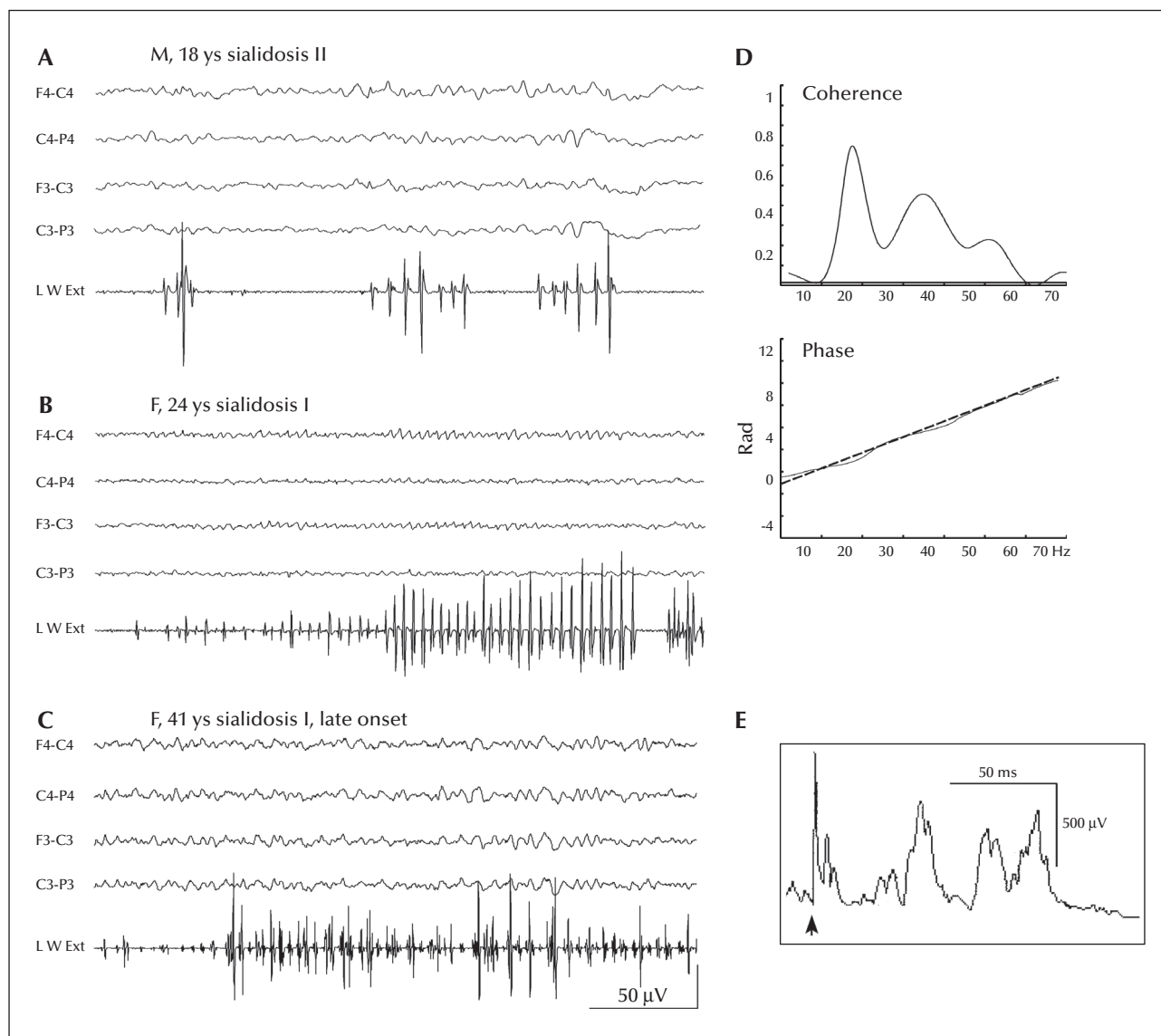


Figure 1. (A, B, C) EEG-EMG recordings performed in 3 patients with sialidosis type II or I. Even if the jerks show a rhythmic course in the all patients, the less severely affected patient with a late onset (C) shows a combination of brief rhythmic sequences and isolated jerks. The panel D shows the coherence and phase function evaluated on EEG-EMG traces in patient B, with an extremely high coherence value and a linear course of the phase indicating a cortical origin of the myoclonic jerks. The panel E shows the multiphasic long-loop response to median nerve stimulation.

consists of a discharge of fast activity, the EEG-EMG coherence analysis appears to be a more reliable method to unequivocally reveal a consistent temporal relationship between the EEG spikes and myoclonic jerks through fast cortico-muscular transfer (Panzica *et al.*, 2003). A study comparing between patients with both type I and II sialidosis and patients with Unverricht-Lundborg disease suggested that this strong rhythmicity and the higher cortico-muscular coherence in sialidoses might account for the particularly severe motor impairment observed in sialidosis patients (Canafoglia *et al.*, 2011). The EEG background

is usually almost normal in patients with type I sialidosis, but polyspike-waves (often associated with spontaneous jerks) are present on the EEG of patients with infantile type II sialidosis.

In some of the reported patients, the presence of high-amplitude somatosensory evoked potentials and enhanced long-loop reflexes (LLR or C-reflex) further confirms the marked neocortical hyperexcitability, which is responsible for 'cortical reflex' and action myoclonus.

The strongly rhythmic recurrence of the jerks reflects on the characteristics of the so-called long-loop

reflexes evoked by median nerve stimulation, which include multiple components resulting from recurrent jerks (Canafoglia *et al.*, 2011). *Figure 1* shows the neurophysiological features of the myoclonus in 3 patients with sialidosis.

Differential diagnosis

Sialidosis type II, presenting in infancy or early childhood with dysmorphic features and skeletal abnormalities, should be differentiated from other storage diseases sharing similar features.

Sialidosis type I, presenting with cortical myoclonus as the main symptom, should be differentiated from other forms of progressive myoclonus epilepsy.

Management

Pharmacological treatment is similar to that for other PMEs (see Nirenberg & Frucht [2005] for a review). Valproate can be considered as the first-line drug, but the treatment of severe myoclonus usually requires two or three additional drugs, including benzodiazepines, levetiracetam, zonisamide or topiramate.

The diversity of clinical phenotypes appears to depend on the type of mutation and the percentage of normal sialidase activity that may protect against most severe forms of the disease. Hence, enzyme replacement therapy is a possible approach to treatment. To date, the effect of enzyme replacement therapy has been evaluated in mouse models. In mice, restored neuraminidase activity persisted for some days, resulting in a significant reduction in lysosomal storage, however, the injected enzyme could not cross the blood-brain barrier. Furthermore, the injected recombinant protein may have induced severe anaphylactic responses (Wang *et al.*, 2005). □

Disclosures.

None of the authors have any conflict of interest to disclose.

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Myoclonus epilepsy in mitochondrial disorders

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ABSTRACT – Mitochondrial disorders is a group of clinical entities associated with abnormalities of the mitochondrial respiratory chain (MRC), which carries out the oxidative phosphorylation (OXPHOS) of ADP into ATP. As the MRC is the result of genetic complementation between two separate genomes, nuclear and mitochondrial, OXPHOS failure can derive from mutations in either nuclear-encoded, or mitochondrial-encoded, genes. Epilepsy is a relatively common feature of mitochondrial disease, especially in early-onset encephalopathies of infants and children. However, the two most common entities associated with epilepsy include MERRF, for Myoclonic Epilepsy with Ragged Red Fibers, and AHS, or Alpers-Huttenlocher syndrome, also known as hepatopathic poliodystrophy. Whilst MERRF is a maternally inherited condition caused by mtDNA mutations, particularly the 8344A>G substitution in the gene encoding mt-tRNA^{Lys}, AHS is typically caused by recessive mutations in *POLG*, encoding the catalytic subunit of polymerase gamma, the only mtDNA polymerase in humans. AHS is the most severe, early-onset, invariably fatal syndrome within a disease spectrum, which also include other epileptogenic entities, all due to *POLG* mutations and including Spino-cerebellar Ataxia and Epilepsy (SCAE). This review reports the main clinical, neuroimaging, biochemical, and molecular features of epilepsy-related mitochondrial syndrome, particularly MERRF and AHS.

Key words: MERRF, MELAS, Alpers-Huttenlocher syndrome, hepatopathic poliodystrophy, mitochondrial DNA, oxidative phosphorylation, mitochondrial respiratory chain, progressive myoclonus epilepsies

The term 'mitochondrial disorders' is, to a large extent, applied to the clinical syndromes associated with abnormalities of the common final pathway of mitochondrial energy metabolism, *i.e.* oxidative phosphorylation (OXPHOS). OXPHOS takes place in the inner mitochondrial membrane involving five enzymatic complexes which form the mitochondrial respiratory chain (MRC). From a genetic perspective, the MRC is unique since it is encoded

by two complementary separate genetic systems: the nuclear and the mitochondrial genomes. Because of this dual genetic control, OXPHOS disorders may be due to mutations in mitochondrial deoxyribonucleic acid (mtDNA) or nuclear DNA genes encoding either structural components of the MRC complexes or factors controlling their expression, assembly, function and turnover (Smeitink *et al.*, 2001). Mitochondria contain the only extra-nuclear

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source of DNA in animal cells (Nass, 1966). MtDNA is a circular, double stranded, 16,569 base-pair molecule of DNA which encodes 37 genes, including 13 polypeptides essential for the formation and function of four of the five MRC complexes, namely complex I, III, IV and V, two ribosomal RNAs (12S and 16S rRNA), and 22 transfer RNAs (tRNA) (Anderson *et al.*, 1981). All other OXPHOS-related proteins, including most of the MRC subunits, MRC assembly factors, factors necessary for mtDNA maintenance and expression, etc., are synthesized in the cytosol and are specifically targeted, sorted and imported to their final mitochondrial location (Mokranjac & Neupert, 2005). The mitochondrial genome has unique features that distinguish it from the nuclear genome; for instance, in sexuate organisms, mtDNA is strictly maternally inherited and present in several hundred to several thousand copies within a single cell, the number varying amongst different cell types, mostly based on the energy demand of each tissue and organ (Taylor & Turnbull, 2005). The mtDNA genes have no introns, hardly any non-coding intervening regions, and for most cases the termination codons are completed by post-transcriptional polyadenylation (Anderson *et al.*, 1981). The genetic code of mtDNA differs between many species, including humans, such that genomes between species may be reciprocally untranslatable; this partly explains why mtDNA is translated *in situ* by protein synthesis machinery that is completely independent from that operating in the cytosol for the translation of nuclear genes. In human mtDNA, the displacement loop (D-loop) is the only major non-coding region of the molecule which is formed by the displacement of the two DNA strands by a third DNA strand, the so-called 7S DNA.

An important contribution to the elucidation of the molecular basis of mitochondrial disorders has come from the discovery of an impressive, ever-expanding number of pathogenic mutations in mtDNA. In cases in which a mtDNA mutation is not found, mitochondrial disease is defined by the detection of a specific biochemical defect in OXPHOS, or the observation of typical morphological clues, or a combination of the two. In many instances, pathogenic mtDNA mutations can coexist alongside non-mutated mtDNA in the same cell, tissue and organism, a condition known as heteroplasmy (Hayashi *et al.*, 1991; Larsson & Clayton, 1995). The percentage of pathogenic heteroplasmy dictates the phenotype, according to a 'threshold effect', i.e. a critical amount below which mutations do not manifest any clinical or biochemical phenotype. This threshold level varies from tissue to tissue and depends on the intrinsic pathogenicity of each mutation but, in general, ranges from 50-60 per cent (DiMauro, *et al.*, 1985; Rosing *et al.*, 1985; Mita *et al.*, 1990; Hayashi *et al.*, 1991; Moraes *et al.*, 1992;

Shoubridge, 1994; Traff *et al.*, 1995; Parikh *et al.*, 2008) for the most severe mutations, to more than 90 percent for the mildest mutations. A paradigmatic example is the m.8993T>G mutation associated with NARP (neuropathy, ataxia, retinitis pigmentosa) syndrome (DiMauro, *et al.*, 1985). The relationship between the mutation load and clinical severity was first documented by Tatuch *et al.*, who showed that around 70 percent heteroplasmy in skeletal muscle results in adult-onset of a slowly progressive syndrome corresponding to the acronymic features of NARP, whereas higher degrees of heteroplasmy (around 90 percent) cause severe, early-onset, maternally inherited Leigh syndrome (MILS) (Rosing *et al.*, 1985).

MtDNA disease has an extremely variable phenotype and can present at any age (Traff *et al.*, 1995). The clinical features usually affect tissues characterized by high metabolic demand, such as the central nervous system, the skeletal muscle, or the heart. However, other tissues are frequently involved, such as the β cells in the pancreas (leading to diabetes), the hair cells of the cochlea (causing deafness), or the renal tubules (leading to kidney dysfunction).

While epilepsy is a recurrent manifestation of mitochondrial disease, its exact prevalence is not known. In contrast, 35-60 per cent of individuals with refractory seizures display biochemical evidence of mitochondrial dysfunction (Parikh *et al.*, 2008). A few studies have systematically examined the epileptic manifestations of mitochondrial disease (Khurana *et al.*, 2008; Lee *et al.*, 2008; El Sabbagh *et al.*, 2010). Although seizures may be the presenting symptom at onset (Hayashi *et al.*, 1991; Canafoglia *et al.*, 2001), in more than 80 percent of cases, the first seizure is preceded by some other symptoms (El Sabbagh *et al.*, 2010), including, for example, failure to thrive, developmental delay, ataxia, or evidence of multi-organ involvement. In children with respiratory chain disorders, different seizure types can occur in as many as 60 percent of cases (El Sabbagh *et al.*, 2010). Whilst clinical identification of mitochondrial epilepsy may be difficult, one of the most common forms is myoclonic epilepsy, either as typical MERRF syndrome (myoclonic epilepsy with ragged red fibres) or within the context of other, complex epileptic manifestations.

Clinical manifestations

In 1921, Ramsay Hunt described six patients with a disorder characterized by ataxia, myoclonus, and epilepsy, which he called '*dyssynergia cerebellaris myoclonica*' (Hunt, 1921). Over 50 years then passed before Tsairis *et al.* linked this entity to mitochondrial abnormalities in the skeletal muscle in one family, hallmarked by the presence of ragged-red fibres

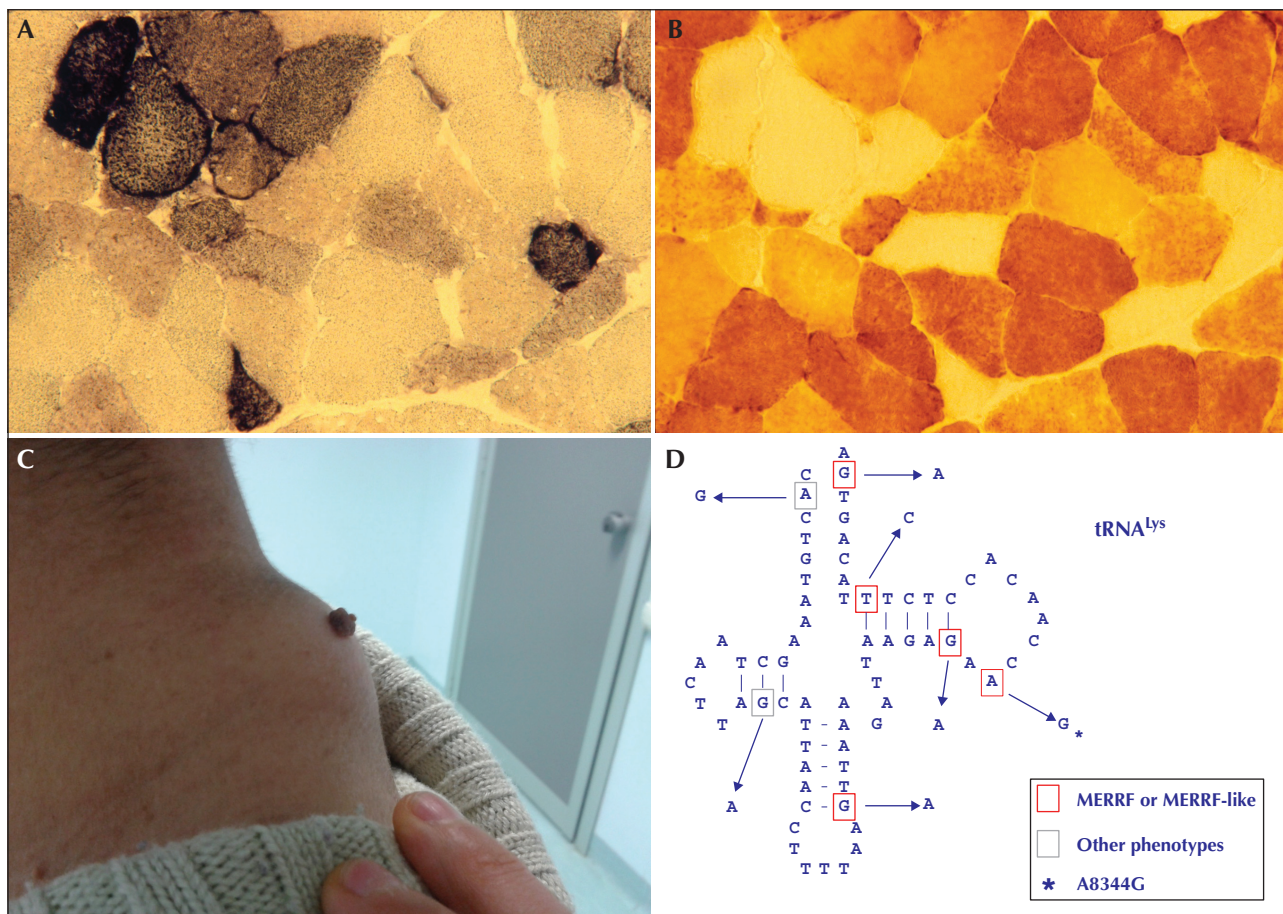


Figure 1. Muscle biopsy of a MERRF patient presenting with COX deficient (A) and intense SDH-positive (B) RRF. (C) Neck lipoma in a 67-year-old MERRF patient. (D) Lys tRNA highlighting the most frequent mutations associated with MERRF or MERRF-Like syndrome.

(Tsairis *et al.*, 1973). This family was described in great detail, leading to the term ‘classic MERRF’ in 1989 (Lombes *et al.*, 1989). MERRF was one of the three major, multisystem syndromes first classified as ‘mitochondrial encephalomyopathy’ (DiMauro *et al.*, 1985). MERRF has two other historical distinctions:

- it was the first well-defined human disease in which maternal inheritance was clearly demonstrated, thus suggesting a mitochondrial DNA defect (mtDNA) (Rosing *et al.*, 1985);
- the first mitochondrial encephalomyopathy in which a molecular mtDNA defect was actually identified.

It is also one of the most common and clinically better defined mitochondrial syndromes. MERRF is, in fact, a multi-system disorder, hallmarked by myoclonus, episodes of generalized epilepsy, progressive ataxia, and ragged-red fibres (RRF) with partial deficiency of cytochrome c oxidase (COX) (19–20–21) (figure 1 A, B). Although the onset is usually in childhood, early development is normal and adult onset is not uncommon. Besides the defining criteria, common clinical manifestations include hearing

loss, peripheral neuropathy, cognitive decay and eventually dementia, short stature, exercise intolerance, and optic atrophy and ataxia. Less common clinical signs (seen in < 50 percent of the patients) include cardiomyopathy, pigmentary retinopathy, pyramidal signs, ophthalmoparesis, and the appearance of multiple lipomas, particularly in the neck and upper trunk (figure 1C). As is usually the case with mitochondrial encephalomyopathies, maternal family members may be symptomatic, oligosymptomatic, or asymptomatic. While several heteroplasmic point mutations, mostly affecting the gene encoding mt-tRNA^{Lys}, are responsible for MERRF, by far the most frequent MERRF mutation is the m.8344A>G substitution in the T-Ψ-C loop of mt-tRNA^{Lys} (figure 1D). A few unusual clinical presentations, characterized by overlapping symptoms between MERRF and MELAS (mitochondrial encephalomyopathy with lactic acidosis and stroke-like episodes), have been reported either as isolated cases or, more often, in pedigrees in which typical MERRF patients were also present. Several studies have reported in detail the wide spectrum of clinical

presentations associated with the m.8334A>G mutation (Berkovic *et al.*, 1991; Hammans *et al.*, 1993; Silvestri *et al.*, 1993; Howell *et al.*, 1996; Mancuso *et al.*, 2013), but certain features and variants deserve special attention. For instance, peripheral neuropathy is not uncommon in MERRF, being usually sensory-motor and contributing to the onset and progression of gait ataxia. At least one case has been reported to be characterized by predominant motor symptoms, thus phenocopying Charcot-Marie-Tooth disease (Howell *et al.*, 1996).

In 1975, long before the molecular defects of MERRF became known, Karl Ekbom (Ekbom, 1975) described multiple lipomas in association with hereditary ataxia, photomyoclonus and skeletal deformities in a family in which the 8344A>G mutation was later documented (Berkovic *et al.*, 1991; Traff *et al.*, 1995). These tumours, varying in size from small subcutaneous nodules to disfiguring masses, are usually located in the nape of the neck and the shoulder area. They have been reported in numerous patients with the 8344A>G mutation (Larsson *et al.*, 1992; Holme *et al.*, 1993; Calabresi *et al.*, 1994; Naumann *et al.*, 1995; Austin *et al.*, 1998). Maternal inheritance was evident in a large family (Rosing *et al.*, 1985) which was again confirmed to harbour the m.8344A>G substitution in the mt-tRNA^{Lys} gene (Shoffner *et al.*, 1990). Not only was this the first molecular defect to be reported for a mitochondrial encephalomyopathy, but also the first to be identified for a specific form of epilepsy. The m.8344A>G mutation is present in about 90 per cent of MERRF patients. Two additional mutations have been associated with MERRF, both affecting the mt-tRNA^{Lys} gene. The first mutation, m.8356T>C, was discovered simultaneously in an American family with typical MERRF (Silvestri *et al.*, 1992), and in an Italian family in which typical MERRF symptoms coexisted with stroke-like episodes and migraines, thus justifying the definition as a “MERRF/MELAS” overlap syndrome (Zeviani *et al.*, 1993). A common clinical feature of both families was hyperthyroidism, which is rather unusual in mitochondrial diseases and may, therefore, be related to this specific mutation. A third family with the same mutation was later reported in Japan; the proband had typical MERRF, but a maternal aunt had stroke-like episodes, another example of MERRF/MELAS overlap (Sano *et al.*, 1996).

The second mutation, m.8363G>A, was first identified in two unrelated American families with maternally inherited cardiomyopathy, which was severe enough to cause early death in several members of one family (Santorelli *et al.*, 1996). Although cardiomyopathy dominated the clinical picture, additional signs included encephalomyopathy, neurosensory hearing loss, progressive external ophthalmoparesis,

intellectual disability, limb weakness, and peripheral neuropathy, variably affecting members of both families. Interestingly, cerebellar symptoms were frequent, including ataxia, dysmetria, slurred speech, and gait instability. Interestingly, one proband had ‘horse collar’ lipomas. The same mutation was later identified in two unrelated Japanese patients with typical MERRF, one of whom also presented cardiomyopathy (Ozawa *et al.*, 1997). Recently a new mutation, m.3291T>C, has been associated with MERRF/MELAS syndrome in a 19-year-old Chinese man (Liu *et al.*, 2014).

The m.8344A>G mutation is virtually always heteroplasmic, and the mutation threshold for typical MERRF is usually high or very high (*i.e.* affecting about 60–90 per cent of total mtDNA), which suggests that this mutation is relatively benign (Shoffner *et al.*, 1990). As mentioned earlier, the three mutations associated with MERRF all affect highly conserved nucleotides in the mt-tRNA^{Lys} gene. The m.8344A>G and the m.8356T>C mutations are located in the T-Ψ-C loop, while the m.8363G>A mutation is located in the aminoacyl acceptor stem of the putative cloverleaf (secondary) structure of the mt-tRNA^{Lys} transcript (Santorelli *et al.*, 1996). A single patient with typical MERRF symptoms, *i.e.* myoclonus, seizures, ataxia, and RRF, but, in addition, peripheral neuropathy, dementia, and neuroradiological evidence of mild cerebral and severe cerebellar atrophy, had no mutation in the tRNA^{Lys} gene, but rather multiple mtDNA deletions in muscle, suggesting impairment in a nuclear gene product controlling mtDNA integrity, such as POLG.

Laboratory tests

Patients with typical MERRF have elevated blood lactate and pyruvate at rest, both increasing abnormally upon moderate exercise. CSF protein levels are often increased, but rarely exceed 100 mg%. Electromyography and nerve conduction studies are usually compatible with a predominantly myopathic pattern, except when motor peripheral neuropathy is clearly present. Electroencephalography is typically characterized by generalized spike and wave discharges with background slowing, but focal epileptiform discharges may also be seen (*figure 2*).

A CT or MRI scan may show brain atrophy and basal ganglia calcifications. Phosphorus magnetic resonance spectroscopy of the gastrocnemius muscle in eight patients (only three of whom showed signs of myopathy) revealed mitochondrial dysfunction in all, as evidenced by increased relative intracellular inorganic phosphate (Pi) concentration and decreased phosphocreatine to Pi ratio (Rahman, 2012). However, no mitochondrial dysfunction was seen in the brain using the same technique.

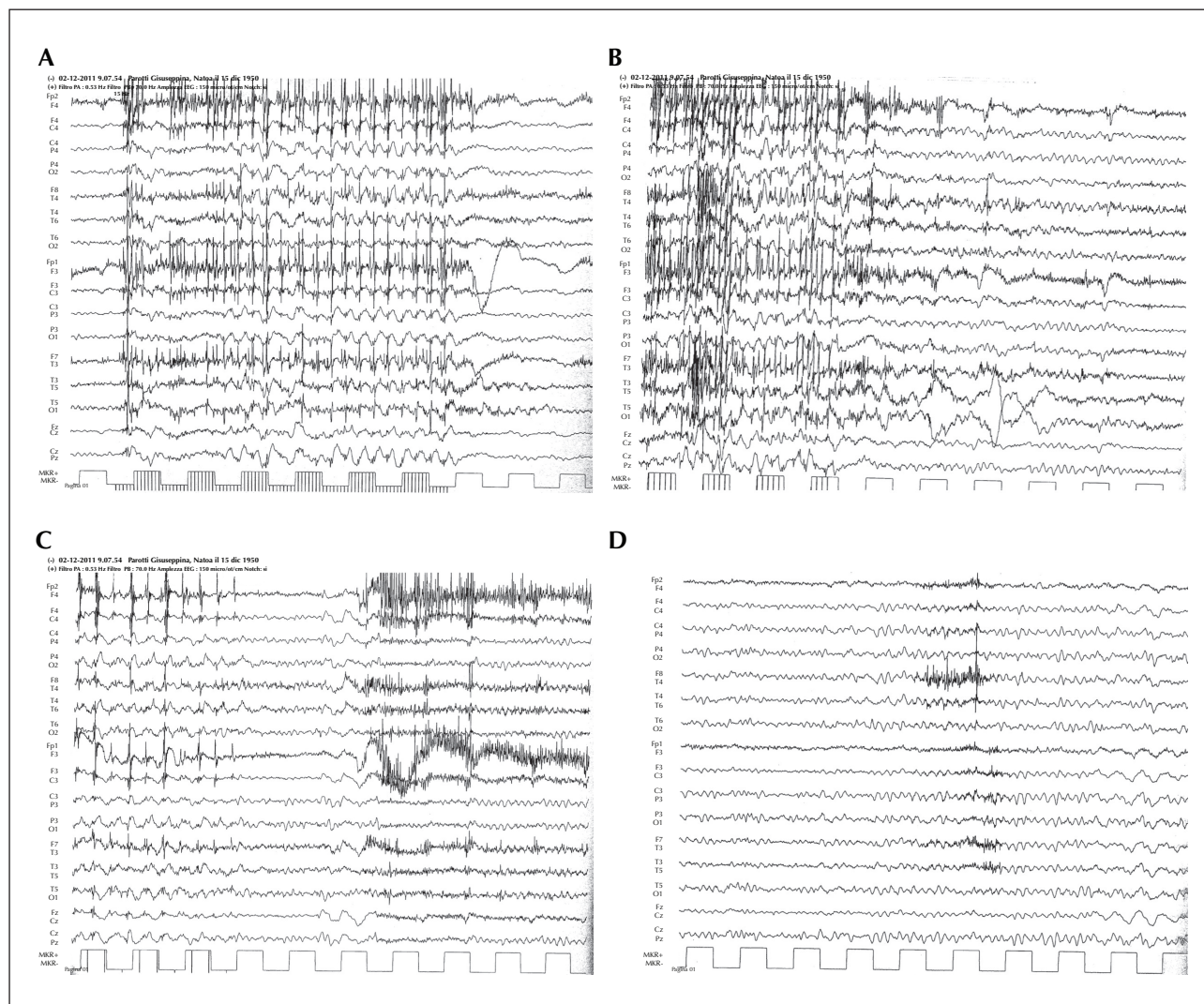


Figure 2. EEG in adult patients with MERRF. (A, B) Generalized spike and waves, as well as myoclonus, during SLI. (C, D) Paroxysmal activity in the temporal region in both hemispheres with right prevalence.

By definition, the muscle biopsy shows RRF using modified Gomori trichrome stain in typical patients (Wolf *et al.*, 2009). These fibres also react intensely to the succinate dehydrogenase (SDH)-specific stain, a more sensitive indicator of excessive mitochondrial proliferation. Both RRF and some non-RRF fail to stain histochemically to COX. Muscle biopsies from MERRF patients can also show strongly SDH-reactive blood vessels (SSVs), similar to those characteristically seen in the muscles of patients with MELAS (Harding, 1990), again emphasizing the concept that the two disease entities may overlap to some extent. However, in contrast to MELAS SSVs, which stain positive for COX, MERRF SSVs are uniformly COX-negative (Lamantea *et al.*, 2002). MRC activities in muscle extracts usually show defects in mtDNA-dependent complexes, particularly COX (Lombes *et al.*, 1989; Silvestri *et al.*, 1993).

Neuronal loss and gliosis predominate in the brains of MERRF patients, preferentially involving the cerebellum, the brainstem, and the spinal cord. In the cerebellum, neuronal loss is particularly severe in the dentate nucleus, an observation originally made by Ramsay Hunt, who described 'primary atrophy of the dentate system' in patients with 'dyssynergia cerebellaris myoclonica' (Hunt, 1921). The inferior olivary nucleus of the medulla oblongata is the most severely affected structure in the brainstem, followed by the red nucleus and the substantia nigra in the mesencephalon. In the spinal cord, severe cell loss has been observed in the thoracic nucleus of Clarke, while milder involvement has been detected in the anterior and posterior horns of the spinal cord. Demyelination preferentially affects the superior cerebellar peduncles and the posterior columns and

lateral spinocerebellar tracts of the spinal cord, while the pyramidal system is usually spared or mildly affected.

Private mtDNA mutations and myoclonic epilepsy

Although MERRF is one of the most common mitochondrial encephalomyopathies, a substantial fraction of paediatric patients with myoclonic epilepsy and MRC defects fails to show MERRF mutations (El Sabbagh *et al.*, 2010). In this group, severe myoclonus may be preceded by other seizure types, such as erratic myoclonus, focal motor seizures, myoclonic absences, or tonic seizures, but becomes predominant during the disease course. Occasionally, the disease may evolve into recurrent myoclonic *status epilepticus*. In the report by El Shabbagh *et al.* on 56 paediatric patients (El Sabbagh *et al.*, 2010), only one showed photosensitivity, with spikes induced by intermittent light stimulation. Brain MRI showed hyper-intense signals on T2/FLAIR (fluid-attenuated inversion recovery) sequences of the basal ganglia ($n = 8$) and/or dentate nuclei ($n = 3$), and cerebellar atrophy ($n = 5$). The basal ganglia were involved irrespective of the age at onset, while cerebellar involvement was present only in patients with early-onset epilepsy, *i.e.* within the first decade of life. Myoclonic seizures were drug-resistant. Six patients died from global neurological failure leading to comatose *status*. Biochemically, eight patients had complex I deficiency, three complex V, two complex IV, one complex II, and four showed multiple defects. In 45 percent of the cases, a mtDNA mutation was identified; in *MT-ATP6* ($n = 3$), *MT-ND3* ($n = 2$), *MT-TK* ($n = 1$), *MT-ND5* ($n = 1$), and *MT-TL1* ($n = 1$). mtDNA depletion in muscles is only rarely associated with progressive myoclonic epilepsy (Rahman, 2012; Minassian *et al.*, 2016).

Alpers-Huttenlocher syndrome

The second most common form of mitochondrial myoclonic epilepsy is Alpers-Huttenlocher syndrome (AHS) (OMIM #203700). AHS is clinically characterized by psychomotor retardation, intractable epilepsy, and liver failure. The onset is in infancy or early childhood, often with seizures and/or hypotonia. *Status epilepticus* is common and most patients die from refractory seizures and liver failure before the age of 3 (Luoma *et al.*, 2004). Liver dysfunction can be present at the onset of the neurological symptomatology, or may manifest following treatment with sodium valproate for the control of seizures. Individuals with AHS

typically present with focal myoclonic and complex seizures. *Epilepsia partialis continua* is also frequently seen and may lead to fatal *status epilepticus*.

The electroencephalogram at onset may point towards the diagnosis, particularly when characterized by unilateral occipital, rhythmic, high-amplitude slow activity with superimposed (poly)spikes, frequently evolving into generalized discharges (Wolf *et al.*, 2009).

Other clinical features include global developmental delay and regression, progressive microcephaly, cortical visual impairment with abnormal visual-evoked potentials, and, importantly, evidence of progressive liver failure, heralded by elevated levels of liver enzymes in the blood and hepatomegaly, followed by overt liver cirrhosis. Brain MRI may be normal in the initial stage of disease, or show non-specific changes, such as progressive cerebral atrophy.

AHS was firstly diagnosed as a neuropathological entity defined by the presence of extensive necrotizing poliodystrophy; histological features include spongiosis, neuronal loss and astrogliosis affecting the cerebral cortex, particularly the calcarine cortex, which explains the cortical visual loss in this condition (Luoma *et al.*, 2004; Van Goethem *et al.*, 2004; Davidzon *et al.*, 2005). Liver histology in AHS may reveal steatosis, hepatocyte loss, bile duct proliferation and fibrosis, evolving into frank cirrhosis (Harding, 1990). By and large, AHS is associated with a few recessive mutations in the *POLG* gene, encoding the catalytic, large subunit of mtDNA polymerase (polymerase gamma). More than 150 mutations have been reported in the *POLG* gene, constituting a major cause of mitochondrial disease. Mutations in this gene are also the most frequent cause of autosomal dominant progressive external ophthalmoplegia (ad-PEO). In adPEO due to *POLG* mutation, distinct features also include severe dysphagia and dysphonia and, occasionally, extra-pyramidal signs, *e.g.* parkinsonism, cerebellar dysfunction, or chorea (Luoma *et al.*, 2004). Recessive mutations of *POLG* may also be responsible for autosomal recessive cases (Lamantea *et al.*, 2002) or apparently sporadic PEO cases characterised by multiple mtDNA deletions (Agostino *et al.*, 2003), with or without additional findings, including parkinsonism, severe peripheral neuropathy, endocrine failure, or psychotic depression (Van Goethem *et al.*, 2003).

In addition, recessive *POLG* mutations are responsible for a wide spectrum of syndromes of increasing severity and precocity, including juvenile sensory ataxic neuropathy, dysarthria, ophthalmoplegia, SANDO (Horvath *et al.*, 2006), childhood cerebellar ataxia and epilepsy, SCAE, and possibly infantile AHS, all characterized by exquisite sensitivity of the liver to valproate-associated damage.

As mentioned above, liver failure occurs spontaneously in AHS, due to severe, liver-specific mtDNA depletion.

The molecular basis of this clinical heterogeneity can be explained, in part, by the structural and functional complexity of the enzyme. Pol- γ A, the 145 kDa catalytic subunit encoded by *POLG*, comprises an N-terminal exonuclease domain, with predominantly proofreading functions, and a polymerase domain, which performs the template-directed synthesis of the nascent mtDNA strands. The 2 most prevalent mutations in AHS, but also in SANDO and SCAE, are p.A467T and p.W748S, which are present at a frequency of approximately 1 percent in the Scandinavian population (Horvath *et al.*, 2006). Rapid molecular diagnosis of AHS syndrome may therefore be achieved by screening these selected 'common' mutations in DNA extracted from blood, however, in several cases, sequence analysis of all exons and exon-intron boundaries is required to identify causative *POLG1* mutations. Liver biopsy of AHS patients shows severe mtDNA depletion while multiple mtDNA deletions are the molecular hallmark in muscle for adPEO or arPEO patients.

Recently, the combination of early encephalopathy, epilepsy, hepatopathy, and sensory axonal neuropathy was found in patients with recessive mutations in the mtDNA helicase, Twinkle (*PEO1*), which co-functions with Pol- γ in mtDNA replication (Saneto & Naviaux, 2010; Lonnqvist *et al.*, 2009). As is the case for *POLG*-associated AHS, patients with recessive mutations in Twinkle also display mtDNA depletion in the liver.

Therapy

There is no specific therapy for MERRF or other mitochondrial encephalomyopathies associated with myoclonic epilepsy. Patients are empirically treated with "cocktails" of vitamins and cofactors, including idebenone at high dosage (150 mg x3 daily) and L-carnitine (1 g daily) (Farge *et al.*, 2007). Myoclonus can be controlled with clonazepam (0.5-1 mg three times a day) or zonisamide. As with all mitochondrial diseases, valproate has to be used with caution and always in combination with L-carnitine because of its well-documented inhibition of carnitine uptake (DiMauro *et al.*, 2000). Hepatic impairment in AHS, SANDO or SCAE may be precipitated by valproate, leading to fulminant liver failure. These conditions, therefore, represent an absolute contra-indication for the use of valproate to control seizures (Tein *et al.*, 1993). Lactic acidosis can be controlled by bicarbonate which, however, has only a transient buffering effect and may exacerbate the cerebral symptoms. Levetiracetam is

the first choice of treatment for myoclonus in MERRF and lamotrigine may exert a neuroprotective effect (Lagruet *et al.*, 2007). □

Disclosures

None of the authors have any conflict of interest to disclose.

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Progressive myoclonus epilepsy associated with neuroserpin inclusion bodies (neuroserpinosis)

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ABSTRACT – Familial encephalopathy with neuroserpin inclusion bodies (FENIB) is a conformational proteinopathy characterised by neuronal inclusion bodies composed of the serine protease inhibitor (SERPIN), neuroserpin. Presenting clinically as a familial dementia-epilepsy syndrome, the molecular mechanism of the pathogenic abnormalities in neuroserpin has been characterised at atomic resolution. There is a remarkable genotype-phenotype correlation between the degree of molecular destabilisation of the several variants of the neuroserpin protein, their propensity to self-associate and the age of onset of the dementia-epilepsy complex. As with other serpinopathies there appears to be a mix of cell-autonomous toxicity, due to neuronal accumulation of neuroserpin, and non-cell autonomous toxicity, caused by loss of protease inhibition, in this case the dysregulated protease is likely to be tissue plasminogen activator (tPA). FENIB should be considered in cases of progressive myoclonic epilepsy and dementia particularly where there is family history of neuropsychiatric disease.

Key words: serpins, serpinopathy, protease inhibitor, conformational disease, neuroserpin, dementia, familial progressive myoclonic epilepsy, FENIB, progressive myoclonus epilepsies

Familial encephalopathy with neuroserpin inclusions bodies (FENIB) has been identified as a cause of pre-senile dementia with frontal symptoms as well as progressive myoclonic epilepsy (Minassian *et al.*, 2016). In affected individuals, mutated neuroserpin accumulates in neuronal inclusions (Collins bodies). FENIB is due to mutations in the *SERPINI1* gene located on chromosome 3q26. Serine protease

inhibitors (serpins) are a large superfamily of proteins that employ a conserved molecular mechanism for the inhibition of a wide range of proteases. Mutations may render serpins prone to polymerisation, *i.e.* ordered aggregation, in the endoplasmic reticulum of the synthetic Acell. In the case of neuroserpin (NS), a neuronal protein, this aggregation causes gain-of-function neuronal dysfunction

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that is thought to underpin the neurological manifestations. NS also inhibits tPA and, in this way, is thought to regulate the sensitivity of neurones to glutamatergic excitatory neurotransmission at the NMDA (N-methyl-D-aspartate) receptor. The epileptic component of FENIB may be due to dysfunction of the NS/tPA pathway.

Neuroserpin (NS) is a member of the serine protease inhibitor, or serpin, superfamily and has sequence and structural homologies to the archetype α_1 -antitrypsin. At the molecular level, serpins are composed of 9 α -helices, 3 β -sheets (A to C) and an exposed mobile reactive centre loop (RCL). The RCL typically contains 20 residues that act as a pseudo-substrate for the target protease (Elliott *et al.*, 1996; Ryu *et al.*, 1996). The main role of NS is to regulate the plasmin proteolytic pathway, in particular by inhibiting tissue-type plasminogen activator (tPA). In this regard, NS plays physiological roles in the development of the central nervous system (Seeds *et al.*, 1999), in learning and memory and also in such pathological events as stroke (Cole *et al.*, 2007) and epilepsy (Qian *et al.*, 1993).

Numerous mutations in human serpins have been linked to a wide range of diseases; examples include emphysema, angioedema, and dementia with progressive myoclonus epilepsy, resulting from mutations in α_1 -antitrypsin, C1-inhibitor, and neuroserpin, respectively. Direct toxicity is invariably a consequence of the intracellular accumulation of serpin aggregates that are termed polymers, and may result in death of the synthetic cell. In the case of neuroserpin, cytotoxicity is seen exclusively in neurons of the central nervous system. There may also be associated loss-of-function effects caused by the deregulated hyperactivity of the target proteases and this may underpin the development of epilepsy in patients carrying neuroserpin variants. To date, only a few families and exceptionally rare non-familial cases of progressive myoclonic epilepsy linked to the *SERPIN1* gene have been described. Age at onset is between 13 and 60 years and the disorder is severe with rapid loss of autonomy and premature death.

Pathophysiology of neuroserpin-related disease

Neuroserpin was first identified as an axonally-secreted glycoprotein in the conditioned medium of embryonic chick dorsal root ganglia cells (Stoeckli *et al.*, 1989; Osterwalder *et al.*, 1996). In mammals it is expressed in the central and peripheral nervous system, predominantly in neurones (Hastings *et al.*, 1997; Osterwalder *et al.*, 1998) where it can inhibit a number of trypsin-like enzymes, including thrombin and plasmin (Osterwalder *et al.*, 1998), however, subsequent

studies indicate that PA is the main physiological target. Accordingly, the highest levels of NS are found in parts of the nervous system that have the highest expression of tPA mRNA and protein (Krueger *et al.*, 1997).

While not all serpins interact with proteases, those such as neuroserpin, which have retained inhibitory activity, employ a highly conserved molecular mechanism. The sequence of events that results in inhibition begins with the serpin's RCL (labelled in red in *figure 1M*) entering the active site of the cognate protease, initially behaving as a substrate. The enzyme and serpin form a transient intermediate called the Michaelis complex (Ye *et al.*, 2001) which precedes the cleavage of the RCL at a specific position, termed the P1-P1' bond. This leads to the release of the P1 residue and the formation of an ester bond between the C-terminal portion of the serpin and the protease. At this stage, however, further hydrolysis, that would otherwise release cleaved serpin and active protease, is prevented by a profound conformational change in the serpin (Wright & Scarsdale, 1995). This transition from a "stressed to relaxed" form occurs as the protease-attached remainder of the RCL inserts as a α -strand in the main β -sheet A of the serpin core. This insertion is highly energetically favourable and violently flips the enzyme from the upper to the lower pole of the serpin (Huntington *et al.*, 2000), causing steric denaturation and inactivation of the protease. Permanent destruction of the protease is subsequently achieved by third-party proteolysis of those domains of the target protease that have been rendered unstructured (Huntington *et al.*, 2000). Experimentally, the enzyme-serpin complex is relatively stable with a dissociation rate constant that is an order of magnitude slower than the association rate (Belorgey *et al.*, 2002). However, dissociation of the serpin-enzyme complex does occur, in which case the released neuroserpin is in the inactive, cleaved conformation. In contrast, when tPA is liberated from the complex, it retains its proteolytic activity (Osterwalder *et al.*, 1998).

Clinical features and genotype-phenotype correlations

Mutations in the neuroserpin gene were first reported to cause an autosomal dominant form of pre-senile dementia (Belorgey *et al.*, 2002), described as familial encephalopathy with neuroserpin inclusion bodies (FENIB), characterized histologically by unique neuronal inclusion bodies and biochemically by polymers of the neuron-specific serpin, neuroserpin. These authors reported 2 unrelated Caucasian families living in the United States, carrying 2 different heterozygous mutations (S49P and S52R). In the larger family, affected individuals presented clinically around the

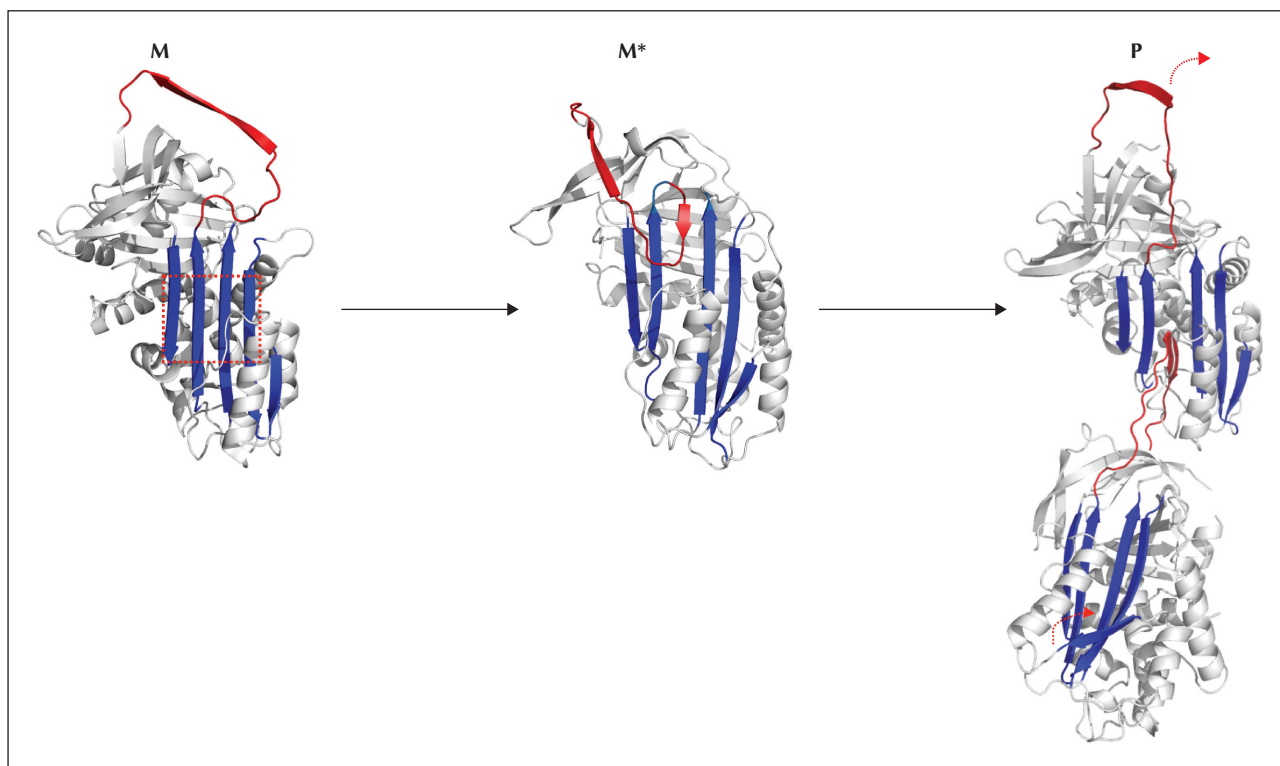


Figure 1. Molecular models of serpin conformers. M represents the active protease inhibitor with the reactive loop shown in red and β -sheet A in blue. Mutations that destabilize sheet A (typically in the so-called 'shutter region' signified by the red dashed box) allow the partial insertion of the reactive loop to form a new strand in sheet A (M^*). The consequent opening of the sheet allows the reactive loop from another serpin to insert forming a serpin dimer (P). The repeated addition of serpins by loop-sheet polymerisation in this way results in large aggregates.

fifth decade of life with cognitive decline, including deficits in attention and concentration, response regulation difficulties, and impaired visuospatial skills. Memory was also impaired, but to a lesser degree than is typically seen in individuals with Alzheimer's disease. After several years of disease progression, most affected individuals were unable to work and eventually required nursing-home care. The second, much smaller family showed an earlier clinical onset, during the second or third decade of life. Affected individuals presented with both epilepsy and progressive cognitive decline and their neuro-histology was dominated by eosinophilic, PAS-positive intraneuronal inclusions in the brain. Later on, a small family with 2 affected siblings featuring progressive myoclonus epilepsy and dementia was described (Belorgey *et al.*, 2002). The affected individuals developed generalized seizures in adulthood and progressed to status epilepticus over several years. In addition, they also developed slow speech, diplopia, nystagmus, dysarthria, and myoclonus in the extremities, with rapidly progressive dementia. Their deceased mother was, reportedly, similarly affected.

Davis *et al.* (1999a) reported additional patients with the disorder: a 23-year-old man with an 8-year history of

progressive myoclonic epilepsy, dementia, tremor, and dysarthria. The second patient was a 13-year-old girl with progressive myoclonus epilepsy with intractable seizures, myoclonus, and dementia. She died at the age of 19 during status epilepticus. Her father was said to be 'mentally deficient', and a paternal uncle had died from epilepsy at the age of 18.

Gourfinkel-An *et al.* (2007) subsequently described a small French family with progressive myoclonic epilepsy associated with a frontal syndrome starting from the age of 18 with severe myoclonus, generalized tonic-clonic seizures, and absences. The EEG of one of the patients showed diffuse slow waves, spikes, and spike-wave discharges superimposed on a slow background, with photic sensitivity at around 1 Hz. Cerebral MRI revealed cortico-subcortical atrophy. The patient's condition progressively worsened and swallowing difficulties were noticed at an early stage. Additionally, cerebellar symptoms and pyramidal signs were also present. Cognitive deterioration was severe (Mini-Mental Status Examination score: 12/30). Signs of frontal dysfunction were observed (emotional lability, distractibility, and poor performance on sequential motor tests) with sparing of long-term explicit memory. In another affected relative, who died at the age of 33,

Table 1. Various mutations in the neuroserpin gene cause onset of dementia and myoclonic epilepsy at a wide range of ages. The most aggregation-prone neuroserpin variants *in vitro* cause juvenile forms of FENIB while less destabilizing mutations are not penetrant until the seventh decade. Histologically, patients with early onset disease exhibit a higher burden of cortical neuroserpin inclusions (+ through to ++++).

| Mutation | Onset (years) | Clinical findings | Inclusions |
|----------|---------------|--|------------|
| S49P | 45-63 | Dementia and terminal seizures | + |
| S52R | 20-40 | Myoclonus, Dementia | ++ |
| L47P | 20-30 | Progressive myoclonus epilepsy | +++ |
| H338R | 15 | Progressive myoclonus epilepsy | +++ |
| G392E | 13 | Progressive myoclonus epilepsy, chorea | ++++ |
| G392R | 8 | Epilepsy with slow wave sleep | ++++ |

symptoms were similar and were first noticed when she was 18. Behavioural problems, depression, and frontal dysfunction were noticed. Epilepsy was drug-resistant and she experienced, during the course of the disorder, two episodes of status epilepticus. She became mute and bedridden and died of inhalation pneumonia at the age of 33. In these patients, a heterozygous S52R missense mutation at position 273 in exon 2 was detected, the same as in the two families from the United States.

A few non-familial cases have been also described. Coutelier *et al.* (2008) reported an 11-year-old girl who had had normal development until 8 years of age, when she developed a rapidly-progressive symptomatology, including aggressive behaviour, intellectual decline, psychic seizures, and subtle seizures with eyelid myoclonus. The EEG was suggestive of epilepsy with continuous spike-and-waves during slow-wave sleep. This patient was the first person identified with the G392R mutation in neuroserpin that resulted in severe juvenile phenotype and had accordingly appeared *de novo*. Hagen *et al.* (2011) more recently reported an additional sporadic patient with progressive myoclonus epilepsy and declining mental status starting in adulthood. Generalized seizures occurred early with myoclonus and progressive gait disturbances. Neuroimaging revealed mild atrophy and multiple periventricular white matter lesions, consistent with demyelination. The course was one of progressive decline with death occurring at the age of 34. Genetic analysis revealed a nucleotide substitution, resulting in a proline to leucine amino acid substitution (L47P). The genotype-phenotype correlations are remarkably strong, with mutations causing increasingly severe clinical features in the following order: S49P, S52R, L47P, H338R, G392E and G392R (*table 1*). In general, increasing clinical severity is characterized by earlier onset of the symptom and an increasing contribution

from the epileptic component of the syndrome. More specifically, individuals with the S49P mutation have diffuse small intraneuronal inclusions of neuroserpin with an onset of dementia between the ages of 45 and 60 (Davis *et al.*, 1999a; Davis *et al.*, 1999b; Bradshaw *et al.*, 2001). People with the S52R, L47P and H338R variants have larger and more numerous intraneuronal inclusions associated with progressively earlier onset of symptoms, during early adulthood (S52R, L47P) and adolescence (H338R) (Hagen *et al.*, 2011; Miranda *et al.*, 2008). In the most severe cases, caused by the most polymerogenic mutations, namely G392R and G392E, the patients exhibit the earliest onset of symptoms, with profound intellectual decline during childhood associated with severe, uncontrolled epilepsy (Coutelier *et al.*, 2008). While FENIB is a rare disease, with only a few known kindreds worldwide, diagnosis should be considered in patients that present with a frontal-type dementia combined with epilepsy, particularly when eosinophilic inclusions are seen on brain biopsy or at post-mortem.

The pathological polymerisation of neuroserpin variants results in FENIB

These remarkable genotype-phenotype correlations that are evident in the clinic are mirrored by the biophysical and biochemical properties of the variant NS proteins. Under physiological conditions, the rate of aggregation of the least clinically-aggressive NS mutant, S49P, is more than 10 times higher than that of the wild type protein, whereas the association rate constant for tPA is essentially unchanged (Belorgey *et al.*, 2002). The next most severe mutation, S52R, results in a further tenfold increase in the polymerization rate and the loss of effective tPA inhibition (Belorgey *et al.*, 2004). Thereafter, the aggregation rate becomes so

high as to be practically immeasurable. In the classic model of serpin polymerisation, proposed by Carrell and Lomas (2002), the functional changes in NS are all caused by the progressive destabilization of the key structural element, termed β -sheet A, which forms the core of NS (*figure 1*). This structural perturbation favours the incorporation of the RCL from a second NS molecule over the physiological process of intramolecular RCL insertion that occurs during protease inhibition (Davis *et al.*, 2002). An initial loop-sheet polymerization event yields a NS dimer that retains a patent β -sheet A at one end and a destabilized RCL at the other. Such a dimer is competent to undergo further loop-sheet polymerization to form trimers and eventually higher-order polymers. Recently, Huntington and colleagues have proposed alternative mechanisms for polymerization that require a more significant domain swap between the serpins in a chain (Sado *et al.*, 2009; Huntington & Whisstock, 2010; Belorgey *et al.*, 2011; Singh & Jairajpuri, 2011; Yamasaki *et al.*, 2011). In either case, the NS polymers gradually become entangled in the neuronal endoplasmic reticulum (ER) and form inclusions, known as Collins bodies. This phenomenon of aggregation or 'polymerization' has been described in other serpins such as α_1 -antitrypsin where it results in hepatocyte inclusions and liver disease (Lomas *et al.*, 1992; Elliott *et al.*, 1996; Huntington *et al.*, 1999). The observation that FENIB was caused by NS mutations (S49P and H338R) that are structurally homologous to substitutions in α_1 -antitrypsin, which also lead to polymerization (S53F and H334D, respectively) (Lomas *et al.*, 1993), argues strongly in favour of a shared molecular mechanism. This was confirmed by the finding that the neuronal inclusion bodies of FENIB were formed by polymers of NS with identical morphology to the polymers of mutant α_1 -antitrypsin present in hepatocytes from a child with α_1 -antitrypsin deficiency-related cirrhosis (Davis *et al.*, 1999a; Carrell & Lomas, 2002).

Dissecting the pathological mechanisms of the dementing and epileptic components of FENIB

The ER-resident inclusions (Miranda *et al.*, 2004) in the neurones of individuals expressing NS variants are likely to represent the toxic gain-of-function that results in this dominantly-inherited neurodegenerative syndrome. Serpin polymerisation exerts a stress on the cell that differs in important ways from other proteinopathies. In most cases, misfolding of proteins in the ER results in activation of the unfolded protein response (UPR). However, the native-like structure of serpin polymers results in a distinct signalling pathway called the ER overload response (Davies *et al.*, 2009).

This ER overload response activates NF- κ B by a pathway that is dependent on raised cytoplasmic Ca^{2+} levels. ER-associated degradation or ERAD is involved in the degradation of mutant NS and may be able to degrade the polymers (Ying *et al.*, 2011), whereas autophagy is more important in the bulk turnover of both wild type and mutant NS (Kroeger *et al.*, 2009). In the presence of polymerogenic mutations in serpins, and with increasing age, these mechanisms are overwhelmed and retention of polymers in the ER leads to cell death. This is apparent in the fly model of FENIB where the accumulation of intracellular polymers is associated with locomotor deficits where there was a correlation between the severity of the behavioural deficits and the degree of polymer accumulation (Miranda *et al.*, 2008).

While the intraneuronal accumulation of NS inclusions may underpin the dementia seen in FENIB, there is evidence that other mechanisms may also contribute to the epileptic propensity of these patients. In particular, it is notable that epilepsy is rarely seen when normal levels of tPA inhibitory activity are present, as is the case for wild type and S49P NS. This has been recently highlighted in relation to stroke, where patients thrombolized with recombinant tPA present a higher level of seizures (Alvarez *et al.*, 2013). When tPA inhibition is lost (for example, for the S52R and 392 mutants) then epilepsy becomes the major clinical feature (Davis *et al.*, 2002). Notably, neuroserpin is able to dampen neuronal sensitivity to excitotoxic stimuli by regulating tPA activity (Yepes *et al.*, 2000; Yepes *et al.*, 2002; Wu *et al.*, 2010). This effect appears to be mediated by the tPA-mediated proteolytic cleavage of the NR1 subunit of the NMDA receptor that increases the functioning of this excitatory glutamate receptor (*figure 2*) (Nicole *et al.*, 2001; Fernandez-Monreal *et al.*, 2004; Samson *et al.*, 2008; Baron *et al.*, 2010).

Excess glutamatergic neurotransmission is also a potent cause of epilepsy (reviewed in Vincent & Mulle [2009]) and increased NMDA signalling in response to dysregulated tPA activity could contribute to FENIB. Indeed, co-injection of neuroserpin with kainate into the hippocampus of the mouse attenuated the spread of consequent epileptic activity when compared to co-injection of kainate with vehicle alone (Yepes *et al.*, 2002). These data suggest a role for tPA in seizure spread in epilepsy and support the use of NS or others tPA inhibitors as a potential therapy.

Conclusion

Familial encephalopathy with neuroserpin inclusion bodies is a recently described neurodegenerative disease that is responsible for progressive myoclonic epilepsy and/or pre-senile dementia. Serpinopathies

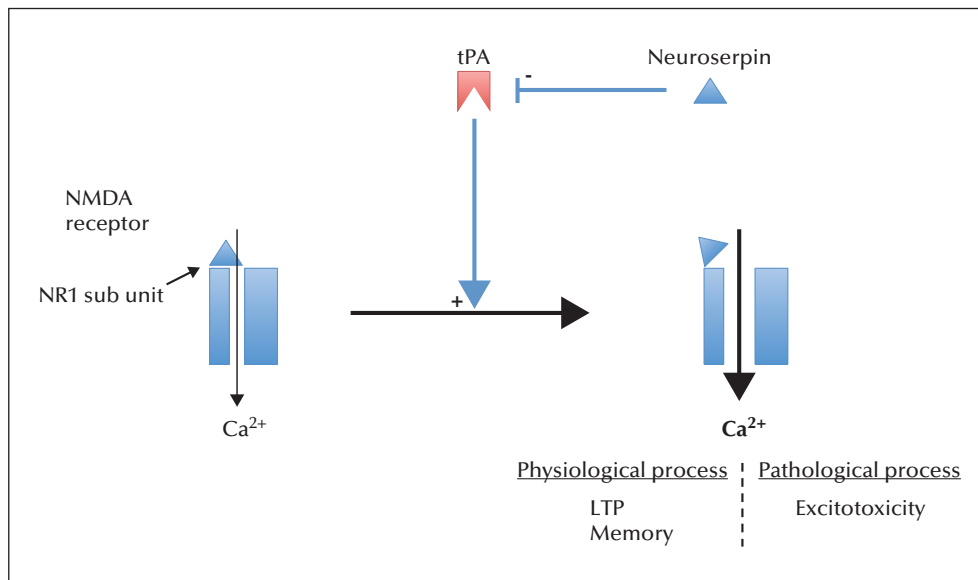


Figure 2. The NMDA receptor mediates the influx of calcium into neurones in the presence of glutamate. tPA has been shown to cleave the NR1 subunit (triangle) of the receptor and, as a consequence, increases the concentration of intracellular calcium. By inhibiting tPA, neuroserpin is thought to protect neurons from the toxic consequences of excessive NMDA activity, termed 'excitotoxicity'.

are unique among conformational diseases because they form native-like aggregates or polymers in the ER of synthetic cells. Serpin variants accumulate as inclusion bodies and thus activate the ER-overload response. It appears that this gain-of-function toxicity is responsible for the neuronal dysfunction and death that underpins dementia in FENIB. The evidence is less clear as to whether similar gain-of-toxicity is the cause of the epilepsy in FENIB or whether this results from dysfunction of the NS/tPA system. The relative importance of these two mechanisms has yet to be clearly elucidated. To date, genetic analysis of the *SERPINI1* gene should be performed in patients with adult-onset PME and early-onset, rapidly progressive cognitive dysfunction or predominantly frontal dementia. □

Disclosures.

None of the authors have any conflict of interest to disclose.

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GOSR2: a progressive myoclonus epilepsy gene

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ABSTRACT – *GOSR2*-associated PME is associated with a homozygous mutation in *GOSR2* (c.430G>T, p.Gly144Trp), a Golgi vesicle transport gene. The functional effect of this mutation is a loss of function that results in failure of the *GOSR2* protein to localize to the cis-Golgi. The main clinical features of the *GOSR2*-associated PME are early-onset ataxia, areflexia, action myoclonus and seizures, scoliosis, elevated creatine kinase levels, relative preservation of cognitive function until the late stages of the disease, and relentless disease course. Severe photosensitive myoclonus is a common feature. *GOSR2*-associated PME is a rare disease with very few cases reported so far and it can be expected that the identification of further patients will contribute to expanding the phenotype and genotype of this condition.

Key words: Progressive myoclonus epilepsy, *GOSR2*, myoclonus, photosensitivity, scoliosis, ataxia

In 2011, Corbett *et al.* reported six patients with a progressive myoclonus epilepsy (PME) (Minassian *et al.*, 2016) phenotype whose main cardinal features were onset of ataxia in the first years of life, appearance of action myoclonus and seizures later in childhood, and loss of independent ambulation in the second decade. Cognition was not typically affected, although mild memory difficulties occurred for some in the third decade. This condition was found to be associated with a homozygous mutation in *GOSR2* (c.430G>T, p.Gly144Trp), a Golgi vesicle transport gene. This p.Gly144Trp mutation gives rise to a loss of function and results in

failure of the *GOSR2* protein to localize to the cis-Golgi. Following this, the clinical and neurophysiological characteristics of PME associated with *GOSR2* mutation were further detailed in 12 patients (including the original six patients described by Corbett *et al.* [2011]); all patients had the same homozygous mutation (c.430G>T, p.Gly144Trp) (Boissé Lomax *et al.*, 2013). Interestingly, the birthplaces of all these patients (including the birthplaces of the ancestors of one Australian patient) clustered around the North Sea (hence the eponym for this type of PME of ‘North Sea progressive myoclonus epilepsy’ by Boissé Lomax *et al.* [2013]). This geographic distribution suggests that the

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identified *GOSR2* mutation may have spread along the North Sea at the time of the Viking conquests in the viiith century, although the dating of the time of mutation indicates that the mutation occurred much earlier than the Viking conquests.

Gene identification

The *GOSR2* gene was identified as a causative gene for PME through the genetic and molecular analyses of a family with one affected child. The proband was an Australian female, born to second-cousin parents of British origin. She, her unaffected brother, and her parents were genotyped using Affymetrix 250K Nsp SNP chips for genome-wide linkage mapping. Homozygosity mapping was performed and a single suggestive linkage region on chromosome 17 was identified, with a maximum possible LOD score of 1.93. Sequence capture followed by next-generation sequencing was carried out and a homozygous variant in *GOSR2* (MIM: 604027), c.430G>T, p.Gly144Trp, (NM_004287.3), was identified as the possible causative mutation. Based on a sequence analysis of a cohort of 73 unrelated individuals with molecularly unsolved PME, five additional individuals (from four families) were identified who were homozygous for the same *GOSR2* variant, c.430G>T, p.Gly144Trp (Corbett *et al.*, 2011). No consanguinity was reported in any of the families. The c.430G>T *GOSR2* variant was not found in 584 chromosomes from unaffected individuals or in dbSNP132, leading to confirmation that this was the causative mutation. Of the four additional families, one was of German ancestry and three were Dutch. Further analysis of one affected individual from each family with both microsatellite markers and Illumina 610 quad SNP chips revealed a founder mutation that was most likely to be of European ancestry (Corbett *et al.*, 2011).

GOSR2 is a member of the Qb-SNARE family of vesicle docking proteins. There are three known alternatively spliced isoforms of *GOSR2* and all are predicted to be affected by the c.430G>T, p.Gly144Trp mutation. The mutated glycine 144 residue (G144) is within the Qb-SNARE domain of the *GOSR2* protein and shows high evolutionary conservation from mammals to yeast. Initial investigations into the pathogenic mechanisms due to the mutation have shown that the *GOSR2* p.Gly144Trp mutant protein fails to localize to the cis-Golgi (Corbett *et al.*, 2011).

Clinical features associated with *GOSR2* mutation

The distinct clinical features of PME associated with *GOSR2* mutations are early-onset ataxia (at around 2 years of age), onset of action myoclonus and seizures

at around 6 years of age, and scoliosis in adolescence and the absence of cognitive deterioration during evolution, although mild cognitive decline may be observed in the later stages. The clinical history of Case 1 in the original report by Corbett *et al.* (2011) is paradigmatic in terms of illustrating the characteristics and course of the disease. This patient began to have difficulty of walking and was found to be areflexic at the age of 2, but development was otherwise normal. At the age of 7, she had a tremor, and it became clear that action myoclonus and occasional absence seizures were present. At the age of 13, she began having drop attacks as well as major convulsive seizures. The patient required a wheelchair from the age of 14 due to falling attacks and was unable to walk unaided from the age of 16. Notably, the patient had severe scoliosis which required surgical correction. By the age of 22, the patient was confined to bed and she died aged 32 as the result of complications associated with uncontrolled myoclonus. The patient's intellect was preserved until the latter few years of her life, when there was mild cognitive impairment. Autopsy showed mild cerebral atrophy and a lack of gross structural abnormalities. Histological examination revealed subtle, Alzheimer type II gliosis in the basal ganglia region (consistent with metabolic derangement related to her agonal state) and minor loss of Purkinje cells and gliosis in the cerebellar vermis.

The clinical features and evolution of PME caused by mutation of *GOSR2* were detailed in the study by Boissé Lomax *et al.* (2013) who described the original 6 patients with the homozygous *GOSR2* c.430G>T, Gly144Trp mutation (Corbett *et al.*, 2011), as well as six new patients (from 11 families) who were molecularly identified with the same mutation. The clinical presentation in the 12 patients was remarkably similar, with features of early-onset ataxia (on average at 2 years of age), followed by myoclonic seizures at the average age of 6.5 years. During the course of the disease, all patients exhibited multiple seizure types, including generalized tonic-clonic seizures, absence seizures, and drop attacks. The patients also uniformly displayed highly photosensitive generalized myoclonus that worsened with action or emotional stress, but was minimal at rest and almost completely absent during relaxation. In some patients, myoclonus and drop attacks were made worse by fever. One patient presented with periods of 'status myoclonicus', characterized by continuous myoclonus lasting for hours up to a day at a time. All patients developed scoliosis by the time they reached adolescence, making this an important diagnostic feature. Additional skeletal deformities were present, including *pes cavus* in 4 patients and syndactyly in two patients. Notably, cognition was preserved in the context of severe motor disability until the later stages of the disease. The progression of the

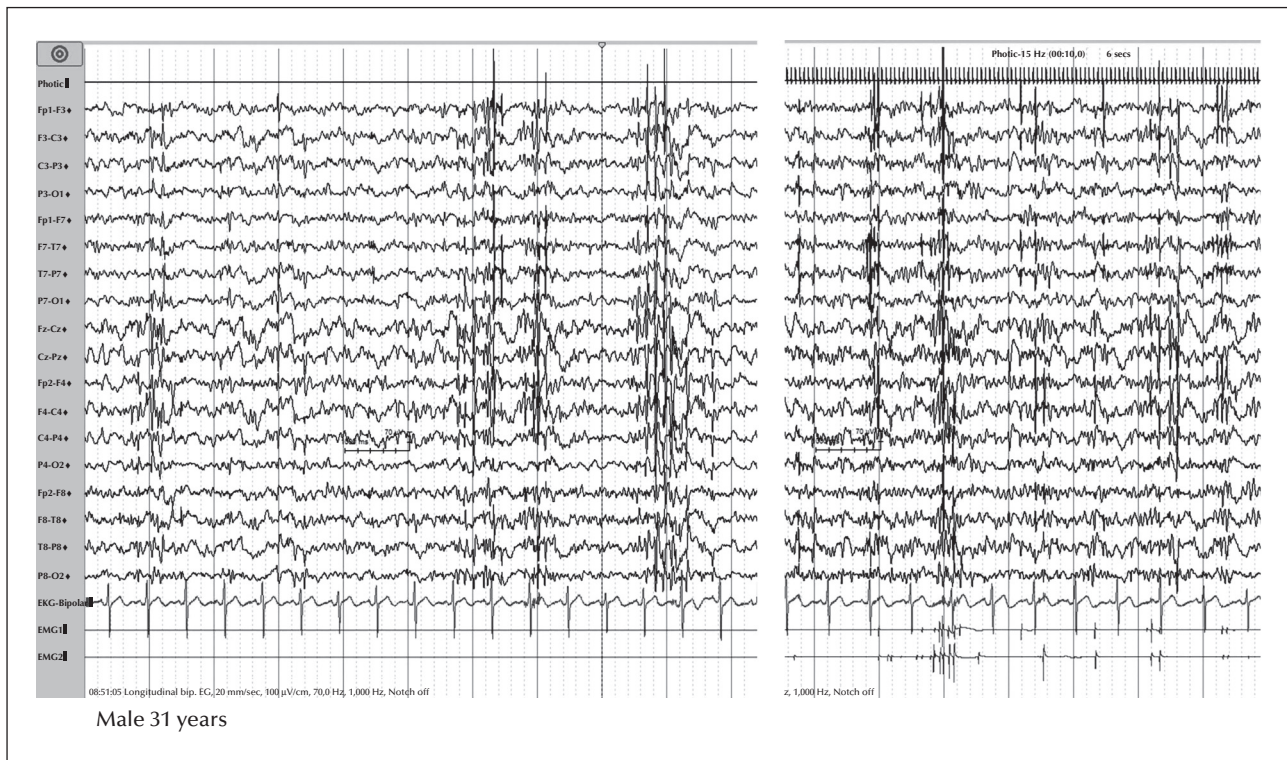


Figure 1. Polygraphic recording in a 31-year-old male with GOSR2-associated PME. Left panel: the patient is at rest. The tracing shows slowing of background activity, bursts of generalized spike-and-wave discharges, and multifocal spikes in both hemispheres. Myoclonic potentials are not evident in the EMG leads. Right panel: photic stimulation at 15 Hz elicits generalized polyspikes and spike-and-wave discharges, often associated with myoclonic potentials at both EMG leads. EMG1: right deltoid; EMG2: left deltoid.

disease showed a relentless decline; patients became wheelchair-bound (at a mean age of 13 years) and four died during their third or fourth decade of life.

An additional patient with PME caused by mutation in *GOSR2* has been reported by Praschberger *et al.* (2015). This patient was a 61-year-old female presenting with a PME phenotype and was found to be compound heterozygous for two *GOSR2* mutations; the known c.430G>T, Gly144Trp mutation and a novel c.491_493delAGA (p.Lys164del) mutation. She presented with mild ataxia at the age of 2 years, as well as transient episodes of motor deterioration triggered by infection and fever. At the age of 14 years, she started to suffer from action myoclonus and seizures. In her thirties, she was wheelchair-bound. Scoliosis and areflexia were also noted. No cognitive deterioration was observed throughout the course of the disease, although mild cognitive decline was detected after repeated neuropsychological testing. The most relevant distinctive clinical feature of this patient was the milder course of the disease, at variance with the previously reported patients with PME associated with the homozygous *GOSR2* Gly144Trp mutation. Finally, van Egmond *et al.* (2014) reported the same homozygous c.430G>T, p.Gly144Trp *GOSR2* mutation

in five Dutch patients with progressive myoclonus ataxia (PMA) (also known as Ramsay Hunt syndrome). These patients showed clinical features that were very similar to those previously described for *GOSR2* mutation-positive PME patients. However, differences include the fact that in the five patients with PMA, cognitive function was not seen to decline (however, only one patient was in his third decade while the others were younger) and scoliosis was observed in only three of the five subjects.

Neurophysiological investigations

EEG analysis of *GOSR2* mutation-positive PME patients reveals generalized spike-and-slow-wave discharges with a posterior predominance, often with a slow background. The generalized discharges are highly photosensitive. Focal or multifocal discharges can also be observed (Boissé Lomax *et al.*, 2013) (figure 1). Nerve conduction studies have been reported to be consistent with a mild, predominantly axonal peripheral neuropathy, while electromyography was normal in all patients with the exception of one in the series reported by Boissé Lomax *et al.* (2013). Signs of

sensory neuronopathy, anterior horn cell involvement, or both, were detected in all patients with absent reflexes reported by van Egmond *et al.* (2014).

Multimodal evoked potentials were shown to be unremarkable (Boissé Lomax *et al.*, 2013).

Brain magnetic resonance imaging studies have displayed essentially normal findings or generalized cerebral and cerebellar atrophy (Boissé Lomax *et al.*, 2013; Prachschberger *et al.*, 2015). Elevation of serum creatine kinase levels (median: 734 IU), in the context of normal muscle biopsies, was reported for all patients in the study of Boissé Lomax *et al.* (2013), but was not a uniform feature in the series described by van Egmond *et al.* (2014).

Conclusions

GOSR2-associated PME has a relatively homogeneous clinical presentation, characterized by a pattern of early-onset ataxia, areflexia, action myoclonus and seizures, scoliosis, elevated creatine kinase levels, relative preservation of cognitive function until the late stages of the disease, and relentless disease course. Thus far, the same homozygous c.430G>T (p.Gly144Trp) mutation has been detected in all reported patients with GOSR2-mediated PME, with the exception of one patient with a milder disease course who was heterozygous for the known c.430G>T (p.Gly144Trp) mutation and a novel c.491_493delAGA (p.Lys164del) GOSR2 mutation. The identification of

additional patients will contribute to further expanding the phenotype and genotype and will add to our knowledge of GOSR2-related disease. □

Disclosures.

None of the authors have any conflict of interest to disclose.

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KCTD7-related progressive myoclonus epilepsy

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ABSTRACT – Progressive myoclonic epilepsy associated with *KCTD7* mutations has been reported in 19 patients from 12 families. Patients show homozygous mutations in the coding regions of the *KCTD7* gene. The disease starts in infancy. Patients typically show an initial severe epileptic disorder, with abundant epileptiform discharges on EEG and myoclonic seizures in the foreground, associated with cognitive regression and ataxia. Continuous multifocal myoclonus aggravated by action is observed in more than half of the cases. After a few years, the disease tends to stabilize and long survival can be expected. Some patients remain able to walk independently. The severity of the disease is variable from one patient to another, even within the same family. It is hypothesized that the epileptic disorder may influence the neurological regression observed in patients.

Key words: KCTD7, progressive myoclonus epilepsies, infancy, encephalopathy

In 2007, three siblings from a consanguineous family with a clinical picture of progressive myoclonic epilepsy (PME) (Minassian *et al.*, 2016) were reported, associated with a homozygous mutation of the gene encoding potassium channel tetramerization domain-containing protein 7 (*KCTD7*) (Van Bogaert *et al.*, 2007). Since this original family, 16 other patients from 11 new families have been reported (Blumkin *et al.*, 2012; Kousi *et al.*, 2012; Krabichler *et al.*, 2012; Staropoli *et al.*, 2012; Farhan *et al.*, 2014). The disease is inherited as an autosomal recessive trait and its incidence is unknown. In a study in which *KCTD7* was screened in a cohort of more than 100 unconfirmed PME

patients after exclusion of neuronal ceroid lipofuscinosis (NCL), five positive families were identified, suggesting that *KCTD7* is not an exceptional cause of PME (Kousi *et al.*, 2012). In a pilot study evaluating a panel of 265 genes, including *KCTD7*, in 33 index patients with various epileptic syndromes randomly selected in Germany and Switzerland, one patient was shown to have a homozygous mutation for *KCTD7* (Lemke *et al.*, 2012). The present article focuses on the clinical characteristics of *KCTD7*-related PME from the 19 published cases reported so far and discusses the pathophysiology and genotype/phenotype correlation of the disease.

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Clinical characteristics

The 12 families reported so far (Van Bogaert *et al.*, 2007; Blumkin *et al.*, 2012; Kousi *et al.*, 2012; Krabichler *et al.*, 2012; Staropoli *et al.*, 2012; Farhan *et al.*, 2014) had variable ethnic origin and showed a high rate of consanguinity (5/12). Epileptic seizures were the first signs in all cases. Onset ranged between five months and three years, after a period of psychomotor development that was described as either normal or slightly delayed. Independent walking was usually acquired. However, one reported patient with onset in the second year of life never walked (Kousi *et al.*, 2012). The first seizures were described as either myoclonic or generalized tonic-clonic seizures, and were precipitated by fever in some cases. Myoclonic seizures were reported in the course of the disease in all but one patient. In 11 patients, continuous multifocal myoclonus, aggravated by action and posture, was reported either at presentation or during the course of the disease. In one patient, myoclonus was associated with opsoclonus, which prompted the authors to consider a diagnosis of opsoclonus-myoclonus syndrome (Blumkin *et al.*, 2012). Other types of seizures (atonic seizures and atypical absences) were also reported. Epilepsy responded poorly to antiepileptic drugs in 2/3 of patients, the most effective drugs being levetiracetam, valproate and clobazam. All patients had cognitive decline leading to severe dementia. Progressive ataxia was reported in all but one patient, and two thirds of them became wheelchair-bound. Pyramidal signs were described in eight patients and microcephaly in six patients.

Concerning complementary examinations, EEGs were described to be abnormal, except for those very early during the course of the disease. The most common findings were very frequent multifocal and/or generalized spike-waves associated with an excess of slow activity. In a number of patients, EEG abnormalities were more prominent in the posterior areas, and intermittent light stimulation evoked generalized or posterior epileptiform discharges. In some patients, the EEG was reported as either hypsarrhythmic or showing continuous spike waves during slow-wave sleep. Action myoclonus was not associated with concomitant epileptiform discharges. One patient had negative focal myoclonus. *Figure 1* illustrates EEG findings from two affected siblings from the first family (Van Bogaert *et al.*, 2007).

Cerebral magnetic resonance imaging was either normal or showed non-specific findings (atrophy or posterior white matter hyperintensities). Metabolic work-up of blood and cerebrospinal fluid was not suggestive of mitochondrial dysfunction. Fundoscopy was normal in most patients.

The search for storage material was performed by skin biopsy in nine patients from eight different families and was normal except for one patient who was reported to have electron-dense storage material in fibroblasts and a lymphoblastoid cell line based on electron microscopy (Staropoli *et al.*, 2012). This finding is noteworthy as it was the only finding of storage material in a patient mutated for *KCTD7*, which prompted the authors to designate *KCTD7* mutations as the cause of a new subtype of NCL that was called 'NCL type 14'. It should be noted that the screening for *KCTD7* mutation performed in a cohort of 22 NCL patients with storage material demonstrated by electron microscopy analysis was negative, suggesting that *KCTD7* is not a common cause of NCL after exclusion of the already known NCL genes (Kousi *et al.*, 2012). Moreover, the patient reported by Staropoli *et al.* (2012), as well as his affected brother, had a very atypical clinical course that differed from other *KCTD7* patients. First, these 2 patients were the only reported cases with severe ocular involvement, *i.e.* optic atrophy leading to visual loss. Second, constant clinical deterioration was observed in these 2 patients who died in their mid-teens. This contrasted with the other reported *KCTD7* patients who showed clinical stabilization after a period of early regression, most of whom were still alive at the last assessment.

To our knowledge, the oldest patient (Patient 2 from the original report [Van Bogaert *et al.*, 2007]) is now aged 25 years and is still able to walk independently.

Differential diagnosis

The typical presentation follows a 3-stage evolution that is quite unique among PME:

- (1) a period of normal development;
- (2) a period of mental and motor regression with epileptic myoclonic seizures that starts at around 1-2 years of age;
- (3) and a period of stabilization after a few years.

Rare cases with optic atrophy and storage material on skin biopsy may complicate differential diagnosis with the classic late-infantile form of NCL. Clues to resolve this differential diagnosis are summarized in *table 1*. The rare presence of associated opsoclonus can evoke opsoclonus-myoclonus syndrome. Finally, in rare patients, ataxia or myoclonus may not be present, which raises the issue of whether such patients have an epileptic syndrome other than PME.

Physiopathology

All 11 different mutations identified so far are within the coding region of *KCTD7*, which is a member of

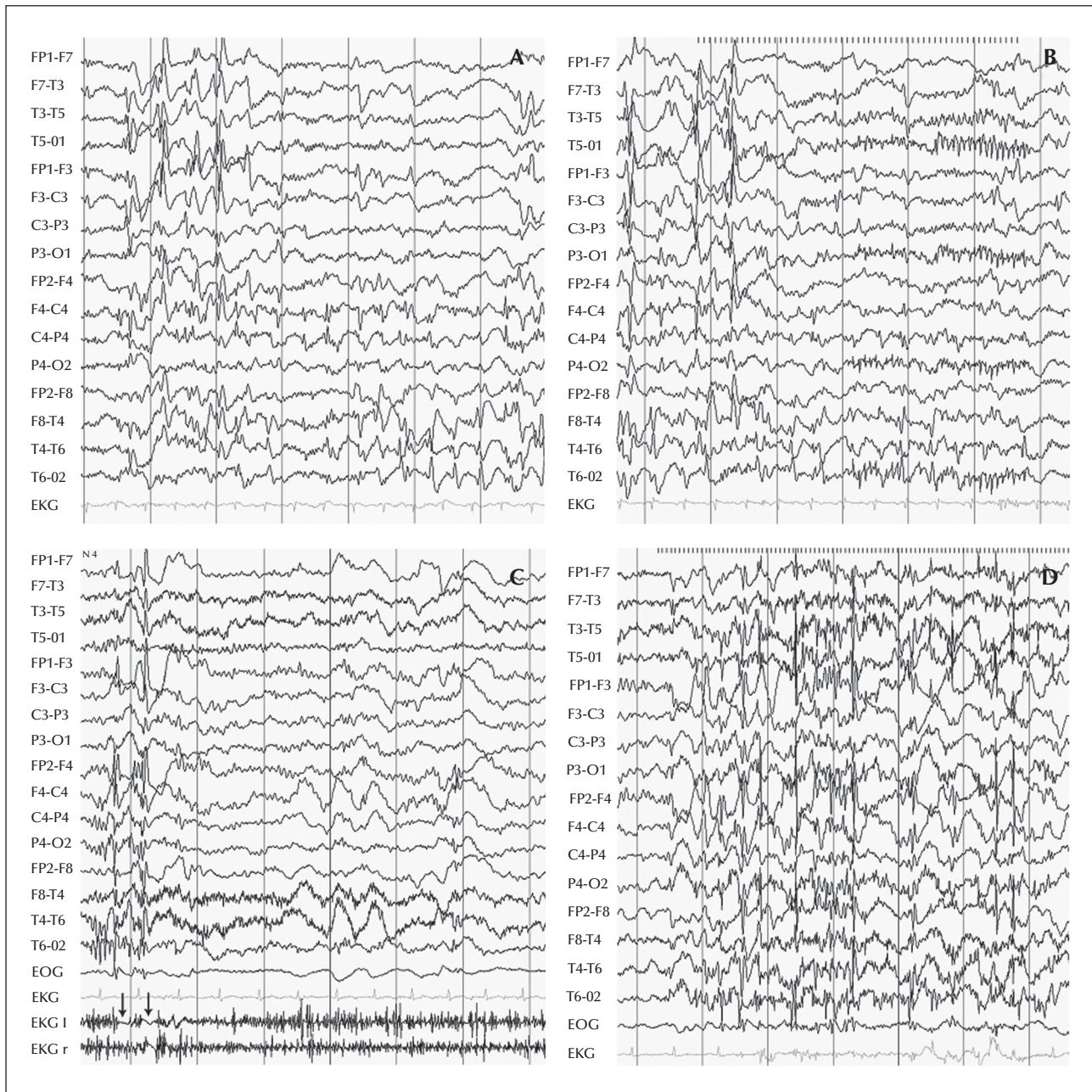


Figure 1. The EEG of Patients 1 and 2 from the original report by Van Bogaert *et al.* (2007). (A) EEG of Patient 1 at the age of 5 years, awake and at rest, showing slow dysrhythmia and multifocal high-amplitude epileptiform discharges. (B) Intermittent light stimulation (ILS) at 12 Hz inducing occipital spikes in addition to the spike-wave complexes still present at rest. (C) EEG of Patient 2 at the age of 14 years, awake with arms maintained in flexion and forearms in extension. On EMG, performed with surface electrodes placed over the left (l) and right (r) deltoid muscles, two negative myocloni appear as flattening of the EMG discharge (arrows), the first one on the left side and the second on both sides. Myoclonus was preceded by right-sided spike-wave discharges, followed by generalized spike-wave discharges. (D) ILS, at 15 Hz, induced generalized spike-waves, together with clinical bilateral myoclonus; these discharges were followed by rhythmic occipital spikes diffusing into a secondary generalized tonic-clonic seizure.

the *KCTD* gene family. This family of proteins shares an N-terminal BTB/POZ domain that demonstrates sequence homology to the T1 domain in voltage-gated potassium channels that allows its tetramerization (Stogios *et al.*, 2005). The *KCTD7* protein is extremely

conserved across species and, in mice, is expressed in the olfactory bulbs, the CA1 and CA3 hippocampal cells, and the Purkinje cells of the cerebellum (Azizieh *et al.*, 2011). *KCTD7* overexpression in transfected primary cultures of murine neurons resulted in

Table 1. Differential diagnosis between KCTD7-related PME and late-infantile NCL.

| | KCTD7-related PME | Late-infantile NCL |
|-------------------------|--|--|
| Gene defect | <i>KCTD7</i> | <i>CLN2</i> gene (<i>TPP1</i>) |
| Age at onset | 0.5-3 years | 2-3 years |
| Epileptic seizures | +++ (myoclonic, GTCS, atypical absences) | +++ (myoclonic, GTCS, atypical absences) |
| Action myoclonus | ++ | ++ |
| Progressive ataxia | +++ | +++ |
| Cognitive regression | +++ | +++ |
| Optic atrophy | - (*) | +++ |
| Retinitis | - | +++ |
| Atrophy on MRI | + | ++ |
| Photosensitivity on EEG | ++ | +++ (low frequency) |
| Storage material on EM | - (*) | +++ (curvilinear bodies) |
| Course | Stabilization (*) | Bedridden at 5-7 years |

+++seen in nearly all patients; ++ seen in >50%; + seen in <50%; - absent; TPP1: tripeptidyl peptidase 1; GTCS: generalized tonic-clonic seizures; EM: electron microscopy; *except in the family reported by Starapoli *et al.* (2012).

hyperpolarization of the resting membrane potential, and decreased their excitability in patch clamp experiments (Azizieh *et al.*, 2011). This is consistent with an epileptogenic effect of a KCTD7 defect. KCTD7 is much smaller than a typical potassium channel subunit, and its computed hydrophobicity profile does not indicate a transmembrane segment. It is hence extremely unlikely that KCTD7 would function as a transmembrane channel for potassium. The demonstration of a direct molecular interaction between KCTD7 and Cullin 3 suggests that the effect on the plasma membrane resting potential is likely to be mediated by Cullin 3 (Azizieh *et al.*, 2011). Although additional experiments are required to understand how the KCTD7 defect causes PME, this may already explain the lack of genotype/phenotype correlation observed in affected patients. Indeed, mutations within the functional BTB/POZ domain (5/11 reported mutations) did not result in a more severe phenotype.

An interesting finding is that the severity of the disease may be highly variable among affected members within the same family (Van Bogaert *et al.*, 2007; Farhan *et al.*, 2014). Since the more severely affected members also had a more severe epileptic disorder, this suggests that the epileptic activity itself played a role in the neurological deterioration observed in patients. From this point of view, KCTD7-related epilepsy could be considered as an epileptic encephalopathy, *i.e.* a condition in which epileptic activity may contribute to progressive neurological decline (Berg *et al.*, 2010),

such that effective antiepileptic intervention might improve developmental outcome. □

Disclosures.

The author has no conflict of interest to disclose.

The present manuscript is part of a *Epileptic Disorders*/Mariani Foundation supplement on Progressive Myoclonus Epilepsies, downloadable as a whole by visiting www.epilepticdisorders.com.

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Autosomal recessive progressive myoclonus epilepsy due to impaired ceramide synthesis

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ABSTRACT – Autosomal recessive progressive myoclonus epilepsy due to impaired ceramide synthesis is an extremely rare condition, so far reported in a single family of Algerian origin presenting an unusual, severe form of progressive myoclonus epilepsy characterized by myoclonus, generalized tonic-clonic seizures and moderate to severe cognitive impairment, with probable autosomal recessive inheritance. Disease onset was between 6 and 16 years of age. Genetic study allowed to identify a homozygous non-synonymous mutation in *CERS1*, the gene encoding ceramide synthase 1, a transmembrane protein of the endoplasmic reticulum (ER), catalyzes the biosynthesis of C18-ceramides. The mutation decreased C18-ceramide levels. In addition, downregulation of CerS1 in neuroblastoma cell line showed activation of ER stress response and induction of proapoptotic pathways. This observation demonstrates that impairment of ceramide biosynthesis underlies neurodegeneration in humans.

Key words: progressive myoclonus epilepsies, epilepsy, ceramide synthase, neurodegeneration, ER stress response

Despite great advances in molecular analytical techniques, aetiology remains undetermined in ~28 per cent of cases of progressive myoclonus epilepsies (PMEs) (Franceschetti *et al.*, 2014; Minassian *et al.*, 2016). We herein report clinical, neurophysiological, and genetic

findings of an Algerian family with a new form of PME with autosomal recessive inheritance, in which four out of six siblings presented a peculiar clinical picture, characterized by epilepsy, action myoclonus, and moderate-to-severe cognitive impairment.

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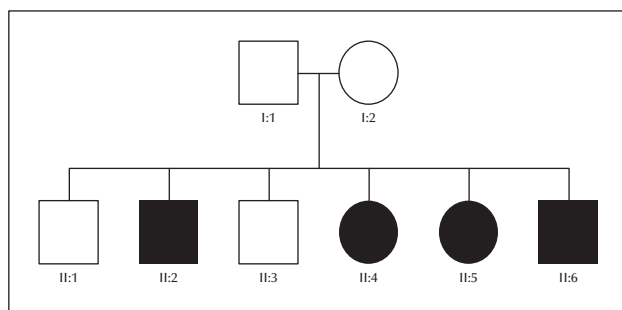


Figure 1. Family pedigree.

This family was first observed at the Hôpital Henri-Gastaut in Marseilles (France), in November 2007, by two of the authors (EF and PG), and was extensively reported by Ferlazzo *et al.* (2009). *Figure 1* shows the pedigree of the family with the four affected siblings (II:2, II:4, II:5, and II:6). The history and the clinical picture of this family appeared unusual and our first impression was that we were faced with a peculiar form of PME. In particular, we were intrigued by the presence of a severe myoclonic syndrome in most (II:2 was wheelchair-bound, and II:4 and II:5 walked with bilateral support) and the persistence of frequent generalized tonic-clonic seizures (GTCS) over the years, the dramatic reduction of action myoclonus following every GTCS, moderate-to-severe intellectual disability, and the unresponsiveness to common antimyoclonic agents (piracetam, levetiracetam and benzodiazepines).

Parents (I:1 and I:2) originated from two small but close Algerian villages and denied consanguinity. All affected siblings underwent neurological examination as well as neurophysiological evaluations, including video-EEG recordings at awakening. Sleep EEG was performed in Patients II:4 and II:6. To evaluate severity of action myoclonus, section 4 of the Unified Myoclonus Rating Scale (UMRS) was completed for all patients (normal score: 0). Serologic screening for coeliac disease was performed for Patients II:2, II:4, and II:6.

Patient II:2 underwent extensive studies including: EMG; a peripheral nerve conduction study; somatosensory, brainstem and visual evoked potentials (SSEP, BAEP, VEP); and brain MRI. He also underwent muscle biopsy to analyze the presence of abnormal mitochondria using routine methodology. This patient also underwent armpit skin biopsies in order to investigate Lafora bodies in sweat duct cells; ultrastructural studies of skin and rectal biopsy specimens were performed to search for typical neuropilofuscinosis inclusions. Biochemical assays for ceruloplasmin, hexosaminidases A and B, arylsulfatase, urine sialyloligosaccharides, leucocitary betagalactosidase, neuraminidase and betaglucoocerebrosidase, as

well as serum and CSF lactate and pyruvate levels, were performed. Molecular analysis for EPM1, EPM2A, EPM2B, MERRF, MELAS and *KCTD7*, were performed according to standard techniques.

Finally, homozygosity mapping was carried out to identify mutations in common using the Web-based tool, HomozygosityMapper. Based on the autosomal recessive pattern of inheritance, we carried out homozygosity mapping of the available family members using the Affymetrix Axiom Genome-Wide Human SNP Array. Genotypes of 567 k single nucleotide polymorphisms were analyzed by HomozygosityMapper software to identify runs of homozygous markers (ROHs) shared by the affected siblings.

Results

Patient II:2

This 29-year-old boy presented his first generalized tonic-clonic seizure (GTCS) at 7 years. Afterwards, GTCS presented at monthly intervals despite trials with different antiepileptic drugs. Onset of myoclonus probably occurred at the same time as the first seizure and worsened over the years; by the age of 20 years, the patient was wheelchair-bound. After each GTCS, myoclonus was reduced over 3-4 days. Lamotrigine worsened myoclonus, and levetiracetam and piracetam were ineffective. Clonazepam and phenobarbital produced a sedative effect. At consultation, the patient was being treated with VPA at 1,250 mg/day. Neurological examination revealed severe action myoclonus (UMRS score: 124). Neuropsychological tests revealed severe cognitive impairment. EEG showed background activity at 4-6 Hz with diffuse or multifocal spike- and polyspike-and-wave discharges (*figure 2*). Intermittent photic stimulation (IPS) was negative. EMG, ENG, BAEP and VEP were normal. SSEP showed giant potentials. Brain MRI showed mild atrophy of the cerebellum and brainstem. Abdominal ultrasound was normal. Fundus oculi and campimetry were normal. Muscle biopsy showed normal morphology and histochemistry without ragged red fibres. No Lafora bodies were observed in skin biopsies. No typical lipofuscin inclusions were present in skin or rectal biopsies. Biochemical assays were normal. Mutation analyses of the cystatin B gene, Lafora genes (laforin and malin), mtDNA for MERRF, MELAS or other mitochondrial diseases, were negative.

Patient II:4

This 24-year-old girl had GTCS that started at the age of 8 years and thereafter occurred monthly. Onset of myoclonus was at around 10 years and worsened

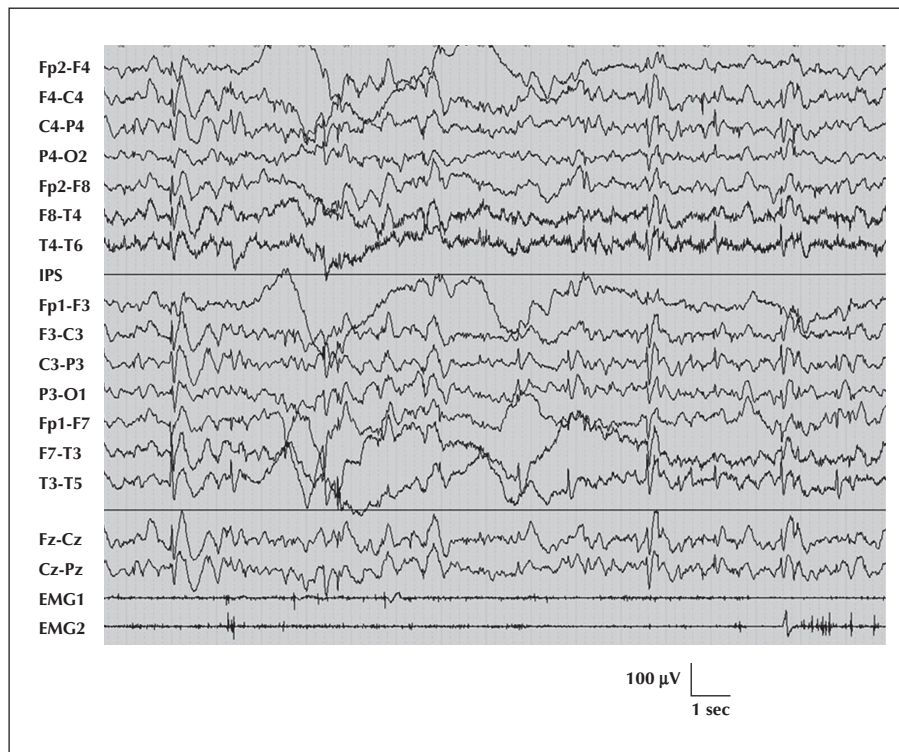


Figure 2. Patient II:2. EEG showing slow background activity in the theta-delta range and low- and mid-voltage spikes, spike- and polyspike-wave discharges, with fast spike components, both diffuse or multifocal, predominant over the centro-parieto-temporal regions. Note myoclonic jerks recorded over both deltoid muscles (EMG1: right deltoid; EMG2: left deltoid).

over the years. The girl started to attend a specialized institution for disabled persons by the age of 13 years. At consultation, she was being treated with VPA at 1,000 mg/day, which was the only drug she was ever given. Neurological examination revealed severe action myoclonus (UMRS score: 106), and she was unable to walk without support. Neuropsychological evaluation showed severe cognitive impairment. Following GTCS, a marked reduction in myoclonus severity was observed over 3 to 4 days. EEG showed slow background activity mixed with low-voltage fast activity, along with diffuse, irregular, spike- and polyspike-and-wave discharges. IPS from 10 to 20 Hz provoked a photoparoxysmal response. Sleep EEG showed spikes and polyspikes over the vertex and the centro-parietal regions during the first and second stages of non-REM sleep (*figures 3-5*).

Patient II:5

Myoclonus began at the age of 6 years. GTCS occurred at the age of 7 years and persisted weekly/monthly. At consultation, she was being treated with VPA at 1,250 mg/day. Neurological examination revealed action and stimulus-sensitive myoclonus (UMRS score: 99). A reduction in myoclonus severity was observed

after each GTCS, which persisted for 3-4 days. Neuropsychological evaluation showed severe cognitive impairment. EEG showed slow background activity and multifocal fast spikes, polyspikes and polyspike-and-wave discharges (*figures 6, 7*). IPS was ineffective.

Patient II:6

This 19-year-old boy presented two GTCS at the age of 16 years (only a few months before consultation) and had never had specialized medical attention. He was described to be completely normal by his father. A very mild action myoclonus was observed according to neurological evaluation (UMRS score: 14). He attended normal school with low performance until age 13, after which he was moved to a specialized school. Neuropsychological evaluation showed moderate cognitive impairment, with visual perception impairment and reduction in spontaneous language. EEG showed slow background activity, as well as diffuse spike- and polyspike-and-wave discharges and multifocal spikes activated by non-REM sleep (*figure 8*). IPS between 6 and 20 Hz provoked a photoparoxysmal response. He was untreated at referral and was prescribed VPA up to 1,000 mg/day.

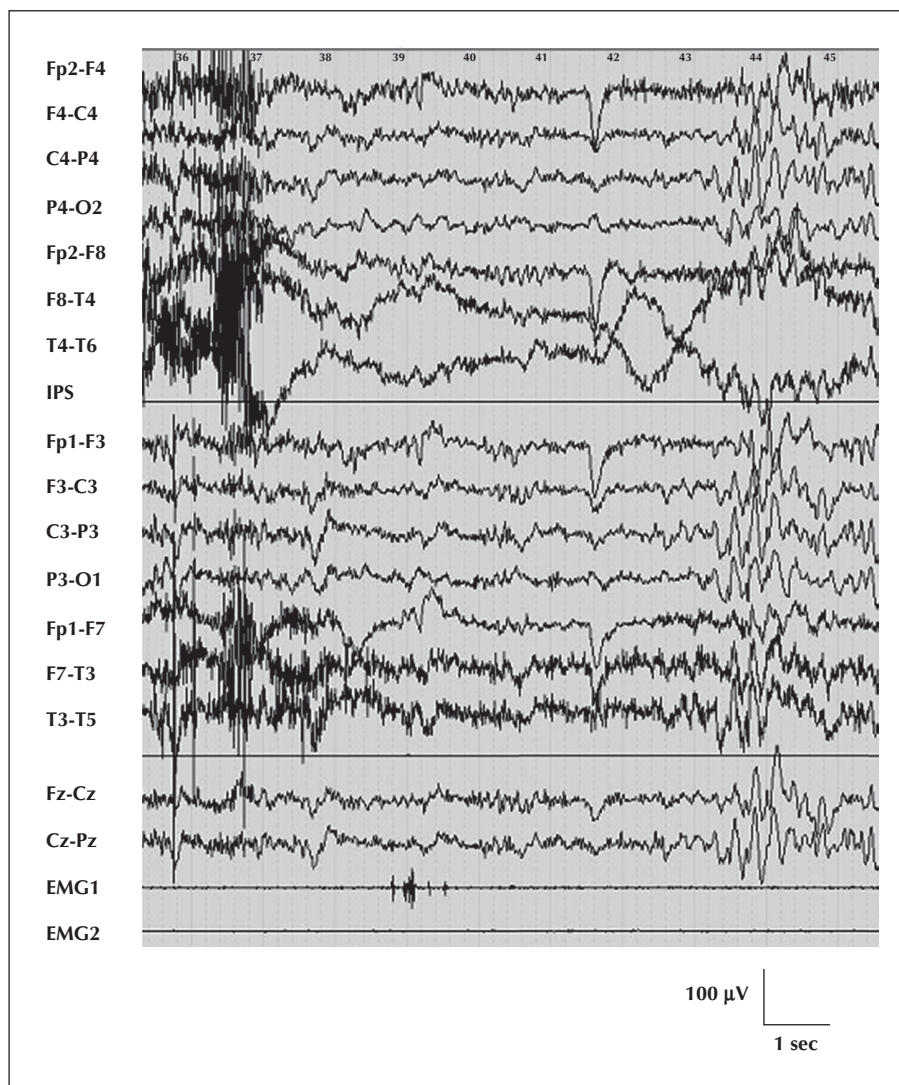


Figure 3. Patient II:4. EEG showing slow background activity at 6 Hz mixed with low-voltage fast activity, and diffuse, irregular, spike-and-polyspike-and-wave discharges at 3-4 Hz. Note myoclonic jerks recorded over the right deltoid (EMG1: right deltoid; EMG2: left deltoid).

Homozygosity mapping

Homozygosity mapping revealed a unique unreported non-synonymous coding variant of the *CERS1* gene (c.549 C>G, NM_021267.3; p.H183Q, NP_067090) (Vanni *et al.*, 2014). The mutation affects a critical histidine conserved in all human ceramide synthases and throughout evolution, which proved to be essential for enzymatic activity. Sanger sequencing confirmed the presence of the homozygous mutation in all affected individuals, whereas both parents and two unaffected siblings were heterozygous. By transfecting wild-type H183-tagged and mutant Q183-CerS1 V5-tagged vectors into HeLa cells, we found that the mutant and wild-type proteins were similarly expressed and that the mutant was correctly localized to the ER compartment (Vanni *et al.*, 2014). However, this mutation

was shown to severely impair CerS1 activity leading to down-regulation of CerS1 in a neuroblastoma cell line, triggering endoplasmic reticulum (ER) stress response and inducing proapoptotic pathways (Vanni *et al.*, 2014).

Discussion

We herein describe clinical, neurophysiological and genetic features of a family with a novel neurodegenerative disorder featuring PME. The main characteristics distinguishing our family from Unverricht-Lundborg Disease (ULD), the most common form of PME, are the presence of relevant cognitive impairment and seizure persistency even in the late phase of the disease. However, EEG findings partially overlap with those of ULD

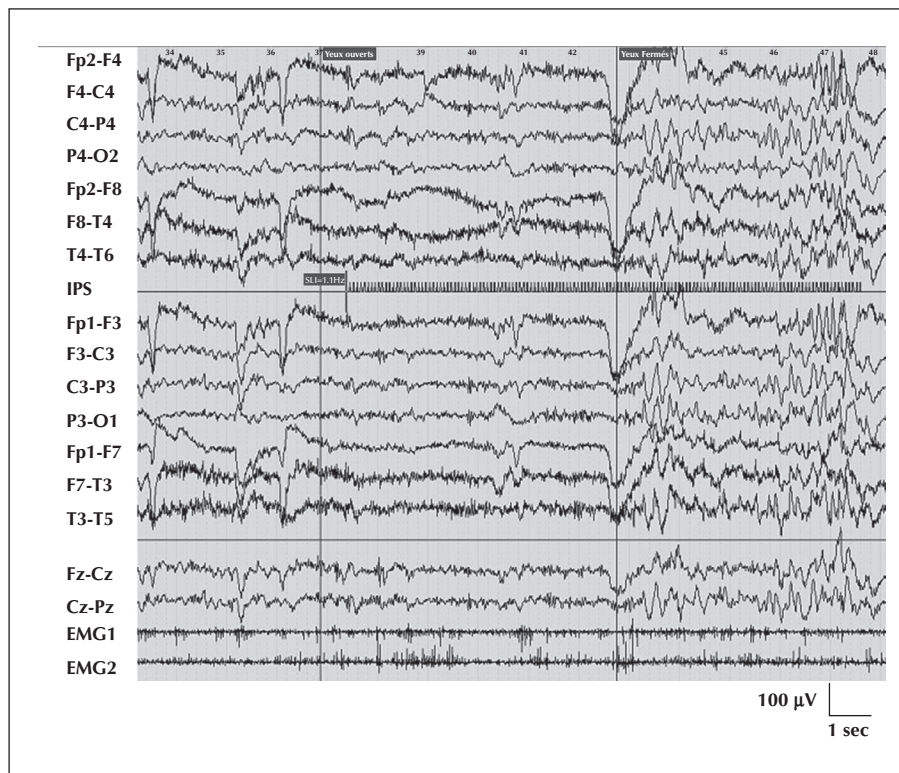


Figure 4. Patient II:4. The same EEG recording as in Fig. 3. Diffuse, irregular spike- and polyspike-and-wave discharges provoked by IPS at 14 Hz, associated with eye closure.

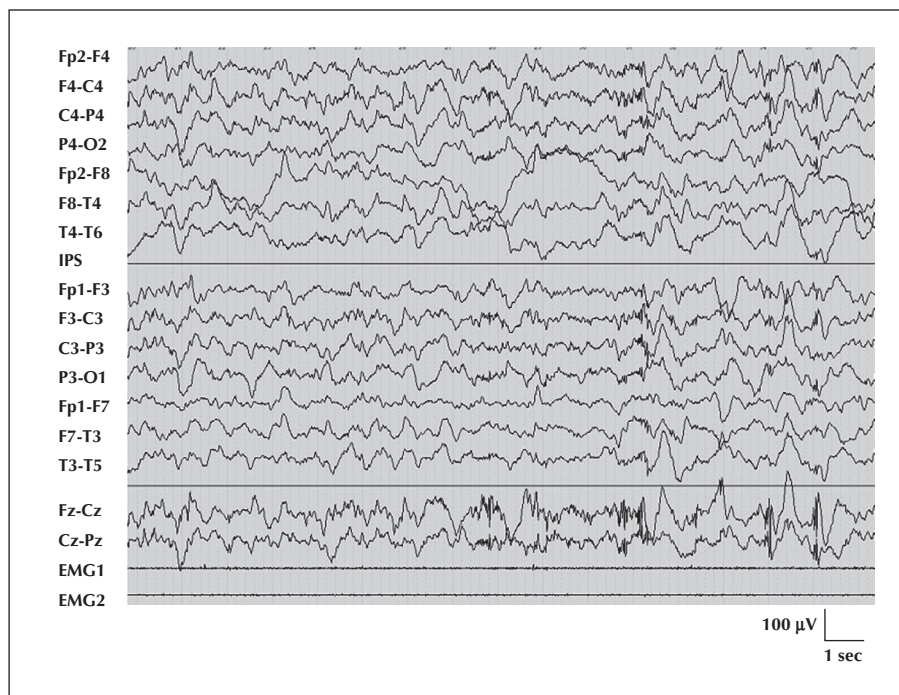


Figure 5. Patient II:4. During stages N1 and N2 of sleep, low- and mid-voltage spikes, polyspikes and polyspike-wave discharges, with fast spike components, were recorded over the vertex and both centro-parietal regions.

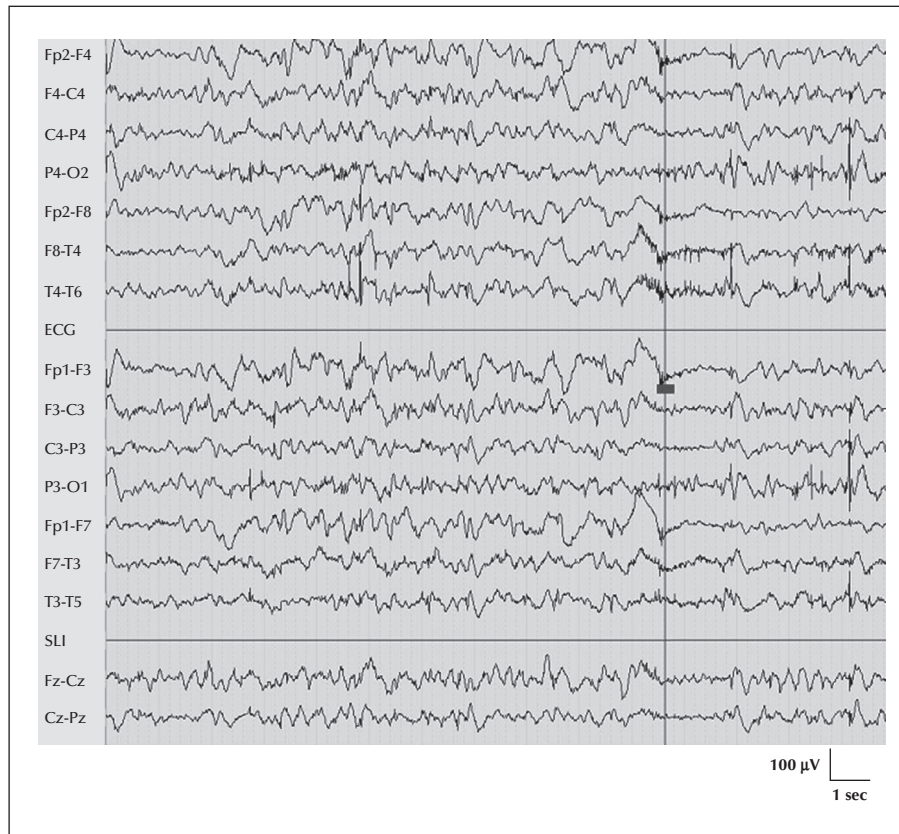


Figure 6. Patient II:5. EEG showing slow background activity in the theta-delta range mixed with low-voltage fast activity, along with multifocal fast spikes, polyspikes and polyspike-and-wave discharges.

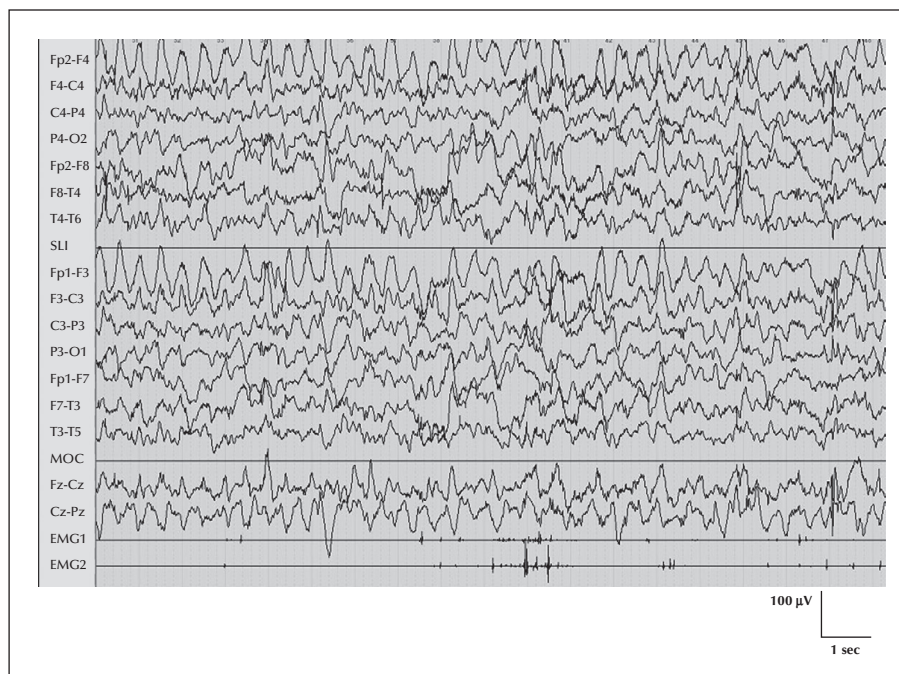


Figure 7. Patient II:5. EEG recorded after a GTCS showing continuous polymorphic delta activity over both frontal regions. Note myoclonic jerks recorded over both deltoid muscles (EMG1: right deltoid; EMG2: left deltoid).

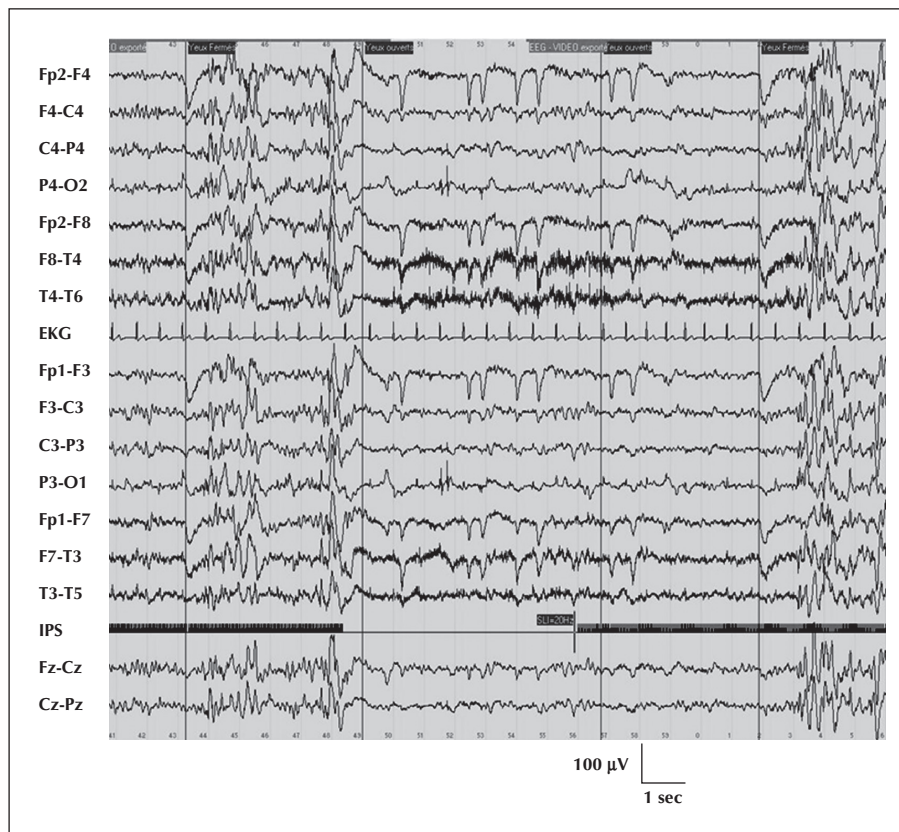


Figure 8. Patient II:6. Diffuse, irregular spike- and polyspike-and-wave discharges provoked by IPS at 16 and 20 Hz, associated with eye closure.

(Magaudda *et al.*, 2006). Indeed, normal or mildly slow background activity, generalized spike- and polyspike-and-wave discharges, photoparoxysmal response at IPS between 10 and 25 Hz, bursts of low voltage, and spikes and polyspikes over the central and vertex regions during sleep are usually found in ULD. Moreover, in ULD, generalized epileptiform discharges, both spontaneous or induced by IPS, tend to decrease after 10 to 15 years of disease (Ferlazzo *et al.*, 2007). For the family reported here, generalized epileptic discharges with a fast spike component, as well as a photoparoxysmal response, were observed in two out of four patients. Moreover, one affected sibling (Patient II:4) showed fast low-voltage spikes and polyspikes over the vertex and both centro-parietal regions during sleep. Unlike ULD, background activity was slow in all patients, and epileptic abnormalities persisted in all subjects for many years after disease onset.

A mutation in the *CERS1* gene, leading to the impairment of C18 ceramide biosynthesis, was shown to be the cause of this peculiar form of PME, providing evidence of the involvement of ceramide metabolism in the pathogenesis of these conditions. Moreover, we observed that impairment of CerS1 activity is associated with up-regulation of key ER stress

markers, highlighting a possible link between ceramide metabolism, ER stress response, and neurodegeneration.

The significance of impaired ceramide synthesis in the development of PMEs was also shown by Mosbech *et al.* (2014) who described a novel 27-kb heterozygous deletion in the *CERS2* gene in a PME patient. This was a 30-year-old man presenting febrile and afebrile seizures at 2 years of age. Developmental delay and myoclonus became evident at the age of 13 years, and GTCS and myoclonia increased over the years despite administration of several antiepileptic drugs. At the last follow-up visit, he presented 4-8 GTCS per month and severe myoclonus. Compared to parental controls, levels of *CERS2* mRNA, protein, and activity were reduced by ~50 per cent in fibroblasts isolated from this proband, resulting in significantly reduced levels of ceramides and sphingomyelins containing the very long-chain fatty acids C24:0 and C26:0.

However, *CERS1* and *CERS2* mutations were not found in a recent whole-exome sequencing study of 84 undiagnosed PME patients with unknown aetiology (Muona *et al.*, 2015) and, to date, have not been reported elsewhere (June 2015). Hence, we cannot

exclude that the mutation reported here represents a private mutation within this family.

Finally, the interest of these findings extends beyond the field of rare forms of PME's because ceramide represents a central element in sphingolipid metabolism, since disturbed sphingolipid levels may be found in ceroid lipofuscinosis (CLN1-6 and CLN8), as well as other common neurodegenerative diseases, such as Parkinson disease, Alzheimer disease, amyotrophic lateral sclerosis, and prion diseases. □

Disclosures.

None of the authors have any conflict of interest to disclose.

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Spinal muscular atrophy associated with progressive myoclonus epilepsy

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ABSTRACT – A rare syndrome characterized by lower motor neuron disease associated with progressive myoclonic epilepsy, referred to as “spinal muscular atrophy associated with progressive myoclonic epilepsy” (SMA-PME), has been described in childhood and is inherited as an autosomal recessive trait. SMA-PME is caused by mutation in the *ASAH1* gene encoding acid ceramidase. Ceramide and the metabolites participate in various cellular events as lipid mediators. The catabolism of ceramide in mammals occurs in lysosomes through the activity of ceramidase. Three different ceramidases (acid, neutral and alkaline) have been identified and appear to play distinct roles in sphingolipid metabolism. The enzymatic activity of acid ceramidase is deficient in two rare inherited disorders; Farber disease and SMA-PME. Farber disease is a very rare and severe autosomal recessive condition with a distinct clinical phenotype. The marked difference in disease manifestations may explain why Farber and SMA-PME diseases were not previously suspected to be allelic conditions. The precise molecular mechanism underlying the phenotypic differences remains to be clarified. Recently, a condition with mutation in *CERS1*, the gene encoding ceramide synthase 1, has been identified as a novel form of PME. This finding underlies the essential role of enzymes regulating either the synthesis (*CERS1*) or degradation (*ASAH1*) of ceramide, and the link between defects in ceramide metabolism and PME.

Key words: spinal muscular atrophy, myoclonus, Farber disease, progressive myoclonus epilepsies

Progressive myoclonus epilepsy (PME) (Minassian *et al.*, 2016) represents a heterogeneous group of epilepsies characterized by myoclonic and generalised seizures with progressive neurological deterioration (Girard *et al.*, 2013). A rare syndrome characterized by lower motor neuron disease associated with progressive myoclonic epilepsy (SMA-PME) has been described in childhood (Jankovic & Rivera, 1979; Halilolu *et al.*, 2002). This condition

is inherited as an autosomal recessive trait and the disease-causing gene has been recently identified. SMA-PME is caused by mutation in the *ASAH1* gene, which encodes acid ceramidase (Zhou *et al.*, 2012). A total of 11 affected individuals have been reported thus far, leading to a relatively precise phenotypic characterization (Zhou *et al.*, 2012; Dymont *et al.*, 2014, 2015; Rubboli *et al.*, 2015; Giráldez *et al.*, 2015).

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Mutations in the same gene are responsible for Farber disease, a very rare autosomal recessive condition with a distinct clinical phenotype, resulting from marked reduction or complete lack of acid ceramidase activity (Sugita *et al.*, 1972; Levade *et al.*, 2009). The marked difference in disease manifestations may explain why Farber and SMA-PME diseases were previously not suspected to be allelic conditions.

Several therapeutic strategies have been adopted to treat Farber disease, including allogeneic bone marrow transplantation and introduction of wild-type human acid ceramidase cDNA, using recombinant oncoretroviral or lentiviral vectors. The latter approach performed in non-human primates was recently shown to result in increased acid ceramidase activity. This underlines the importance of screening for *ASAH1* or acid ceramidase activity in patients with undiagnosed SMA or PME, for the purposes of diagnosis and future treatment.

Clinical description of SMA-PME

Based on the clinical description of cases reported to date ($n = 11$), SMA-PME is mainly characterized by progressive lower motor neuron degeneration followed by progressive myoclonic epilepsy (*table 1*) (Jankovic & Rivera, 1979; Haliloglu *et al.*, 2002; Zhou *et al.*, 2012; Dymant *et al.*, 2014, 2015; Rubboli *et al.*, 2015; Giráldez *et al.*, 2015).

Lower motor neuron disease manifestations

Early development milestones are usually normal, with an ability to walk unaided between 14 to 17 months of age. Progressive symmetric weakness and muscle atrophy of the lower and then the upper limbs are the main clinical features, usually occurring between 2.5 to 6 years of age (*table 1*). The patients begin to show slowness of gait and difficulties in standing from the sitting position. The motor deficit is purely peripheral, with a decrease in or absence of deep tendon reflexes. The disease is progressive, leading to the inability to stand up from the floor, the inability to sit unsupported, a lack of head control, difficulty swallowing, fasciculations of the tongue, severe scoliosis, and subsequent respiratory insufficiency. Electromyography (EMG) shows a chronic denervation process while muscle biopsy shows neurogenic atrophy, although there are no changes suggestive of a mitochondrial disorder.

However, in 1 of these 11 patients, atonic and absence seizures and myoclonic jerks were the main and first clinical features. Once the *ASAH1* gene mutations were identified, an EMG was performed, showing

evidence of motor neuron disease despite only mild proximal muscle weakness (Dymant *et al.*, 2014, 2015).

Progressive myoclonus epilepsy characteristics

Later on, myoclonic epilepsy is observed from 3 to 12 years of age (*table 1*). This is characterized by brief myoclonic seizures without loss of consciousness, generalized epileptic seizures and myoclonic jerks, myoclonic seizures, absences with head drop or postural lapses in the upper limbs, atonic seizures, or upper limb myoclonic jerks. The EEG shows subcortical myoclonic epileptiform abnormalities which are sensitive to hyperventilation, paroxysmal activity consisting of frequent, diffuse bursts of sharp waves and polyspike and wave complexes, bursts of generalized spike- and polyspike-and-wave complexes associated with myoclonic phenomena, generalized polyspike-and-wave discharges, or interictal bursts of posterior delta activity and ictal generalized spike-wave and polyspike-wave discharges. Brain MRI is most often normal or displays mild supratentorial and subtentorial cortical atrophy. Myoclonic seizures are most often refractory to antiepileptic drugs.

Other symptoms

Variable degrees of cognitive impairment occur. When reported, cognition is usually mildly impaired. Neither skin, joint abnormalities, or hoarseness of the voice, as observed in Farber disease, has been reported. Ophthalmological examination did not reveal corneal opacities or cherry red spots.

SMA-PME is caused by mutations in *ASAH1*

This condition is inherited as an autosomal recessive trait. Genome-wide linkage analysis combined with exome sequencing in two multiplex families and one sporadic case suffering from childhood SMA-PME revealed mutations in *ASAH1* which were responsible for the disease (Zhou *et al.*, 2012). *ASAH1*, located on chromosome 8, was the only gene to have a mutation that was shared by the unrelated affected individuals. The same missense mutation in exon 2 of *ASAH1* (NM_177924.3; c.125C>T; p.Thr42Met) was found in the affected individuals. The p.Thr42Met missense substitution affected an evolutionarily conserved amino acid among different species and was predicted to be damaging. In 2 families, both parents were heterozygous and affected siblings were homozygous for this missense mutation. In one family, the affected child carried compound heterozygous

Table 1. Literature review.

| | Zhou <i>et al.</i> , 2012 | | Rubboli <i>et al.</i> , 2015 | | | Dyment <i>et al.</i> , 2014 & 2015 | Giráldez <i>et al.</i> , 2015 |
|-------------------------------|--|--|---|--|---|---|--|
| | Family D | Family ITA | Family ITB | Case 1 | Case 2 | Case 3 | |
| Nb. of affected individuals | 3 | 2 | 1 | 1 | 1 | 1 | 1 |
| MOTOR DEVELOPMENT | | | | | | | |
| Ability to walk (m.) | 14 | Normal | Normal | Normal | 17 (abnormal deambulation) | Not reported | 17 (unsteady gait) |
| Age of onset of weakness (y.) | 5 | 4 to 5 | 5 | 6 | 2,4 | ? | 3 |
| Muscle symptoms | Proximal weakness | Progressive muscle weakness | Progressive muscle weakness | Progressive muscle weakness | Progressive muscle weakness and limb tremor | Mild proximal muscle weakness | Proximal muscle weakness |
| EMG | Chronic denervation process | Denervation process | Denervation-reinnervation process | Chronic denervation process | Chronic denervation process | Evidence of motor neuron disease | Chronic denervation process |
| Muscle biopsy | Neurogenic atrophy | Denervation process | Denervation-reinnervation | Neurogenic damage | Neurogenic damage | Neurogenic damage | Neurogenic atrophy |
| EPILEPSY | | | | | | | |
| Age of onset (y.) | 7 | 12 | 10 | 8 | 3 | 10 | 7 |
| Clinical symptoms | Brief myoclonic seizures without loss of consciousness | Generalized epileptic seizures and myoclonic jerks | Loss of consciousness associated with myoclonic jerks | Brief episodes of impairment of consciousness associated with “jerks” at the upper limbs | Staring and myoclonic jerks | Absence and atonic seizures; frequent myoclonic jerks | Multiple and brief absences, upper limb myoclonic jerks and/or head nodding episodes |

Table 1. (Continued)

| Zhou <i>et al.</i> , 2012 | | Rubboli <i>et al.</i> , 2015 | | | Dyment <i>et al.</i> , 2014 & 2015 | | Giráldez <i>et al.</i> , 2015 | |
|--|--|------------------------------|--|---|------------------------------------|---|---|---|
| Family D | Family ITA | Family ITB | Case 1 | Case 2 | Case 3 | | | |
| EEG | Subcortical myoclonic epileptiform abnormalities sensitive to hyperventilation | Not reported | Paroxysmal activity consisting in frequent diffuse bursts of sharp waves and poly-spike and wave complexes | Bursts of generalized spike- and polyspike-and-wave complexes associated with myoclonic phenomena | Similar to case 1 | Similar to case 1 | Generalized polyspike and wave discharges | Frequent interictal bursts of posterior delta activity and ictal generalized polyspike-wave and polyspike-wave discharges |
| Brain MRI | Normal | Normal | Normal | Diffuse supratentorial and subtentorial cortical atrophy | Normal | Normal then mildly increased size of the III and IV ventricles and mild volume loss | Normal | Normal |
| EVOLUTION | Progressive | Progressive | Progressive | Progressive | Progressive | Not reported | Progressive | Progressive |
| Age of death (y.) | 13-17 | na | 15 | 19 | na | na | na | na |
| ASAH1 MUTATION | | | | | | | | |
| Nucleotide (NM_177924.3) | c.125C>T; homozygous | c.125C>T; ASAH1 deletion | c.125C>T; homozygous | c.223_224insC; c.125C>T | c.177C>G; c.456A>C | c.850G>T; c.456A>C | c.125C>T; the other allele (deletion ?) | c.125C>T; lack of the other allele (deletion ?) |
| Protein | p.Thr42Met | p.Thr42Met | p.Thr42Met | p.Thr42Met | p.Val75Alafs*25; p.Thr42Met | p.Tyr59*; p.Lys152Asn | p.Thr42Met | p.Thr42Met |
| Acid ceramidase activity (% of normal value) | 32% | 32% | 32% | Not reported | Not reported | 5.5% | Not reported | Not reported |

Nb.: number, y. : year, m. : month ; nd : not done; na: not applicable

mutations, the c.125C>T missense mutation on one allele and a deletion of the whole gene on the other. Transient expression of the mutant cDNA in immortalized fibroblasts derived from an individual with Farber disease with very low acid ceramidase activity (less than 3.5 per cent of control value), revealed a mild reduction in acid ceramidase activity when compared to that of the wild type cDNA (about 32 per cent of the control value) (Zhou *et al.*, 2012). To analyze the effect of *ASAH1* loss-of-function *in vivo*, a morpholino antisense oligonucleotide of the *ASAH1* ortholog was used to knock down *ASAH1* in zebrafish embryos. Analysis of this model revealed a marked defect in motor neuron axonal branching associated with a significant increase in apoptosis in the spinal cord (Zhou *et al.*, 2012).

For other SMA-PME patients reported to date, *ASAH1* mutations were identified using sanger or whole-exome sequencing (Rubboli *et al.*, 2015; Dymment *et al.*, 2014, 2015; Giráldez *et al.*, 2015). The same c.125C>T mutation was identified in three additional patients, and other mutations including missense, non-sense, or large deletion of *ASAH1* were identified (table 1). Although acid ceramidase activity was reported *in vitro* to be mildly reduced as the consequence of the p.Thr42Met mutation (Zhou *et al.*, 2012), acid ceramidase activity in patient fibroblasts carrying the compound *ASAH1* heterozygous mutation (p.Gly284*; p.Lys152Asn) was markedly reduced (5.5 per cent of normal activity), similar to that observed in Farber patients. Surprisingly, this patient did not develop manifestations of Farber disease (Dymment *et al.*, 2014, 2015).

Pathogenesis of SMA-PME is linked to *ASAH1* gene mutations

Ceramide is synthesized in the endoplasmic reticulum and transported by the ceramide-transfer protein, CERT, to the trans-Golgi membrane where it is converted to sphingomyelin by the sphingomyelin synthase-1,2. Ceramide and the metabolites (sphingosine and sphingosine 1-phosphate) participate in various cellular events as lipid mediators. The catabolism of ceramide in mammals occurs in lysosomes through the activity of ceramidase. Three different ceramidases (acid, neutral, and alkaline) have been identified and characterized according to optimum pH and primary structure; these include the acid (*ASAH1*), neutral (*ASAH2*), and alkaline ceramidases (*ASAH3*). The three families of ceramidase appear to play distinct roles in sphingolipid metabolism.

The enzymatic activity of acid ceramidase is deficient in two rare inherited disorders, Farber disease

and SMA-PME. Importantly, in addition to the role of acid ceramidase in ceramide catabolism, the enzyme may have other functions depending on its subcellular location and the local pH. The other putative functions of *ASAH1* may account for the clinical spectrum of the disease associated with *ASAH1* mutation. Refined characterization of acid ceramidase activity in the subcellular domain should contribute to a better understanding of the genotype-phenotype correlation.

Ceramides are the precursors to complex sphingolipids, which are important for normal functioning of both the developing and mature brain. Recently, a mutation in *CERS1*, the gene encoding ceramide synthase 1, has been identified as a novel cause of PME (Vanni *et al.*, 2014). This data underlies the essential role of enzymes in terms of regulating the synthesis (*CERS1*) or degradation (*ASAH1*) of ceramide and the link between ceramide metabolism defects and PME.

Molecular basis of Farber versus SMA-PME diseases

Farber disease is a rare and severe autosomal recessive lysosomal storage disorder (Levade *et al.*, 2009) which, like SMA-PME, is linked to the disease gene, *ASAH1*. It is characterized by a marked deficiency in acid ceramidase activity (Sugita *et al.*, 1972). Farber disease is characterized by a severe early onset (from 2 weeks to 4 months of age) of a unique triad of clinical manifestations including painful and progressively deformed joints, subcutaneous nodules, and progressive hoarseness as the result of laryngeal involvement. The illness advances rapidly, with progressive neurological deterioration, leading to death at a mean age of 1.45 years. At late stages of disease progression, 13 per cent of Farber disease patients display hypotonia and muscular atrophy with reduced or absent deep tendon reflexes and signs of denervation, as observed on EMG. These data strongly suggest that while a dramatic reduction in acid ceramidase activity leads to Farber disease, a milder reduction in enzymatic activity leads to a later onset of symptoms restricted to spinal cord motor neurons and subsequently other areas of the CNS responsible for PME. However, as reported recently (Dymment *et al.*, 2014, 2015), one patient presenting with features of PME, with mild motor weakness, had a marked reduction in acid ceramidase activity, similar to that observed in Farber patients, but did not present any manifestations of Farber disease, suggesting that additional factors are likely to be involved in the different phenotypic expression.

Animal models and therapeutic research strategies

Previously, homozygous *ASAH1* knockout mouse embryos were shown not to live beyond the four-cell stage, indicating that acid ceramidase plays an essential role during development (Eliyahu *et al.*, 2007). Recently, a knock-in mouse model was created by introducing a single-nucleotide mutation, identified in human Farber patients, into the murine *ASAH1* gene (Pro362Arg) (Alayoubi *et al.*, 2013). Homozygous mutant mice displayed Farber disease manifestations and died within 7–13 weeks. Treating mutant mice during the neonatal period with a single injection of lentivector expressing human acid ceramidase diminishes the severity of the disease. It would be of great interest to determine whether, using this model, partial correction of acid ceramidase deficiency could prevent both Farber and SMA-PME disease manifestations or only the Farber phenotype, which should reinforce the hypothesis that the clinical expression of the disease correlates with residual acid ceramidase activity. This model should contribute to a better understanding of the pathogenesis of Farber and SMA-PME diseases and represents a valuable tool towards developing therapeutic strategies.

While bone marrow transplantation has been reported to be effective in relieving joint contractures and subcutaneous nodules in a patient with Farber disease (Yeager *et al.*, 2000), the affected individual developed progressive muscle weakness and features consistent with the occurrence of lower motor neuron involvement, the main clinical features found in SMA-PME individuals. More recently, a preclinical gene therapy study for Farber disease involving a lentiviral vector was performed in non-human primates. Acid ceramidase activity was detected above normal levels in various cell types, including bone marrow cells, spleen and liver 1 year after lentiviral vector transduction (Walia *et al.*, 2011).

Genetic counselling

The identification of the *ASAH1* gene mutations found in SMA-PME patients has greatly improved diagnostic testing and family-planning options of SMA-PME family members. Because the risk of recurrence of the disease in the sibling of an affected child is high (1/4), information should be given to parents early after diagnosis. Explanations regarding molecular pathology, risk evaluation, and prenatal testing possibilities should be given to the parents and their relatives within the context of a genetic counselling visit.

Conclusion

Mutations in *ASAH1*, which encodes acid ceramidase, are the major cause of SMA-PME. Based on an overview of patients reported to date, the main clinical characteristic features are onset, most often within the first 6 years of age, characterized by lower motor neuron disease leading to progressive muscle weakness, followed by the occurrence of clinical and EEG characteristics of myoclonus epilepsies, and a progressive course. This data underlies the link between ceramide metabolism defects and PME. Taking into account ongoing therapeutic research for Farber disease, a disease allelic to SMA-PME, screening for *ASAH1* or acid ceramidase activity should be proposed for the diagnosis and future treatment of patients with PME or SMA. □

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Myoclonus epilepsy and ataxia due to potassium channel mutation (MEAK) is caused by heterozygous *KCNC1* mutations

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ABSTRACT – Progressive myoclonus epilepsy (PME) is a distinct group of seizure disorders characterized by gradual neurological decline with ataxia, myoclonus and recurring seizures. There are several forms of PME, among which the most recently described is MEAK - myoclonus epilepsy and ataxia due to potassium channel mutation. This particular subtype is caused by a recurrent *de novo* heterozygous mutation (c.959G>A, p.Arg320His) in the *KCNC1* gene, which maps to chromosome 11 and encodes for the Kv3.1 protein (a subunit of the Kv3 subfamily of voltage-gated potassium channels). Loss of Kv3 function disrupts the firing properties of fast-spiking neurons, affects neurotransmitter release and induces cell death. Specifically regarding Kv3.1 malfunctioning, the most affected neurons include inhibitory GABAergic interneurons and cerebellar neurons. Impairment of the former cells is believed to contribute to myoclonus and seizures, whereas dysfunction of the latter to ataxia and tremor. Phenotypically, MEAK patients generally have a normal early development. At the age of 6 to 14 years, they present with myoclonus, which tends to progressively worsen with time. Tonic-clonic seizures may or may not be present, and some patients develop mild cognitive impairment following seizure onset. Typical electroencephalographic features comprise generalized epileptiform discharges and, in some cases, photosensitivity. Brain imaging is either normal or shows cerebellar atrophy. The identification of MEAK has both expanded the phenotypic and genotypic spectra of PME and established an emerging role for *de novo* mutations in PME.

Key words: MEAK, *KCNC1*, progressive myoclonus epilepsies, myoclonus, seizures, potassium channel mutations, ataxia

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The term ‘progressive myoclonus epilepsy’ (PME) (Minassian *et al.* 2016) provides us with a broad, but reliable, characterization for

this distinct subgroup of seizure disorders. ‘Progressive’ depicts the gradual neurological decline seen in patients affected with a PME.

‘Myoclonus’ and ‘epilepsy’ summarize the other two cardinal clinical features observed in these individuals (Berkovic *et al.*, 1986). Aetiologically, most PME are the result of genetic abnormalities. The vast majority are inherited in an autosomal recessive fashion, although some cases exhibit autosomal dominant or mitochondrial inheritance.

Despite the impressive range of specific forms of PME, their phenotypes are fairly similar, which makes accurate clinical diagnosis very challenging, especially in the early stages. However, significant heterogeneity exists in terms of the genetic causes of the subtypes of progressive myoclonus epilepsies.

As is the case with countless other diseases in the realm of neurology, a proportion of PME cases still have no identified genetic basis. In this context, a cohort of 84 unconfirmed, unrelated PME patients was extensively investigated in a recent multicentre study. With a view to trying to elucidate their molecular diagnoses, all cases underwent whole-exome sequencing, and Sanger sequencing was performed for a secondary cohort of 28. The most remarkable finding was the identification of a recurrent heterozygous *de novo* mutation in the *KCNC1* gene, which encodes for a subunit of a specific potassium channel. The new PME subtype was named ‘myoclonus epilepsy and ataxia due to potassium channel mutation’ (MEAK) (Muona *et al.*, 2015). Notably, prior to this study, *KCNC1* had never been associated with any human disease. In addition, this group of researchers importantly demonstrated that *de novo* mutations play a significant role in the genesis of progressive myoclonus epilepsies.

This article aims to review the most recent PME subtype described: MEAK. We begin by introducing the micro-world of potassium channels, since an understanding of this is of fundamental importance. We then focus on how *KCNC1* was identified to be associated with MEAK and discuss how the effects of such a mutation are translated into clinical signs and symptoms. Finally, the phenotypes of all the reported individuals with MEAK are discussed.

Potassium ion channels

Potassium ion channels are ubiquitous membrane proteins responsible for a range of cellular functions, including maintenance of membrane potential, regulation of cell volume, and electrical excitability modulation. The latter is particularly important for the membrane physiology of excitable cells, such as neurons. According to their functional properties, potassium channels can be categorized into several groups, such as **voltage-gated potassium channels**, calcium-activated potassium channels, and sodium-activated potassium channels, among others.

Furthermore, each of these groups can be subdivided into families and subfamilies, based on molecular similarity (Sansom *et al.*, 2002).

Voltage-gated potassium channels (Kv channels) play an essential role in the generation and propagation of electrical impulses in the nervous system. By enabling the selective flow of potassium ions through neuronal membranes (upon changes in transmembrane potentials), Kv channels help set the resting potential and degree of excitability of the membrane (repolarization), influence action potential waveforms and firing patterns, and modulate synaptic activity (Ried *et al.*, 1993). Voltage-gated potassium channels are classified into four subfamilies: Kv1, Kv2, Kv3, and Kv4.

Kv3 channels, in particular, are known for their high activation threshold and fast activation and deactivation properties (Rudy & McBain, 2001). Kv3 channels are crucial components of the circuitry of neurons that are able to fire action potentials at high frequencies or follow high frequency inputs (Wang *et al.*, 2007). The Kv3 subfamily is composed of four subunits, Kv3.1, Kv3.2, Kv3.3, and Kv3.4, which are encoded by four genes, *KCNC1* to *KCNC4*, respectively.

These four subunits assemble as either homomers or heteromers to form voltage-gated tetrameric potassium channels. Each of the four subunits consists of 6 transmembrane segments (S1–S6) with a re-entrant P-loop region. The transmembrane segments S1–S4 are referred to as the voltage-sensing domain, of which the primary voltage-sensing unit is S4. The segments S5–S6 represent the ion-conducting pore domain. Upon membrane de- or hyper-polarization, which is sensed by positively charged arginine residues at the S4 segment, the S4 segment undergoes the largest reorientation, leading to channel opening and generation of transient gating currents (Aggarwal & MacKinnon, 1996; Chanda & Bezanilla, 2008) (*figure 1*).

Within the *KCNC* gene family, *KCNC3* has been previously reported as a human disease associated gene. *KCNC3* mutations have been recognized as a cause of spinocerebellar ataxia type 13 (*SCA13*) (Herman-Bert *et al.*, 2000; Middlebrooks *et al.*, 2013). Only very recently, *KCNC1* mutations have been identified as a cause of human disease: myoclonus epilepsy and ataxia due to potassium channel mutation (MEAK).

Discovering *KCNC1* mutations in myoclonus epilepsy and ataxia (MEAK)

Eighty-four clinically confirmed cases of PME without clear genetic aetiology from multiple centres (including Europe, North America, Asia, and Australia), were investigated using whole-exome sequencing. A recurrent heterozygous missense mutation, c.959G>A

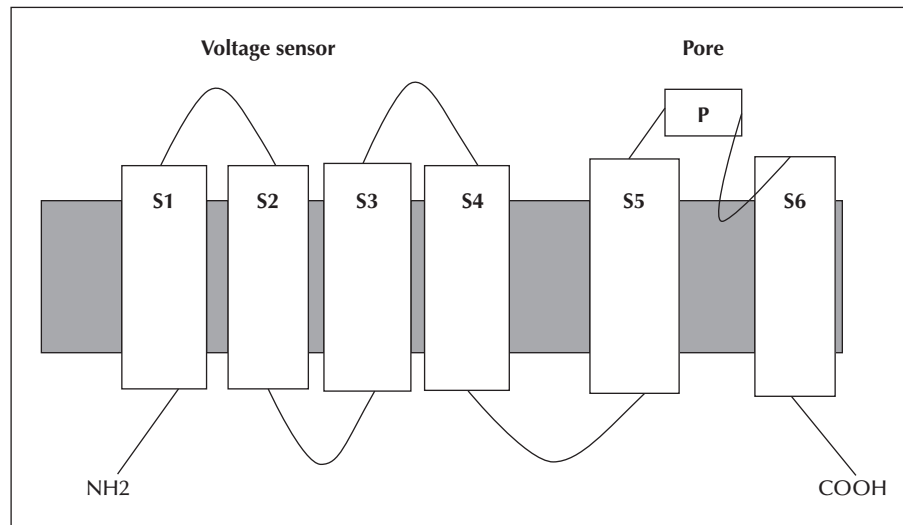


Figure 1. Transmembrane topology of a Kv channel subunit.

(p.Arg320His), in *KCNC1* was identified in 11 unrelated patients (13.1 per cent). Sanger sequencing confirmed the mutation in two new cases, as well as in an affected sister and two affected children of one of the original 11 cases. In total, 16 cases, 13 of them unrelated, had the same *KCNC1* mutation. The parents of all patients with the *KCNC1* mutation were unaffected. Segregation analysis, where DNA was available for both parents, revealed that in each case the mutation occurred *de novo*. The family with four MEAK cases was further evaluated to rule out parental mosaicism (Muona *et al.*, 2015). Based on a recently published mutation model (Samocha *et al.*, 2014), the rate of this specific mutation was estimated at 1.75×10^{-7} mutations per person, representing an occurrence in one out of every 5,700,000 conceptions.

Pathogenicity of *KCNC1* mutations

KCNC1 maps to chromosome 11, at 11p15.1, and encodes the Kv3.1 potassium channel (Ried *et al.*, 1993). The c.959G>A mutation causes a substitution of histidine for arginine at codon 320 of the Kv3.1 protein (p.Arg320His). The targeted arginine is a highly evolutionary conserved residue in segment S4, which constitutes the main voltage sensor of Kv3 channels. Functional analysis showed that, upon membrane depolarization, mutant channels produced hardly detectable currents (Muona *et al.*, 2015). Interestingly, experiments also indicated a dominant-negative effect of the mutant channel. When the mutant subunit and the wild-type protein were co-expressed at a ratio of 1:1, an 80 per cent decrease in the expected current was observed (Muona *et al.*, 2015).

Previous studies have shown that loss of Kv3 function (such as that caused by p.Arg320His in *KCNC1*) disrupts

the firing properties of fast-spiking neurons (Erisir *et al.*, 1999; Issa *et al.*, 2011; Rudy & McBain, 2001), affects neurotransmitter release (Sabatini & Regehr, 1997), and induces cell death (Irie *et al.*, 2014). Specifically, in terms of Kv3.1 malfunctioning, the most affected neurons include inhibitory GABAergic interneurons (Gan & Kaczmarek, 1998; Rudy & McBain, 2001) and cerebellar neurons. Consequently, within the context of a p.Arg320His mutation, the physiology of both these types of neuronal cells would be compromised and could, eventually, degenerate. The implications of these outcomes are useful in attempting to explain the clinical scenario of subjects with MEAK. The first implication would be disinhibition following impaired firing of fast-spiking GABAergic interneurons, which is believed to contribute to myoclonus and tonic-clonic seizures. The second implication would be the dysfunction and degeneration of cerebellar neurons, which is hypothesized to contribute to motor impairment and ataxia.

MEAK phenotypes

The phenotypes of the 16 patients with MEAK were fairly similar. Early development was, as a rule, normal. The first symptom tended to be myoclonus, with an onset ranging from the age of 6 to 14 years. Myoclonus progressively worsened. Gait disturbances due to myoclonus led to the use of a walking aid or wheelchair by adolescence or early adulthood in most patients. Tonic-clonic seizures were present, albeit infrequent. Learning difficulties before seizure onset were not common. Mild cognitive decline subsequent to seizure onset, nonetheless, was shared by roughly half of the individuals. On electroencephalogram, these patients had generalized epileptiform

discharges, with photosensitivity in some cases. Magnetic resonance imaging was either normal or showed cerebellar atrophy.

The early clinical presentation and evolution of MEAK resembles that of Unverricht-Lundborg disease (ULD). The age at onset, moderate-to-severe myoclonus, infrequent tonic-clonic seizures, and mild, if any, cognitive decline, are characteristic of both MEAK and ULD. As MEAK evolves, however, it might be clearly distinguished from ULD, as patients with MEAK usually suffer a more severe course. In terms of molecular diagnosis, however, the two entities are easily differentiated, irrespective of the time of diagnosis. While MEAK is caused by *KCNC1* mutations, ULD is most frequently a result of mutations in the promoter region of *CSTB*, which leads to a massive reduction in levels of cystatin B, a lysosomal protease inhibitor (Rinne *et al.*, 2002; Girard *et al.*, 2013).

Conclusion

The identification of MEAK and its genetic basis have widened the phenotypic and genotypic spectra of PME. Given that pathogenic *KCNC1* mutation is estimated to affect one in every 5,700,000 conceptions, many undetermined PME cases worldwide may now be solved. Diagnosing these individuals is likely to have a direct impact on counselling and education for patients and caregivers.

In addition, an emerging role for *de novo* mutations in progressive myoclonus epilepsies has been established. Finally, an understanding of the pathophysiology of MEAK provides singular insights into potential therapeutic interventions. □

Disclosures.

None of the authors have any conflict of interest to disclose.

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Autosomal dominant cortical tremor, myoclonus and epilepsy

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ABSTRACT – The term ‘cortical tremor’ was first introduced by Ikeda and colleagues to indicate a postural and action-induced shivering movement of the hands which mimics essential tremor, but presents with the electrophysiological findings of cortical reflex myoclonus. The association between autosomal dominant cortical tremor, myoclonus and epilepsy (ADCME) was first recognized in Japanese families and is now increasingly reported worldwide, although it is described using different acronyms (BAFME, FAME, FEME, FCTE and others). The disease usually takes a benign course, although drug-resistant focal seizures or slight intellectual disability occur in some cases. Moreover, a worsening of cortical tremor and myoclonus is common in advanced age. Although not yet recognized by the International League Against Epilepsy (ILAE), this is a well-delineated epilepsy syndrome with remarkable features that clearly distinguishes it from other myoclonus epilepsies. Moreover, genetic studies of these families show heterogeneity and different susceptible chromosomal loci have been identified.

Key words: cortical tremor, myoclonus, epilepsy, genetics, autosomal dominant, progressive myoclonus epilepsies

In 1990, Ikeda *et al.* described an action and postural tremor originating from the cerebral cortex, which was thus defined as a cortical tremor. Due to its electrophysiological features, this involuntary movement was considered to be a variant of cortical reflex myoclonus (Ikeda *et al.*, 1990). Uyama and colleagues reported 54 patients in 7 families, estimating a prevalence of approximately 1:35,000 based on their observation in Kumamoto Prefecture (Uyama *et al.*, 2005).

Cortical tremor was recognized to occur in families often in association with generalized tonic-clonic

seizures, and this observation led to the definition of a peculiar autosomal dominant (AD) trait named ‘benign adult familial myoclonic epilepsy’ (BAFME). BAFME was first described in Japanese families and the genetic locus mapped to chromosome 8q24. However, several European families fulfilling the diagnostic criteria for BAFME have been described and in these families, linkage to chromosome 8q24 has been excluded (reviewed in Striano & Zara [2010]). These findings suggest a worldwide distribution and genetic heterogeneity for this condition.

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Guerrini *et al.* (2001) described a family who also presented drug-resistant focal seizures and mild intellectual disability in three affected individuals and considered this phenotype as a peculiar syndrome named 'AD cortical myoclonus and epilepsy' (ADCME). The mapping of a gene on chromosome 2p11.1-q12.2 reinforced the hypothesis that ADCME was an independent (genetically distinct from BAFME) clinical entity. A founder effect may possibly explain the high frequency of families originating from the same topographic areas of Japan and southern Italy (Uyama *et al.*, 2005; Madia *et al.*, 2008). An in-frame insertion/deletion in the α_2 -adrenergic receptor subtype B gene (*ADRA2B*), encoding the α_2 -adrenergic receptor subtype B, has been reported in two apparently unrelated ADCME pedigrees of Italian origin (De Fusco *et al.*, 2014). This mutation alters several conserved residues of the third intracellular (3i) loop and alters the binding with the scaffolding protein called spinophilin upon neurotransmitter activation, thus increasing epinephrine-stimulated calcium signalling.

Additional loci have also been mapped to 5p15.31-p15 and 3q26.32-q28 in single families of French (Depienne *et al.*, 2010) and Thai (Yeetong *et al.*, 2013) origin. Stogmann *et al.* (2013) recently described a consanguineous Egyptian family presenting focal epilepsy, neuropsychiatric features, borderline cognitive level, and myoclonus, resembling BAFME, but inherited in an autosomal recessive manner. A homozygous deletion (c.503_503delG) leading to a frameshift in the coding region of *CNTN2* and segregating in the 5 affected family members was identified. The *CNTN2* gene (mapped at 1q32.1) encodes for contactin 2, a glycosylphosphatidylinositol-anchored neuronal membrane protein, which is necessary to maintain voltage-gated potassium channels at the juxtaparanodal region. Finally, a unique south Indian community, including 241 patients with ADCME belonging to 48 families has been described. The 48 families are domiciled in 2 southern districts of Tamilnadu, India, which belongs to the Nadar community, and their origin is confined to these southern districts, with reported unique genetic characteristics. This is the largest single report of ADCME worldwide (Mahadevan *et al.*, 2016).

The pathophysiological and biochemical bases of this condition also remain largely speculative. Both clinical and electrophysiological features of the syndrome suggest a cortical hyperexcitability, which may be the result of decreased cortical inhibition by the cerebellum via its cerebello-thalamo-cortical projections (Striano *et al.*, 2005). Sporadic post-mortem histological studies have shown evidence of cerebellar pathology (Uyama *et al.*, 2005).

Clinical features

Age at onset is highly variable (ranging from 11 to 50 years) but the disease usually starts with a slight hand 'tremor' within the second decade of life and progresses to rare tonic-clonic seizures and myoclonus by the third or fourth decade of life. Prevalence is unknown, but this condition is likely to be under-recognized. The dominant clinical picture is characterized by cortical tremor, myoclonus and epilepsy.

Cortical tremor is an action and postural fine shivering movement consisting of continuous, arrhythmic, mainly distal, fine twitches in the hands. There is no significant progression over time, but a worsening of the disturbance may be observed over the age of 70. The cortical tremor is enhanced by emotion or fatigue and may be easily misdiagnosed as essential tremor, but may be distinguished from the latter clinically (figure 1A). To definitively distinguish between cortical tremor and benign essential tremor requires a neurophysiological demonstration of cortical origin (Striano *et al.*, 2005).

In addition to cortical tremor, most patients present distal segmental, arrhythmic, erratic myoclonic jerks in the upper limbs which are exacerbated by posture and action. The involvement of more proximal, as well as facial, muscles, particularly the eyelids, is also possible. The onset of myoclonus is difficult to clearly establish but usually starts at around the same age as that for cortical tremor (Striano & Zara, 2010).

Most patients experience generalized tonic-clonic seizures starting later than the tremor. The age at first seizure ranges widely between 12 and 67 years, with a peak around the age of 30. Seizures are generally rare (up to 5-10 episodes over the person's lifetime) and are not preceded by any warning. However, in some cases, they may be heralded by progressively increasing myoclonic jerks. Precipitating factors, such as sleep deprivation, emotional stress, and photic stimulation are often reported (Striano *et al.*, 2005; Uyama *et al.*, 2005). In rare cases, patients may also present with drug-resistant complex partial seizures and focal EEG abnormalities (Guerrini *et al.*, 2001).

Patients usually present normal cognitive levels. However, mild-to-moderate intellectual disability may be present in some cases, particularly at a more advanced age (figure 2) (Striano *et al.*, 2005; Coppola *et al.*, 2011). Night blindness, with a reduced b-wave response on electroretinography, has been reported in three patients from Japan and migraine attacks have been reported as a predominant feature in a Turkish family (reviewed by Striano *et al.* [2005] and Uyama *et al.* [2005]).

Instrumental diagnostic procedures

Detailed electrophysiological investigations are essential to confirm the cortical origin of myoclonus. However, some of these electrophysiological features may be masked by antiepileptic treatments. The EEG

background activity is usually normal or slightly slow in the slower alpha band. Generalized paroxysmal abnormalities and photoparoxysmal responses are frequently found in patients without therapy (Striano *et al.*, 2005; Uyama *et al.*, 2005). Furthermore, a photomyogenic response (*i.e.* a muscular, mainly anterior

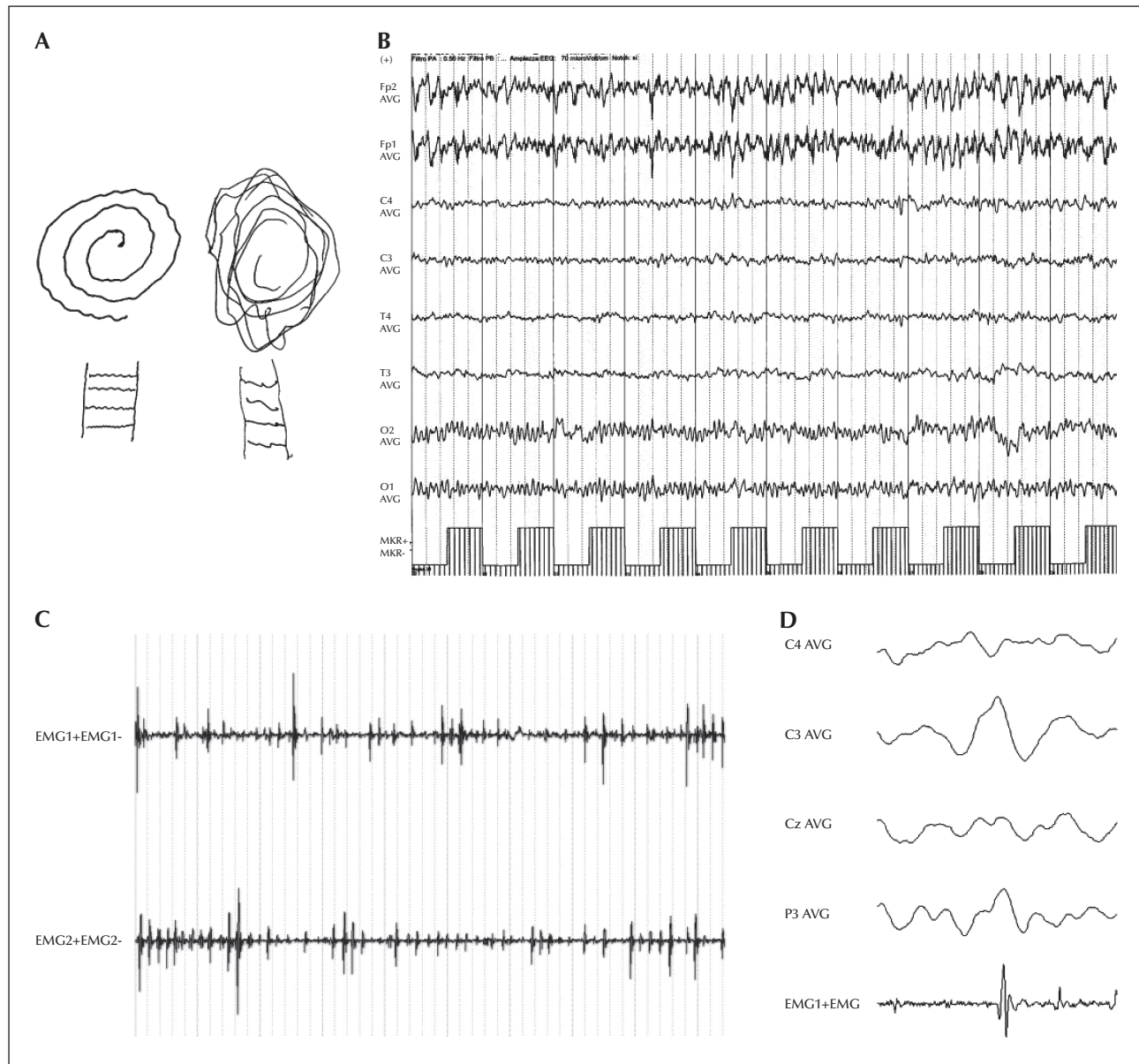


Figure 1. Electroclinical and MRI features of familial cortical tremor, myoclonus and epilepsy. (A) free-hand drawing (Archimedes' spiral and ladder) showing the differences between essential (left) and cortical (right) tremors. The cortical tremor is fairly irregular, and sudden, brisk jerks cause disruption to the drawing. (B) EEG of a patient during photic stimulation with eyes closed, showing the photomyoclonic response consisting of increasing, mainly anteriorly, myogenic potentials related to each flash stimuli. (C) EMG recording of bursts between agonist and antagonist muscles (EMG1: right wrist extensor; EMG2: right wrist flexor) with extended arms; irregular, high-frequency, short EMG bursts without the regular alternating pattern typically found in tremors. (D) Jerk-locked averaging analysis shows a positive-negative potential, recognizable over the left centroparietal electrodes, preceding myoclonus by about 30 ms (right wrist extensor muscle; number of triggers = 100).

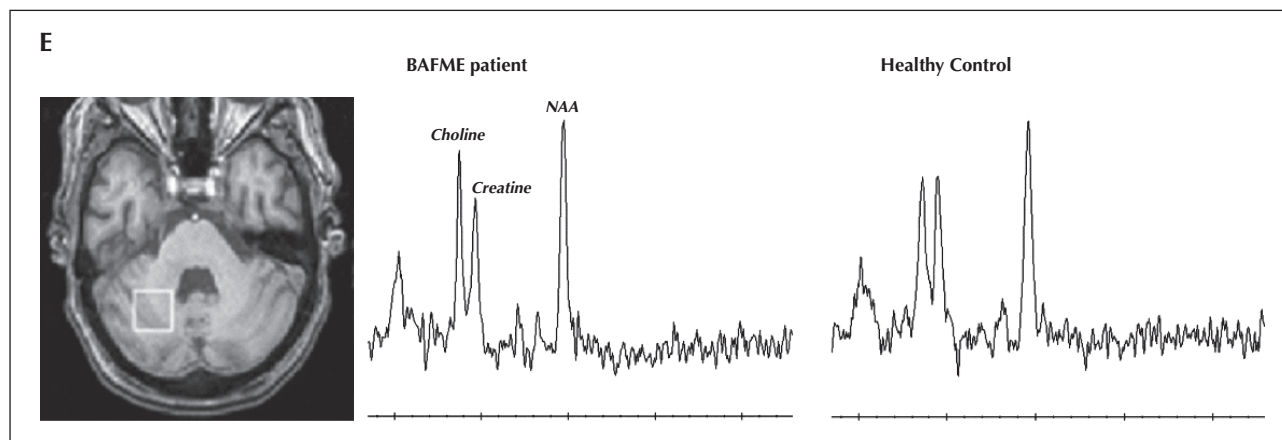


Figure 1. (E) 1H-MR Spectroscopy using a PRESS sequence (TR 1,500 ms; TE 144 ms) showing abnormal spectral peak areas at 3.22 ppm, corresponding to choline (location of the 8 cm³ voxel: right cerebellar hemisphere). Reproduced from Striano and Zara (2010), with permission.

response, synchronous with photic stimulation) may be present (*figure 1B*). Focal paroxysmal activity can occur in some patients, in addition to generalized EEG abnormalities (Guerrini *et al.*, 2001; Striano & Zara, 2010).

Polymyographic recording helps to differentiate between tremor and myoclonus. The electromyographic (EMG) pattern is consistent with irregular, arrhythmic or semi-rhythmic, high-frequency (around 10/s) myoclonic jerks. EMG bursts last about 50 ms and are usually synchronous between agonist and antagonist muscles, but do not regularly alternate between agonist/antagonist muscles, as in essential tremor (*figure 1C*). Jerk-locked averaging analysis commonly discloses a positive-negative, biphasic, premyoclonic spike or a more complex pattern of wavelets related to myoclonus on the contralateral sensorimotor regions (*figure 1D*). The evaluation of somatosensory evoked potentials and long-loop reflex I may show an enlargement of cortical components (P25-N33 amplitude larger than 8.5–15 μ V) and enhanced long-latency (40 ms) C-reflex response evoked by stimulation of the peripheral nerve. A reduction in the resting motor threshold intensity and post-motor evoked potential silent period has been reported in a few patients evaluated by transcranial magnetic stimulation, indicating that central motor inhibitory mechanisms are impaired in these cases (Guerrini *et al.*, 2001). MRI examination is usually normal although minor, non-specific abnormalities (such as mild enlargement of the subarachnoid spaces of the lateral ventricles) are sometimes reported. An MRI spectroscopy study revealed an elevated choline/creatine ratio in the cerebellum cortex of patients compared with controls (*figure 1E*) (Striano *et al.*, 2009).

Differential diagnosis

Cortical tremor may be easily misinterpreted as essential tremor and seizures may be overlooked or considered to be coincidental, or interpreted as a side effect of valproate treatment (Striano *et al.*, 2005). The clinical observation and demonstration of cortical reflex myoclonus by means of electrophysiological investigation enables confirmation of the diagnosis. ADCME must be differentiated from epilepsy syndromes with prominent myoclonus features. In particular, patients may easily be misdiagnosed with juvenile myoclonic epilepsy (JME) because of the occurrence of myoclonic jerks and generalized tonic-clonic seizures. However, JME clinically differs with regards to the absence of cortical tremor and predominant proximal myoclonic seizures which typically occur upon awakening. The absence of ataxia and dementia, adult onset, and the usually benign outcome of epilepsy differentiate ADCME from progressive myoclonus epilepsies (Striano & Zara, 2010; Minassian *et al.*, 2016).

Treatment and evolution

Cortical tremor is not responsive to alcohol or L-dopa/carbidopa but improves with antiepileptic drugs (Ikeda *et al.*, 1990; Striano *et al.*, 2005; Uyama *et al.*, 2005). Valproate, levetiracetam and benzodiazepines produce the greatest benefit against cortical tremors and myoclonus, combining both antiepileptic and antimyoclonic activity. In some cases, epilepsy may be difficult to treat.

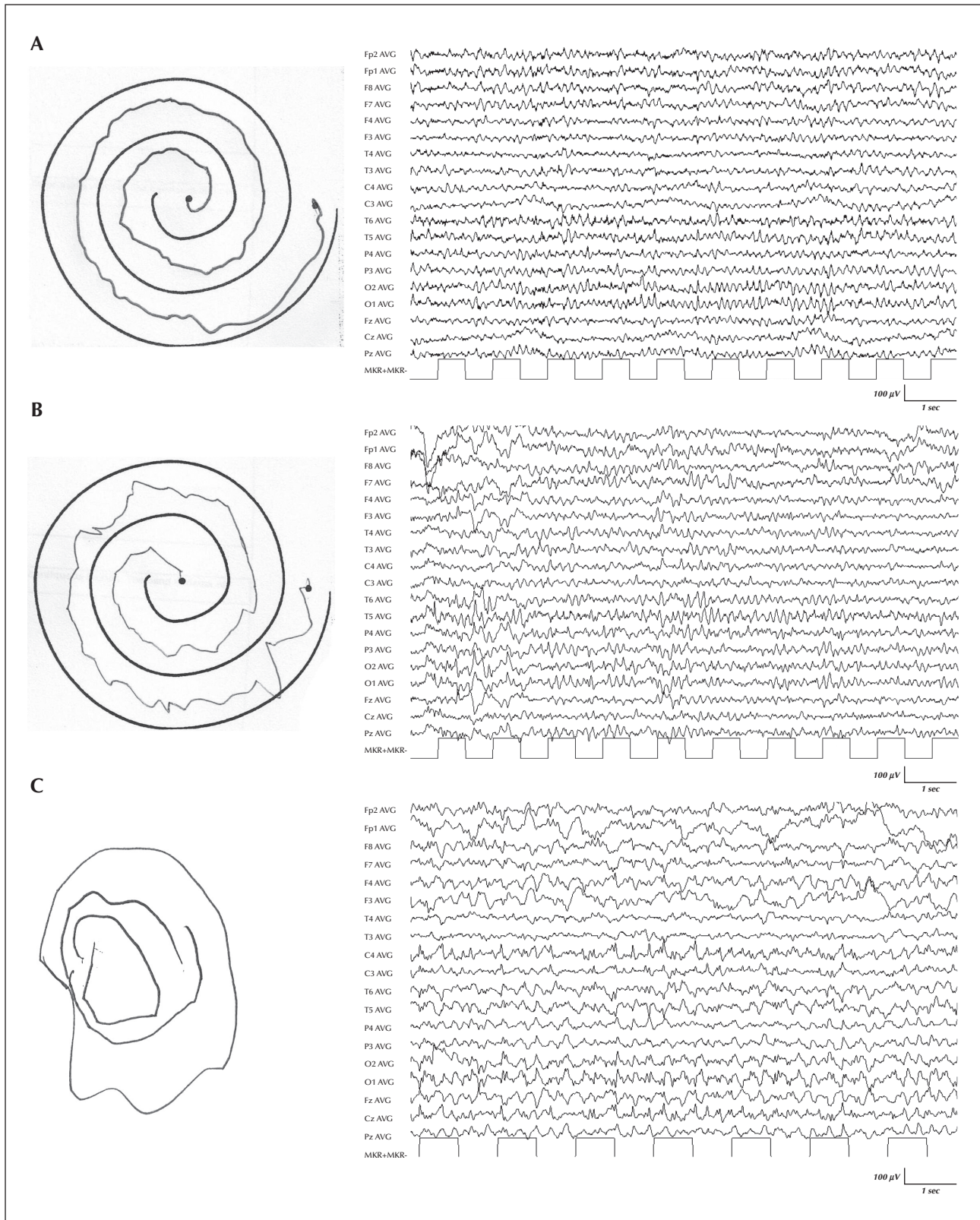


Figure 2. Free-hand drawing, Archimede's spiral hands (left) and basal EEG of patient C/3 obtained at the age of 59 (A), 70 (B), and 80 years (C), showing worsening of myoclonus and progressive slowing of EEG background activity. Reproduced from Coppola *et al.* (2011), with permission.

As for other idiopathic generalized epilepsies, some antiepileptic drugs may precipitate myoclonic status. In these cases, a correct diagnosis and prompt discontinuation of the drug may reverse a potentially severe, life-threatening condition (Striano *et al.*, 2007). In advanced age, worsening of the myoclonus is possible, as well as difficulty in walking and mild ataxia (Striano & Zara, 2010; Coppola *et al.*, 2011). □

Disclosures.

None of the authors have any conflict of interest to disclose.

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Myoclonus and seizures in progressive myoclonus epilepsies: pharmacology and therapeutic trials

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ABSTRACT – Generalized motor seizures, usually tonic-clonic, tonic-vibratory, myoclonic or clonic, and stimulus-sensitive/action myoclonus are typical features of progressive myoclonus epilepsies (PMEs). Despite the introduction of many anticonvulsants, the treatment of these symptoms, particularly myoclonus, remains challenging, due to the incomplete and often transitory effects of most drugs. Moreover, treatment is only symptomatic, since therapy targeting the underlying aetiology for these genetic conditions is in its infancy. Traditional antiepileptic drugs for the treatment of PMEs are valproate, clonazepam, and phenobarbital (or primidone). These drugs may improve the overall performance of PME patients by decreasing their generalized seizures and, to a lesser extent, their myoclonic jerks. Newer drugs which have been shown to be effective include piracetam, levetiracetam, topiramate, zonisamide, and possibly perampanel for Lafora disease. The potential of other drugs (such as L-tryptophan and N-acetylcysteine) and procedures (such as vagal and deep brain stimulation) has also been discussed. The available data on the efficacy of drugs are mainly based on small series or anecdotal reports. Two prospective, randomized, double blind studies investigating the novel SV2A ligand, brivaracetam, in genetically confirmed Unverricht-Lundborg patients have been performed with disappointing results. When treating PMEs, particular care should be paid to avoid drugs known to aggravate myoclonus or myoclonic seizures, such as phenytoin, carbamazepine, oxcarbazepine, lamotrigine, vigabatrin, tiagabine, gabapentin, and pregabalin. The emergency treatment of motor status, which often complicates the course of PMEs, consists of intravenous administration of benzodiazepines, valproate, or levetiracetam.

Key words: progressive myoclonus epilepsies, myoclonus, valproate, brivaracetam, perampanel, drugs worsening myoclonus

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Progressive myoclonus epilepsies (PMEs) (Minassian *et al.*, 2016) are a group of uncommon diseases characterized by the association between myoclonus, epileptic seizures, and neurological deterioration, particularly ataxia and dementia (Marseille Consensus Group, 1990). A large number of rare specific genetic disorders may cause PME, but 5 principal causes are responsible for most cases of PME worldwide: Unverricht-Lundborg disease (ULD), Lafora disease (LD), mitochondrial encephalomyopathies (ME) with the phenotype of myoclonic epilepsy with ragged red fibres (MERRF), and sialidosis and neuronal ceroid lipofuscinoses (NCL). In order to establish a precise diagnosis, the clinical and neurophysiological characteristics of each PME are crucial to guide the clinician to the correct diagnostic algorithm, which may include complex and sometimes invasive procedures (Berkovic *et al.*, 1986; Shahwan *et al.*, 2005). Despite extensive investigations, a number of PMEs remain undiagnosed (Franceschetti *et al.*, 2014); however, these unsolved cases are decreasing over time as a result of recent genetic advances (including whole-genome/exome sequencing), which have led to the discovery of new major causative genes, such as *SCARB2* (Dibbens *et al.*, 2009) and *KCNC1* (Muona *et al.*, 2015). Myoclonus, the main symptom of PMEs, is typically fragmentary and multifocal, involving the musculature of the face and distal limbs. Bilateral massive myoclonic jerks, which tend to involve proximal limb muscles, may also occur, sometimes causing the patient to fall to the ground. Myoclonus may be spontaneous or, more often, induced or exacerbated by a variety of stimuli (such as light, sound, touch, and emotional strain), as well as movement or posture. Action myoclonus, in which muscular jerking is induced by movement or attempts at movement, is the most frequent and disabling form, commonly seen in almost all conditions underlying PME. A mixture of positive and negative myoclonus is the rule in patients with PME, especially LD. The generators for myoclonus in PME remain controversial, but neurophysiological studies suggest that cortical reflex myoclonus is the most common type, while reticular reflex myoclonus is less frequent (Tassinari *et al.*, 1998b). Myoclonus is often difficult to control in these conditions and is usually a significant cause of disability in daily life.

PMEs are characterized by a wide range of epileptic seizures (Michelucci *et al.*, 2002). Generalized tonic-clonic seizures are usually reported at the onset of LD and ULD and may, therefore, suggest the alternative diagnosis of idiopathic generalized epilepsies. They may occur without any warning or after a long build-up of myoclonias. Polygraphic recording may reveal these generalized motor seizures to be tonic-vibratory seizures (as in LD) or true myoclonic or clonic seizures (as in ULD). Absences or focal seizures

(such as occipital seizures in LD) have also been encountered.

Other symptoms, which appear at various times during the course of the illness, include ataxia, cognitive dysfunction (sometimes leading to dementia), pyramidal signs, and a variety of other neurological and extraneurological signs (particularly in mitochondrial diseases).

Overall, the treatment of PMEs remains palliative, since there is no specific treatment for most genetic disorders underlying a PME syndrome (Uthman & Reichl, 2002; Shahwan *et al.*, 2005). Despite a variety of anticonvulsants on the market, effective treatment remains challenging due to the fact that although these drugs may control major convulsive seizures, myoclonus does not really respond to the use of classic antiepileptic drugs (AEDs). Moreover a pathogenetic variety exists among the subtypes of PME, which means that medications which benefit one patient may be less effective in patients with another particular type of PME. Another problem is that clinical trials are difficult to perform due to the small number of patients, the progression of the clinical condition, and the choice of reliable efficacy endpoints. Hence, available data on the efficacy of newer antiepileptic medications in PME are primarily anecdotal or observational, based on individual responses in very small groups of patients. In the present review, we summarize the results of the treatment of PMEs using a number of drugs (including AEDs) and procedures. The issues of designing and performing controlled clinical trials, as well as the emergency treatment, for patients with PMEs are also addressed.

Pharmacological treatment

Traditional AEDs used to treat PMEs are valproate, clonazepam, and phenobarbital (or primidone, a parent compound). These drugs may improve the overall performance of PME patients by decreasing their generalized seizures and, to a lesser extent, their myoclonic jerks. Other drugs which have shown to be effective include piracetam, levetiracetam, topiramate, zonisamide, brivaracetam, and perampamil (at least for LD). The overall therapy of PMEs is summarized in *table 1*.

Valproate

Valproate (at doses ranging from 15 mg/kg to 60 mg/kg) is the treatment of choice for PMEs. It may be used as monotherapy in mild cases or combined with other drugs in more serious cases. Its usefulness was clearly demonstrated by Iivanainen & Himberg (1982) in a prospective study conducted with 26 Finnish adults

Table 1. Treatment of PMEs. (A combination of multiple drugs is commonly needed).

| First-line AEDs | Second-line AEDs | Third-line strategies | Emergency treatment |
|-----------------|------------------|-------------------------|-----------------------------------|
| Valproate | Zonisamide | 5-hydroxy-L-tryptophan | Benzodiazepines i.v. ¹ |
| Clonazepam | Levetiracetam | Lamotrigine | Levetiracetam i.v. |
| | Topiramate | N-acetylcystein | Valproate i.v. |
| | Piracetam | VNS or DBS | Phenytoin i.v. ² |
| | Phenobarbital | Experimental drugs | |
| | | Brivaracetam | |
| | | Perampanel ³ | |

¹ Diazepam, lorazepam, clonazepam, midazolam.

² Phenytoin may be successfully used to treat motor status epilepticus in late-stage PMEs (Miyahara *et al.*, 2009).

³ Used thus far for Lafora disease.

with severe forms of PME, likely to be related to ULD. These patients were severely disabled at the onset of the study (due to stimulus-sensitive myoclonus and ataxia) and were receiving chronic treatment with different combinations of AEDs, including carbamazepine, phenytoin, phenobarbital, primidone, and diazepam. These drugs were discontinued and treatment with valproate and clonazepam was commenced simultaneously, with rapid titration to optimal doses (1,500 to 1,800 mg for valproate and 6 to 10 mg for clonazepam). If the patients still had generalized tonic-clonic seizures, 50 to 100 mg of phenobarbital was added. The patients showed a dramatic improvement, particularly in locomotor ability, which continued in the 19 patients who were followed for 6 years. According to Iivanainen & Himberg (1982), these favourable results contributed to improving the prognosis of 'Baltic' PME and were also due, at least in part, to the discontinuation of phenytoin.

Despite its widespread use for all types of PME, valproate should be given with caution for mitochondrial disorders, due to its inhibitory effect on complex IV (cytochrome c oxidase) activity in the respiratory chain (Lam *et al.*, 1997) and carnitine uptake (Tein *et al.*, 1993). If used, therefore, supplementation with L-carnitine is recommended (Tein *et al.*, 1993).

Clonazepam

The efficacy of clonazepam for the treatment of myoclonus and myoclonic seizures in different clinical contexts is well established (Nanda *et al.*, 1977; Obeid & Panayiotopoulos, 1989; Tassinari *et al.*, 1998a). The role of clonazepam in PMEs was highlighted in the study of Iivanainen & Himberg (1982), as described in the above section. Clonazepam is used as add-on therapy, at doses ranging from 3 to 16 mg/day for adults

and 0.2 mg/kg/day for children. Due to the possible development of tolerance in some patients after one to 6 months of treatment, the dosage may need to be adjusted. Since it is commonly used in combination with other drugs for PMEs, the sedative effects of clonazepam may increase with the concomitant administration of phenobarbital. The effect of other, commonly used benzodiazepines, such as diazepam or clobazam, has not been assessed. However, these drugs may be used alternatively, especially when tolerance to clonazepam has developed.

Phenobarbital

Phenobarbital is a major AED with a wide efficacy spectrum. In PMEs, it may be used as an add-on treatment, at doses ranging from 30 to 200 mg/day (adults) and 3 to 8 mg/kg/day (children), particularly for the control of generalized tonic-clonic seizures (Iivanainen & Himberg, 1982). Particular attention should be paid to the inhibitory effect of valproate on phenobarbital elimination, resulting in phenobarbital accumulation and increased somnolence. Toxic signs may also be precipitated by elevated blood ammonia levels, because the magnitude of valproic acid-induced hyperammonaemia is increased in patients comedicated with phenobarbital (Michelucci *et al.*, 2009). Primidone, which is at least partially metabolized into phenobarbital, has also been used with positive results in patients with myoclonus (Obeso, 1995).

Piracetam

Piracetam, a pyrrolidone derivative with a potent antimyoclonic effect and good tolerability profile, has long been used for the treatment of PMEs. Koskiniemi *et al.* (1998) reported a significant

reduction in myoclonic jerks, and improvement in gait, in a double-blind, placebo-controlled trial in 20 patients with ULD, especially with the highest dose used (24 g/day). In this study, a linear dose-effect relationship of piracetam was established. High piracetam doses (up to 45 g/day) were also used by Genton *et al.* (1999) to obtain a stable and long-lasting improvement in 12 patients with PME.

Other reports have also stressed the positive antimyoclonic effects of 20 g/day for advanced forms of PMEs (Fedi *et al.*, 2001). The major drawback of these higher doses is the number of tablets taken and their bulk, sometimes making adherence to treatment difficult. Piracetam is well tolerated and the side effects (usually consisting of gastrointestinal discomfort) are rare, transitory, and mild even at high doses and can be avoided by slow titration (Ikeda *et al.*, 1996).

Levetiracetam

Levetiracetam, a potent AED with a wide spectrum and a unique mechanism of action (mediated by binding to presynaptic vesicular protein, SV2A), was developed within the class of pyrrolidone derivatives and belongs to the same family of piracetam. For PMEs, levetiracetam has been evaluated in several series and appears to be effective for both myoclonus and generalized seizures (Genton & Gelisse, 2000; Frucht *et al.*, 2001; Kinrions *et al.*, 2003; Crest *et al.*, 2004; Magaouda *et al.*, 2004; Lim & Ahmed, 2005; Mancuso *et al.*, 2006; Papacostas *et al.*, 2007).

Overall, of 23 patients with ULD treated with levetiracetam in open label trials at doses ranging from 1,000 to 4,000 mg, 15 (65 per cent) showed some clinical improvement while 8 (35 per cent) were unchanged. Seven patients (30 per cent) showed a dramatic improvement, which tended to subside, however, with long-term treatment. Levetiracetam is usually well tolerated in this group of patients and does not interact with concomitant drugs used for PMEs.

Topiramate

Topiramate, a sulfamate-substituted monosaccharide molecule, is a widely recognized broad-spectrum AED which is particularly effective for the treatment of refractory focal seizures, as well as primary or secondary generalized seizures. There are scattered reports in the literature showing that topiramate, when used for PMEs, may cause a marked decrease in myoclonus and myoclonic seizure frequency, and an improvement in daily functioning (Uldall & Bucholt, 1999). These effects do not appear to be specific to any given PME, but have been particularly studied in LD (Aykutlu *et al.*, 2005; Demir *et al.*, 2013). In one study, however, topiramate efficacy tended to decrease

over time and the drug was discontinued in 2 out of 5 patients because of a rapid increase in cognitive impairment and vomiting (Aykutlu *et al.*, 2005).

Zonisamide

Zonisamide, a sulfonamide derivative chemically distinct from any of the previously established AEDs, is indicated for the treatment of refractory partial epilepsy, but is also useful for a variety of generalized epilepsies, including epileptic encephalopathies, such as Lennox-Gastaut and West syndrome.

A number of case reports and small studies have suggested that zonisamide may be effective for treating patients with PME. More specifically, almost all patients with ULD who were treated with zonisamide as add-on therapy showed a dramatic reduction in myoclonus and a marked improvement in generalized tonic-clonic seizures and daily functioning, although this effect may subside over time (Henry *et al.*, 1988; Kyllerman & Ben-Menachem, 1998; Yoshimura *et al.*, 2001).

Tassinari *et al.* (1999) investigated the efficacy and tolerability of zonisamide in the treatment of severely disabling action myoclonus. In the 4 patients with PME (2 with ULD and 2 cryptogenic), the authors observed a dramatic improvement, as documented by videopolygraphic recording of the patients before and at various times after the start of zonisamide therapy.

Vossler *et al.* (2008) used add-on zonisamide (up to 6 mg/kg/die) in 30 patients with a variety of PME syndromes refractory to common AEDs. They found a ≥ 50 per cent decrease in myoclonic seizure frequency, measured over a 24-hour period, in 38 per cent of patients. About half of the patients experienced side effects consisting of anorexia, asthenia, and somnolence.

Italiano *et al.* (2011) carried out a pilot, open-label trial of add-on zonisamide (up to 6 mg/kg/day) in 12 patients with EPM1 and studied the effect on myoclonus by means of the Unified Myoclonus Rating Scale, obtained for each subject before and after zonisamide add-on treatment. The authors observed a significant reduction in myoclonus severity following the introduction of zonisamide, associated with a good tolerability profile.

Lamotrigine

Lamotrigine, a triazine derivative developed from a series of folate antagonists, is a useful medication for a wide variety of epilepsies, including partial and generalized conditions. Its plasma levels are markedly increased by valproate co-administration, an interaction which largely explains the significant increase in lamotrigine potency after the addition of valproate.

The clinical experience with the use of lamotrigine for the treatment of myoclonus has given conflicting results. Wallace (1998) reviewed the effect of lamotrigine on different myoclonic syndromes and concluded that lamotrigine could be useful for the control of myoclonus for a variety of conditions, including PMEs. It was noted to be particularly efficacious for the treatment of seizures (of any type) of infantile and juvenile neuronal ceroid lipofuscinosis (Aberg *et al.*, 1999). More recently, lamotrigine was revealed to improve disabling myoclonus in a patient with a mtDNA A3243G mutation (Costello & Sims, 2009).

In juvenile myoclonic epilepsy, however, lamotrigine was reported to exacerbate myoclonus in some cases. A similar effect was reported in some children with Dravet syndrome (Guerrini *et al.*, 1998). Genton *et al.* (2006) retrospectively analyzed the effect of add-on lamotrigine in 5 patients with ULD and observed either an aggravation of myoclonic jerks or a lack of improvement. The authors concluded that lamotrigine is not a sensible treatment option for ULD.

Brivaracetam

Following the discovery of the mechanism of action of levetiracetam, which involves a specific binding site on the presynaptically located synaptic vesicle protein, SV2A, great efforts were made to identify molecules which had structural analogy to levetiracetam and showed high binding constants to SV2A. Brivaracetam, a novel molecule with a 10-fold higher affinity for the SV2A binding site than levetiracetam, was proposed as a drug with high potential efficacy for myoclonus, as suggested by preclinical studies in an established rat model of cardiac arrest post-hypoxic myoclonus (Tai & Truong, 2007) and a phase IIa trial in which its efficacy in the photoparoxysmal response model was analyzed for photosensitive epilepsies (Kasteleijn-Nolst Trenité *et al.*, 2007). In this study, brivaracetam, administered at a daily dose of 80 mg, eliminated the photoparoxysmal responses in 14/18 patients for a period of time exceeding the half-life of the drug.

In 2005, brivaracetam received orphan drug designation by the European Medicines Agency for development in PME and from the US Food and Drug Administration for the treatment of symptomatic myoclonus. Two prospective, multicentre, randomized, double-blind, placebo-controlled, parallel-group studies of brivaracetam as adjunctive treatment in 50 and 56 patients, respectively, with genetically ascertained ULD have recently been completed (N01187, N01236) (Kälviäinen *et al.*, 2009). In the first study, dosages of 50 and 150 mg/day were tested and in the second study, dosages of 5 and 150 mg/day were applied. These patients suffered from

moderate-to-severe action myoclonus and were stratified for concomitant use of levetiracetam or piracetam. Both studies failed to reach the primary goal of reducing the severity of action myoclonus, as measured by the Unified Myoclonus rating scale. However, a favourable trend was observed with brivaracetam at a dose of 50 mg/day in the 12-week maintenance period, and a significant improvement in quality of life (measured by QOLIE-31-P) was found with brivaracetam at daily doses of 50 and 150 mg. Moreover, 87 per cent of patients who completed the studies entered the long-term follow-up study. Overall, brivaracetam was well tolerated in this patient population, as evidenced by the high retention rate and the fact that it did not aggravate seizures or myoclonus. The most frequently reported adverse events were dizziness, headaches, and somnolence.

Although these controlled studies did not appear to support significant efficacy of brivaracetam for the treatment of ULD, a number of factors could have biased the results, including the severity and long duration of the disease, the small scale of the patient population, a high inter- and intra-patient variability, and, perhaps most importantly, the fact that many patients were already on high doses of levetiracetam and/or piracetam, which also act on the SV2A vesicles. A considerable number of patients continued to use brivaracetam after the controlled phase of the trial, for over 6 years, which appears to indicate that this compound has some benefits.

Other drugs and procedures

Perampanel, one of the newer AEDs, is a selective non-competitive antagonist of the AMPA-type glutamate receptors and was recently licensed as adjunctive therapy for the treatment of refractory focal-onset seizures (French *et al.*, 2012; Krauss *et al.*, 2012).

Two reports document its efficacy when used as add-on therapy for the treatment of LD. The first case was a 21-year-old female patient who was administered perampanel at a dose of 8–10 mg, in addition to a regimen that included clonazepam, levetiracetam, piracetam, valproate, zonisamide, a ketogenic diet, and VNS. This therapeutic change resulted in seizure remission for more than 3 months and led to a reduction in the number of epileptiform discharges on EEG (Schorlemmer *et al.*, 2013). The second case was a 15-year-old girl who experienced a dramatic decrease in her seizure frequency, as well as improvement in neurological and cognitive function following initiation of treatment with 10 mg perampanel, administered as monotherapy. Perampanel was therefore proposed as the first potentially efficacious treatment for LD (Dirani *et al.*, 2014). In line with the serotonergic hypothesis of myoclonus, which suggests that serotonergic

hypofunction is involved in the genesis of myoclonus in PMEs and other myoclonic disorders, 5-hydroxy-L-tryptophan, a precursor of serotonin, was used for the treatment of PMEs. In 1980, Koskiniemi *et al.* (1980) performed a double-blind, placebo-controlled, cross-over study with 2 g L-tryptophan in 7 patients with ULD and found a significant improvement in 6 patients, mostly concerning ambulation, myoclonic jerks, and general condition. With long-term L-tryptophan treatment, however, the effect disappeared or was even reversed in 3 of the 7 patients after 3 to 4 weeks. Similar positive short-term results were obtained in a further group of 11 Finnish patients with ULD who received up to 100 mg/kg of L-tryptophan plus carbidopa (in order to prevent metabolism outside of the brain) over a 6-week period (Leino *et al.*, 1981). In contrast, Pranzatelli *et al.* (1995) reported no significant change in myoclonus or ataxia evaluation score in a double-blind, cross-over study with L-5HTP in 8 patients with a variety of PME. Overall, this drug does not appear to have a place in the modern treatment of ULD.

N-acetylcysteine is a sulfhydryl antioxidant that increases cellular glutathione and the activity levels of several antioxidant enzymes. Hurd *et al.* (1996) reported marked beneficial effects on mobility, speech, and seizures in at least 2 of 4 severely affected siblings with ULD treated with N-acetylcysteine in combination with other antioxidants (riboflavin, vitamin E, selenium, and zinc).

Antioxidant vitamins and cofactors, including coenzyme Q₁₀ and L-carnitine, are empirically used to treat mitochondrial disorders (Shahwan *et al.*, 2005; Di Mauro & Mancuso, 2007). Baclofen, a muscle relaxant normally used to treat spasticity by inhibiting both monosynaptic and polysynaptic reflexes at the spinal level, has shown promising results in a few PME cases with prominent spasticity and polymyoclonus (Awaad & Fish, 1995). Ropirinole, a dopamine agonist commonly used to treat Parkinson's disease, was shown to improve myoclonus, writing, and muscular balance in a single patient with ULD (Karvonen *et al.*, 2010). Ethosuximide is active against negative myoclonus, which is often found in PMEs, in association with positive myoclonus.

Alcohol was proven to have some beneficial effects in patients with myoclonus by decreasing myoclonic jerks and improving speech and gait (Genton & Guerrini, 1990). This compound, however, can be used only occasionally to improve the quality of a patient's social life; in contrast, regular use can induce the development of tolerance or even dependence.

A high-fat and low-carbohydrate diet (with a ratio of fat to carbohydrate of 3:1 or 4:1), also known as a ketogenic diet, has been shown to be useful for a variety of severe, drug-resistant epilepsies, including

infantile myoclonic seizures. An Italian study of 5 patients with LD, a condition which causes a specific glycogen metabolism disorder, showed that a ketogenic diet, though well tolerated, was unable to stop disease progression (Cardinali *et al.*, 2006). It is now hypothesized that potential targets of new molecules for LD could involve the inhibition or modulation of glycogen synthesis (Pedersen *et al.*, 2013).

Different *stimulation procedures* have also occasionally been employed for patients with PMEs. Vagal nerve stimulation was implanted in an adult patient with a ULD-type PME, who was followed for 1 year, and the procedure resulted in a marked reduction in seizures (more than 90 per cent) and a significant improvement in cerebellar function, as demonstrated on neurological examination (Smith *et al.*, 2000). Chronic high-frequency deep brain stimulation (DBS) of the subthalamic nucleus has been used in an adult patient with an undiagnosed form of PME who was disabled due to frequent seizures, despite vagal nerve stimulation and a complex antiepileptic regimen (Vesper *et al.*, 2007). After a 12-month follow-up, the seizures were reduced in intensity and frequency by 50 per cent. More recently, 5 adult patients with PME underwent chronic high-frequency DBS (Wille *et al.*, 2011). Electrodes were implanted in the substantia nigra pars reticulata (SNr)/subthalamic nucleus (STN) region in the first patient and additionally in the ventral intermediate nucleus (VIM) bilaterally in the next four cases. After a mean follow-up of 24 months, a reduction in myoclonic seizures was observed in all patients, ranging between 30 and 100 per cent, as quantified by a standardized video protocol. All patients reported clinically relevant improvements of various capabilities, such as free standing, walking, and improved fine motor skills. The best clinical effects were seen with SNr/STN DBS in all patients.

Drugs and circumstances to avoid

Interestingly, rather than being beneficial, some AEDs have the potential to exacerbate myoclonic seizures and should be used with caution in patients with PME (table 2). More specifically, sodium channel blockers (carbamazepine, oxcarbazepine, and phenytoin) and GABAergic drugs (vigabatrin and tiagabine), as well as gabapentin and pregabalin, should, in general, be avoided as they may aggravate myoclonus and myoclonic seizures (Medina *et al.*, 2005).

Phenytoin has also been found to aggravate neurological symptoms and cerebellar ataxia in ULD and its widespread use in the past has been proposed as an explanation for the poor prognosis of ULD described in the early series reports (Iivanainen & Himberg, 1982; Elridge *et al.*, 1983).

Table 2. Antiepileptic drugs and effects on myoclonus in PME.

| Antimyoclonic | Potentially aggravating | To be used with caution | Not documented |
|-------------------------|-------------------------|-------------------------|-----------------|
| Valproate | Phenytoin | Lamotrigine | Lacosamide |
| Clonazepam | Carbamazepine | Valproate for MERRF | Felbamate |
| Phenobarbital/Primidone | Oxcarbazepine | | Rufinamide |
| Piracetam | Vigabatrin | | Ethosuximide |
| Levetiracetam | Gabapentin | | Eslicarbazepine |
| Topiramate | Pregabalin | | |
| Zonisamide | Tiagabine | | |

Emergency treatment of PMEs

In situations where myoclonic jerks are exacerbated, leading to a series of jerks or status myoclonicus, loud noises and bright lights should be avoided and the patient should be treated in a quiet room, as calmly as possible. Emergency treatment includes the intravenous use of benzodiazepines (diazepam, lorazepam, clonazepam, and midazolam), valproate, and levetiracetam (Fernandez-Baca Vaca *et al.*, 2012). Phenytoin, although usually contraindicated for PMEs, has proven to be useful in selected cases of refractory status epilepticus, particularly when this occurs in the late stages of a variety of PMEs or in the presence of focal status (Riguzzi *et al.*, 1997; Kälviäinen *et al.*, 2008; Miyahara *et al.*, 2009).

Conclusions

The treatment of PME disorders essentially continues to involve the management of seizures and myoclonus, together with palliative, supportive, and rehabilitative measures. The treatment of myoclonus and seizures in PME can prove to be difficult and both tend to be refractory to conventional medications. Available data on the efficacy of drugs are primarily anecdotal or observational based on small groups of patients. It is difficult to conduct controlled clinical trials in these patients because the incidence of these disorders is exceedingly rare; however, collaborative trials involving many specialized centres could be designed to bring together a sufficient number of patients with a genetically verified diagnosis. Following the availability of brivaracetam, a potentially effective antimyoclonic agent, 2 multicentre, randomized, placebo-controlled studies on genetically verified ULD have been performed, but the effect of this drug on action myoclonus was statistically not significant. However, a favourable trend was observed with the 50-mg dose and it was

argued that various factors could have negatively influenced the results.

Traditional AEDs used for the treatment of PMEs are valproate, clonazepam, phenobarbital, or primidone. Newer drugs which have been shown to be effective include piracetam, levetiracetam, topiramate, zonisamide, and, possibly, perampanel for LD. Care must be taken to avoid antiepileptic medications that clearly worsen myoclonus, such as vigabatrin, carbamazepine, phenytoin, and gabapentin. Lamotrigine has an unpredictable effect on myoclonus and must be used with caution.

Although recent advances in molecular genetics have led to the identification of several genes, mutations, and proteins involved in the pathogenesis of PME disorders, therapy targeting the underlying aetiology remains in the experimental phase and results, to date, have not been encouraging. It is expected, however, that future treatments with gene therapy and enzyme replacement, or the identification of drugs that interact with new targets and mechanisms, may help to modify and improve the course of these progressive disorders. □

Disclosures.

None of the authors have any conflict of interest to disclose.

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Post-modern therapeutic approaches for progressive myoclonus epilepsy

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ABSTRACT – While the PME are arguably the severest epilepsies and neurological disorders, the vast majority are monogenic. Additionally, many affect straightforward biochemical pathways. Finally, by definition, they occur in previously healthy and well-developed brains. As such, their therapies should be easier than in complex, albeit often less severe, neurological developmental disorders where the complex, poorly understood, and extremely difficult-to-correct, neural network of the brain is affected. This last article reviews the latest cutting edge technologies in monogenic disease therapy, with some examples provided applicable to a number of disease. It aims to give a sense of where we are and how much closer we are, to the goal of making an actual organic difference.

Key words: Cas9, CRISPR, gene therapy, AAV9, small molecule, progressive myoclonus epilepsies

By and large, progressive myoclonus epilepsies (PMEs) described in this supplement (Minassian *et al.*, 2016) are devastating and can be lethal. However, most are monogenic, rendering them much more amenable to future therapies relative to other complex diseases, for two reasons; firstly, the pathophysiology is easier to understand, and secondly, therapy may be targeted to a single gene, function, or pathway. In this article, some of the cutting-edge therapeutic modalities applicable to monogenic brain diseases is reviewed. In each case, one example is provided, simply as an illustration. It is not possible to provide a comprehensive review since even as this state-of-the-art supplement is being published, discoveries are being made regarding new molecular targeting and delivery systems.

Small molecules

We are accustomed to therapy in the form of a pill ingested daily, the active compound of which travels through the gut and blood, crosses the blood-brain barrier (BBB), and enters cells to impart therapeutic change. The size of small molecules is their abiding therapeutic advantage for PMEs and other brain diseases, and size matters regarding passage through the BBB, but of course, small size is not all it takes. For example, the presence of a monoester (phosphate) bound to a small molecule prevents it from not only crossing the BBB, but even cell membranes.

However, a small molecule will not replace a gene function, but could, as a chaperone, stabilize a gene product which is present but simply

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misfolded and thus non-functional. In other words, for diseases where at least one of the mutations is a missense mutation that deconvolutes or otherwise destabilizes the tertiary structure of a disease gene product, a small chaperone molecule may stabilize the structure to a sufficient extent to render it functional, effectively above a clinical threshold.

Tay-Sachs disease is a neurodegenerative disease due to mutations in the hexosaminidase A (*HEXA*) gene. In its classic form, it is an infantile disease that is fatal by age 2 or 3. However, some “mild” mutations are associated with a milder neurodegenerative course. The FDA-approved antimalarial BBB-penetrant agent, pyrimethamine, was shown to stabilize certain *HEXA* missense mutants in fibroblasts of patients with late-onset Tay-Sachs disease, and raise enzymatic activity threefold (Maegawa *et al.*, 2007). This led to a pilot clinical study in 4 patients in whom *HEXA* activity rose 2.24 fold. The rise was unfortunately not sustained and the study was clinically unsuccessful (Osher *et al.*, 2015), but nonetheless this was a start for an otherwise intractable disease.

While small molecules cannot replace gene functions, they can very well affect other parts of a disease pathway, whether metabolic or not, or a second messenger. Often, the absence of a gene causes the activity of a downstream protein to be up or down-regulated, which when corrected through interference by a small molecule, even partially, may revert disease pathogenesis and be therapeutic. mTOR pathway interference with rapamycin and its analogues is a case in point. Tuberous sclerosis and several other epilepsies are caused by genetic disruptions of this pathway, leading to focal or more widespread dysplasias and tumours (usually benign), which drive epilepsy. This therapy, which partially regulates the pathway and presents an entirely novel, more direct mechanism, therefore provides intrinsic support as treatment for these epilepsies (Ricos *et al.*, 2015).

Identifying a small molecule drug commonly requires establishment of *in vitro* assays; *in vitro* (biochemical) and/or cell culture assays. These assays are based on the associated molecular or biochemical disease pathway(s) which are investigated through standard gene function-based research. The assays are used to screen ever-expanding libraries of small molecules in order to identify, by chance, one or more that may affect the assay in the desired way. Huge amounts of work follow in translating ‘hits’ to something useful. This involves establishing safety and efficacy in animal models, mouse and larger animals, BBB passage, and other pharmacological aspects. Usually, the initial ‘hit’ is less than optimal, and a prolonged process of medicinal chemistry towards optimization follows, until eventually a drug can be ready for testing in human subjects. This type of work is long and expensive. However, the

infrastructure is increasingly in place in many universities, and even more so in pharmaceutical companies. The latter appear to be moving very much in the direction of developing drugs for orphan diseases. These are usually so devastating that families and societies will incur the costs of these drugs which will always be priced high, in part, to recover the large investment of development.

While “chance” through high-throughput screening is the common and strongest approach to identify interfering molecules, the vast expansion of crystal structures of proteins available as well as the bioinformatics tools to study these structures and their functions, are allowing intelligent drug design, e.g. *in silico* development of chemicals that might fit an enzyme’s active site. This is presently in its infancy, but clearly holds much promise. One of the biggest problems with small molecules is their usual incomplete selectivity, i.e. their interference with other targets, leading to side effects. This too could partially be managed in the future with complete understanding of protein structures and even stronger informatics tools. Clearly, oral drugs are the most desired therapeutic approach due to non-invasiveness, but drugs are neither permanent solutions, nor, for the moment, easy to tailor to ensure a high degree of target specificity. As such, other approaches are needed, as discussed below. Some are invasive, but this is mitigated by the huge need for them, given the devastating nature of the PMEs.

Targeting RNA

As well as targeting proteins, it is possible to target messenger RNA (mRNA). The same general principles hold, in that it is generally much easier to downregulate mRNA than increase it, and the same issues regarding BBB passage exist. There are a number of anti-RNA approaches, the most advanced of which is the use of antisense oligonucleotides (ASOs). These oligonucleotides are designed to interfere with an mRNA and downregulate it. One of these is already on the market as a drug for familial hyperlipidaemia (Phillips *et al.*, 2015).

There are currently many ASOs in clinical trials for various neurological diseases (e.g. Huntington disease and spinocerebellar ataxia [Keiser *et al.*, 2015]), but all share the same problem in that they do not cross the BBB and require intra-cerebrospinal fluid administration. This problem is mitigated by specific chemical properties that allow their longevity, and the patient does not require an injection more than once every two months. This is clearly still far from ideal, but for the moment, is the best available treatment. Interestingly,

it has been reported that ASOs can actually activate a function which has been lost.

The most common cause of spinal muscular atrophy (SMA) is loss-of-function mutation of the *SMN1* gene. It so happens that evolution has led to the *SMN1* gene being located adjacent to the *SMN2* gene. In SMA, *SMN2* is also generally inactive due to a sequence that leads to skipping over exon 7. ASOs have been designed to interfere with this skipping of exon 7 splicing, and thus lead to inclusion of this exon, thus rendering *SMN2* functional. This “drug” is presently in an advanced clinical trial (Faravelli *et al.*, 2015).

Targeting DNA

Bacteria, of course have an immune system; how else would they survive the onslaught of viruses over millions of years? This immunity consists of generating guide RNAs that specifically target enzymes (e.g. Cas9) to viral genomes. The Cas9 then nicks and inactivates the viral DNA.

Impressive recent studies have addressed the following questions. Could Cas9 cut human DNA? The answer is yes. Could Cas9 be directed by specific guide RNA sequences to specific sites in the human genome to cut specific genes? Yes. Would the human genome be able to mend itself after being cut? Yes. Does this correction maintain the coding frame of a target gene? No. *In toto*, this CRISPR (which refers to the guide RNA)/Cas9 system can specifically target a desired gene and break it, causing the cell's DNA repair system to mend the chromosomal break, however, usually not in-frame, *i.e.* the targeted gene is knocked out, but the chromosome is otherwise untouched (Ran *et al.*, 2015). The major advantage of this genome editing approach is that it is permanent. Presently, the system only knocks out genes. If a disease pathway is identified in which a protein function in the pathway is increased, and its elimination would correct pathogenesis, then CRISPR/Cas9 could be utilized to remove this protein and treat the disease. The greatest therapeutic promise of the system, however, is in future possibilities. If the system can be targeted to a mutant gene, specifically to the mutation site, such that the mutation is removed and replaced with normal sequence, then the disease would be corrected at the source.

There are countless difficulties to overcome before the CRISPR system delivers. Firstly, Cas9 and guide RNA have to be delivered to the brain, no small feat as this is a large protein. It would be possible to deliver CRISPR/Cas9 through viral gene therapy, but the gene for the commonly used Cas9, from *Strep. viridans*, is too large to fit in the AAV viral vectors that have the greatest promise for gene therapy. This problem, though, appears to have been solved with the

identification of a smaller, but equally effective, Cas9 gene from *Staph. aureus*, which does fit in AAV particles (Ran *et al.*, 2015).

Another problem is the need to deliver to large numbers of neurons, *i.e.* ‘edit’ most neurons, which is a limitation, though perhaps not serious, of present viral vectors (see next section). Then there is the likely immunogenicity of Cas9 delivered to humans. However, Cas9 will not need to be delivered many times, because its effect would be permanent. As such, the patient could be immunosuppressed during the treatment.

Another problem is the risks associated with off-target nicking. While bioinformatics is used to design specific CRISPRs against specific sequences, the genome is vast enough that there is at least the risk of off-target cutting, with potential for ‘side-effects’, including oncogenesis.

Off-target effects, however, from most recent data, appear to be not nearly as concerning as initially thought (Iyer *et al.*, 2015). In addition, there is, of course, the looming issue that gene deletion as a therapeutic approach would be useful in only very few situations. What is desired is mutation correction, and the CRISPR system is not quite there yet. There have been successes, but correction of gene mutation remains inefficient and much work is required to optimize it. However, if this system proves to be successful, it would allow, as mentioned, correction of the root of the problem in essentially all the diseases described in this book. An important step forward has been taken recently in this endeavour. Yet another bacterial DNA nickase has been identified, which, unlike Cas9, cleaves DNA via a staggered cut, leading to overhanging ends, which should make the replacement of a deleted segment by a corrected sequence much easier (Zetsche *et al.*, 2015).

Gene therapy

Most of the conditions described in this supplement are autosomal recessive diseases caused by a loss of function of single genes. As such, they can potentially be remedied by replacing the gene function. Gene replacement therapy stalled for a long time in the 1990s due to premature use of immunogenic viruses for gene delivery with subsequent harm to patients. However, gradual careful work has brought to the fore the adeno-associated viruses, in particular AAV9 and AAVrh10, which have superb properties. These viruses are normally present in the human brain, implying natural BBB passage capability and low immunogenicity. However, low immunogenicity does not mean no immunogenicity. The high load of virus that needs to be given generates immune reactions in animal models, although, presumably, this problem can be

averted by immunosuppression during treatment. AAV viruses can package genes of average size, which means they are useful, potentially, for most diseases. The DNA vectors are not silenced, leading to persistent expression. Their DNA does not integrate into the human genome, *i.e.* there is low to no risk of interfering with genomic DNA, including tumour suppressor genes. Finally, AAV9 is capable of delivering its cargo to a large proportion of cells, in particular, when injected in the CSF (Meyer *et al.*, 2015).

Gene therapy is indeed in resurgence and it is hoped the technical difficulties will continue to be overcome, to allow gene replacement, and, as discussed above, gene correction. In fact, clinical trials are ongoing for multiple diseases, including some neuronal ceroid lipofuscinoses reviewed in this supplement, as well as SMA, as mentioned above. While the ASO approach for SMA aims at activating *SMN2*, the AAV-delivered gene therapy approach aims to replace under-expressed *SMN1*. Over a dozen babies with this fatal disease have been injected so far with virus carrying *SMN1*, and they are doing well (B. Kaspar, *personal communication*); publication of this major potential therapeutic success by the group which has been working on it for many years is anticipated (Mandel, Kaspar and colleagues).

While gene therapy mostly utilizes viruses, it must be noted that there are other approaches. These include packaging the gene of interest in liposomes, which merge with membranes and can potentially make their way through the BBB to neurons. Immunoliposomes are another development on this concept. Here, the liposome membrane contains an antibody that allows it to bind to membrane receptors that are then internalized, thus supporting transduction into the brain.

Protein replacement therapy

In this last section, we touch on the possibility of directly replacing the missing protein, instead of its gene. Multiple approaches are used. This relies, as for immunoliposomes, on antibodies against the transferrin receptor or other proteins. The antibody is covalently bound to the protein to be delivered, and once it interacts with its target, it and the protein are internalized into vesicles, which then merge with lysosomes. As such, this type of therapy would be useful for lysosomal diseases, which make up a large part of PME.

Another exciting possibility is the use of diphtheria toxin. This toxin binds a receptor on the endothelial cell membrane and is internalized into a vesicle into the endothelial cells that make up the BBB. This vesicle translocates to the other side of the cells and

releases the toxin into the CSF space. The toxin then binds to its receptor at the surface of neurons, is again internalized into a vesicle, but this time it is injected out of the vesicle into the neuronal cytoplasm, where it blocks protein synthesis and kills the neuron. It was shown that the toxicity of this toxin can be completely eliminated by a point mutation, whilst retaining its translocation properties. Finally, it was shown that a cargo protein, almost of any size, can be attached to this toxin and co-delivered to cells (so far in cell culture experiments only) with maintained function of the delivered protein (Auger *et al.*, 2015).

There are other post-genetic era therapeutic approaches in the works, and indeed a lot of excitement that the expression of certain genes will follow Mendelian genetics. For example, for Lafora disease, one of the prototypical PMEs described in an earlier chapter of this supplement (Turnbull *et al.*, 2016), progress in understanding the disease has allowed us to 'cure' Lafora mice simply by reducing the amount of glycogen synthase (GS), the enzyme that gives rise to the neurotoxic Lafora bodies that underlie the disease. As such, we are presently applying almost all the methods above to attempt to find a therapy for this horrendous condition. This involves screening for small molecule inhibitors of GS, using ASOs against GS and its activator protein PTG, experimenting with CRISPRs against the latter two proteins, using AAV9 to replace the two respective disease-causing genes, and using inactivated diphtheria toxin-mediated delivery of amylase to mouse brain. Amylase is the only enzyme known to digest Lafora bodies. We have already shown that we are able to deliver amylase to cells in culture in this fashion and that amylase remains functional after crossing the cell membrane (Auger *et al.*, 2015). This disease, which we are particularly familiar with, exemplifies the immense possibilities ahead for all our patients with any form of PME. □

Disclosures.

The author has no conflict of interest to disclose.

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